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(57) Abstract

The invention provides isolated nucleic acids molecules, designated hVR-1, hVR-2, and rVR-2 nucleic acid molecules, which encode novel members of the Capsaicin/Vanilloid receptor family. The invention also provides antisense nucleic acid molecules, recombinant expression vectors containing hVR-1, hVR-2, and rVR-2 nucleic acid molecules, host cells into which the expression vectors have been introduced, and nonhuman transgenic animals in which an hVR-1, hVR-2, and rVR-2 gene has been introduced or disrupted. The invention still further provides isolated hVR-1, hVR-2, and rVR-2 proteins, fusion proteins, antigenic peptides and anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies. Diagnostic methods utilizing compositions of the invention are also provided.

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NOVEL MEMBERS OF THE CAPSAICIN/VANILLOID RECEPTOR FAMILY OF PROTEINS AND USES THEREOF

Background of the Invention

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Pain is initiated when the peripheral terminals of a subgroup of sensory neurons are activated by noxious chemical, mechanical or thermal stimuli. These neurons, called nociceptors, transmit information regarding tissue damage to pain-processing centres in the spinal chord and brain (Fields, H.L. Pain, McGraw-Hill, New York, 1987). Nociceptors are characterized in part, by their sensitivity to capsaicin, a vanilloidcontaining compound, and a natural product of capsicum peppers that is the active ingredient of many "hot" and spicy foods. In mammals, exposure of nociceptor terminals to capsaicin leads initially to excitation of the neuron and the consequent perception of pain and local release of inflammatory mediators. With prolonged exposure, nociceptor terminals become insensitive to capsaicin, as well as to other noxious stimuli (Szolcsanyi, J. in Capsaicin in the Study of Pain (ed. Wood, J.) 1-26 (Academic, London, 1993). This latter phenomenon of nociceptor desensitization underlies the seemingly paradoxical use of capsaicin as an analgesic agent in the treatment of painful disorders ranging from viral and diabetic neuropathies to rheumatoid arthritis (Campbell, E. in Capsaicin and the Studyof Pain (ed. Wood, J.) 255-272 (Academic, London, 1993); Szallasi, A. et al. (1996) Pain 68, 195-208). Some of this decreased sensitivity to noxious stimuli may result from reversible changes in the nociceptor, but the long-term loss of responsiveness can be explained by death of the nociceptor or destruction of its peripheral terminals following exposure to capsaicin (Jancso, G. et al. (1977) Nature 270, 741-743).

The cellular specificity of capsaicin action and its ability to evoke the sensation of burning pain have led to speculation that the target of capsaicin action plays an important physiological role in the detection of painful stimuli. Indeed, capsaicin may elicit the perception of pain by mimicking the actions of a physiological stimulus or an endogenous ligand produced during tissue injury (James, I.F., Kinkina, N.N. & Wood, J.N. in *Capsaicin in the Study of Pain* (ed. Wood, J.N.) 83-104 (Academic, London, 1993).

-2-

Caterina M.J. et al. have recently determined the molecular basis underlying this phenomenon by characterizing a functional cDNA that encodes a vanilloid receptor (VR-1) in rat sensory ganglia (Caterina M. J. et al., (1997) Nature 389:816-824). VR-1 is a vanilloid-gated, nonselective cation channel that resembles members of the transient receptor potential (TRP) channel family, first identified as components of the *Drosophila* phototransduction pathway (Montell et al. (1989) Neuron 2:1313-1323).

Table of Contents

	A.	Sumr	nary of	the inv	vention	-3-	
10	B.	Brief	ief Description of the Drawings			-8-	
	C.	Detailed Description of the Invention					
		I.	Isolated Nucleic Acid Molecules				
		II.	Isolated hVR-1, hVR-2, and rVR-2 Proteins and				
			Anti- hVR-1,Anti-hVR-2, and Anti-rVR-2 Antibodies				
15		III.	Recombinant Expression Vectors and Host Cells				
		IV.	Pharmaceutical Compositions				
		V.	Uses and Methods of the Invention				
			A.	Scre	ening Assays	-60-	
			B.	Dete	ection Assays	-67-	
20				1.	Chromosome Mapping	-67-	
				2.	Tissue Typing	-70-	
				3.	Use of Partial hVR-1, hVR-2, and rVR-2		
					Sequences in Forensic Biology	-71-	
			C.	Pred	ictive Medicine	-72-	
25				1.	Diagnostic Assays	-73-	
				2.	Prognostic Assays	-75-	
				3	Monitoring of Effects During Clinical Trials	-81-	

	D. Metho	Methods of Treatment		
	1.	Prophylactic Methods	-84-	
	2.	Therapeutic Methods	- 84-	
	3.	Pharmacogenomics	-85-	
5	D. Examples		-88-	

Summary of the Invention

The present invention is based, at least in part, on the discovery of novel members of the Capsaicin/Vanilloid family of receptors. Described herein is the isolation of the human orthologue of rat VR-1 (rVR-1), referred to herein as hVR-1, as well as another previously unknown member of the VR family of receptors, referred herein as VR-2, and specifically as human VR-2 (hVR-2, including an alternate form which contains a deletion) and rat VR-2 (rVR-2) nucleic acid and protein molecules. The hVR-1, hVR-2, and rVR-2 molecules of the present invention are useful as targets for developing modulating agents to regulate a variety of cellular processes, *e.g.*, cellular processes involved in pain. Accordingly, in one aspect, this invention provides isolated nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins and fragments thereof, as well as nucleic acid fragments suitable as primers or hybridization probes for the detection of hVR-1, hVR-2, and rVR-2-encoding nucleic acids.

In one embodiment, an hVR-1, hVR-2, or rVR-2 nucleic acid molecule of the invention is at least 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical to the nucleotide sequence (*e.g.*, to the entire length of the nucleotide sequence) shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof.

In another embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof. In another embodiment, the nucleic acid molecule includes at least 10, 15, 20, or more contiguous nucleotides of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

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In another embodiment, an hVR-1, hVR-2, and rVR-2 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11. In one embodiment, an hVR-1, hVR-2, and rVR-2 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more identical to the entire length of the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.

Another embodiment of the invention features nucleic acid molecules, preferably hVR-1, hVR-2, and rVR-2 nucleic acid molecules, which specifically detect hVR-1, hVR-2, and rVR-2 nucleic acid molecules relative to nucleic acid molecules encoding non-hVR-1, non-hVR-2, and non-hVR-2 proteins. For example, in one embodiment, such a nucleic acid molecule is at least 100-150, 1150-200, 200-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, 550-600, 600-700, 700-800, 800-900, 900-1000, 1088, or more nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In preferred embodiments, the nucleic acid molecules are at least 15 (e.g., contiguous) nucleotides in length and hybridize under stringent conditions to nucleotides 1-17, 3696-3863, or 3901-3909 of SEQ ID NO:1. In other preferred embodiments, the nucleic acid molecules comprise nucleotides 1-17, 3696-3863, or 3901-3909 of SEQ ID NO:1. In yet other preferred embodiments, the nucleic acid molecules consist of nucleotides 1-17, 3696-3863, or 3901-3909 of SEQ ID NO:1. In preferred embodiments, the nucleic acid molecules are at least 15 (e.g., contiguous) nucleotides in length and hybridize under stringent conditions to nucleotides 1944-2003 of SEQ ID NO:4. In other preferred embodiments, the nucleic acid molecules comprise nucleotides 1944-2003 of SEQ ID NO:4. In yet other preferred embodiments, the nucleic acid molecules consist of nucleotides 1944-2003 of SEQ ID NO:4.

In other embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions and is encoded by the same locus as hVR-1, hVR-2 or rVR-2.

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Another embodiment of the invention provides a nucleic acid molecule that encodes a naturally occurring orthologue of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, *e.g.*, the coding strand of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule.

Since the hVR2 (the alternate form) and rVR2 sequences represent fragments of the entire coding regions of these genes, another embodiment of the invention provides the complete gene sequences. A skilled artisan can readily isolate such molecule using the sequences disclosed herein.

Another aspect of the invention provides a vector comprising an hVR-1, an hVR-2, or a rVR-2 nucleic acid molecule. In certain embodiments, the vector is a recombinant expression vector. In another embodiment, the invention provides a host cell containing a vector of the invention. In yet another embodiment, the invention provides a host cell containing a nucleic acid molecule of the invention. The invention also provides a method for producing a protein, preferably an hVR-1, hVR-2, and rVR-2 protein, by culturing in a suitable medium, a host cell, e.g., a mammalian host cell such as a non-human mammalian cell, of the invention containing a recombinant expression vector, such that the protein is produced.

Another aspect of this invention features isolated or recombinant hVR-1, hVR-2, and rVR-2 proteins and polypeptides. In one embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain. In another embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain and at least one proline rich domain. In yet another embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain. In yet another embodiment, the protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one proline rich domain, and at least one ankyrin repeat domain and has

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an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11. In another embodiment, the protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain and plays a role in the development and regulation of pain. In yet another embodiment, the protein, preferably an hVR-1, hVR-2, and rVR-2 protein. includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

In another embodiment, the invention features fragments of the protein having the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15, 30, 40, 50, 60, 70, 80, 90, or 100 amino acids (e.g., contiguous amino acids).

In another embodiment, the invention features an isolated protein, preferably an hVR-1, hVR-2, and rVR-2 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence at least about 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous to a nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof. This invention further features an isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule consisting of the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof.

The proteins of the present invention or portions thereof, *e.g.*, biologically active portions thereof, can be operatively linked to a non-hVR-1, non-hVR-2, or non-rVR-2 polypeptide (*e.g.*, heterologous amino acid sequences) to form fusion proteins. The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind proteins of the invention, preferably hVR-1, hVR-2, and rVR-2 proteins. In addition, the hVR-1, hVR-2, and rVR-2 proteins or biologically active

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portions thereof can be incorporated into pharmaceutical compositions, which optionally include pharmaceutically acceptable carriers.

In another aspect, the present invention provides a method for detecting the presence of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide in a biological sample by contacting the biological sample with an agent capable of detecting an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide such that the presence of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide is detected in the biological sample.

In another aspect, the present invention provides a method for detecting the presence of hVR-1, hVR-2, and rVR-2 activity in a biological sample by contacting the biological sample with an agent capable of detecting an indicator of hVR-1, hVR-2, and rVR-2 activity such that the presence of hVR-1, hVR-2, and rVR-2 activity is detected in the biological sample.

In another aspect, the invention provides a method for modulating hVR-1, hVR-2. and rVR-2 activity comprising contacting a cell capable of expressing hVR-1, hVR-2, and rVR-2 with an agent that modulates hVR-1, hVR-2, and rVR-2 activity such that hVR-1, hVR-2, and rVR-2 activity in the cell is modulated. In one embodiment, the agent inhibits hVR-1, hVR-2, and rVR-2 activity. In another embodiment, the agent stimulates hVR-1, hVR-2, and rVR-2 activity. In one embodiment, the agent is an antibody that specifically binds to an hVR-1, hVR-2, and rVR-2 protein. In another embodiment, the agent modulates expression of hVR-1, hVR-2, and rVR-2 by modulating transcription of an hVR-1, hVR-2, and rVR-2 gene or translation of an hVR-1, hVR-2, and rVR-2 mRNA. In yet another embodiment, the agent is a nucleic acid molecule having a nucleotide sequence that is antisense to the coding strand of an hVR-1, hVR-2, and rVR-2 mRNA or an hVR-1, hVR-2, and rVR-2 gene.

In one embodiment, the methods of the present invention are used to treat a subject having a disorder characterized by aberrant hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity by administering an agent which is an hVR-1, hVR-2, and rVR-2 modulator to the subject. In one embodiment, the hVR-1, hVR-2, and rVR-2 modulator is an hVR-1, hVR-2, and rVR-2 protein. In another embodiment the hVR-1, hVR-2, and rVR-2 modulator is an hVR-1, hVR-2, and rVR-2 nucleic acid molecule. In

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yet another embodiment, the hVR-1, hVR-2, and rVR-2 modulator is a peptide, peptidomimetic, or other small molecule. In a further embodiment, the disorder characterized by aberrant hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression is a pain disorder, e.g., hyperalgesia.

The present invention also provides a diagnostic assay for identifying the presence or absence of a genetic alteration characterized by at least one of (i) aberrant modification or mutation of a gene encoding an hVR-1, hVR-2, and rVR-2 protein; (ii) mis-regulation of the gene; and (iii) aberrant post-translational modification of an hVR-1, hVR-2, and rVR-2 protein, wherein a wild-type form of the gene encodes a protein with an hVR-1, hVR-2, and rVR-2 activity (as described herein).

In another aspect the invention provides a method for identifying a compound that binds to or modulates the activity of an hVR-1, hVR-2, and rVR-2 protein, by providing an indicator composition comprising an hVR-1, hVR-2, and rVR-2 protein having hVR-1, hVR-2, and rVR-2 activity, contacting the indicator composition with a test compound, and determining the effect of the test compound on hVR-1, hVR-2, and rVR-2 activity in the indicator composition to identify a compound that modulates the activity of an hVR-1, hVR-2, and rVR-2 protein.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

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Brief Description of the Drawings

Figure 1 depicts the full length cDNA sequence and predicted amino acid sequence of human VR-1 (hVR-1). The nucleotide sequence corresponds to nucleic acids 1 to 3909 of SEQ ID NO:1. The amino acid sequence corresponds to amino acids 1 to 839 of SEQ ID NO:2. The coding region without the 5' and 3' untranslated regions of the human VR-1 (hVR-1) gene is shown in SEQ ID NO:3.

Figure 2 depicts the full length cDNA sequence and predicted amino acid sequence of human VR-2 (hVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 2809 of SEQ ID NO:4. The amino acid sequence corresponds to amino acids 1 to 764 of SEQ ID NO:5. The coding region without the 5' and 3' untranslated regions of the human VR-2 (hVR-2) gene is shown in SEQ ID NO:6.

Figure 3 depicts the partial cDNA sequence and partial predicted amino acid sequence of an alternate form of human VR-2 (hVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 1489 of SEQ ID NO:7. The amino acid sequence corresponds to amino acids 1 to 436 of SEQ ID NO:8. The coding region without the 5' and 3' untranslated regions of the alternate form of human VR-2 (hVR-2) gene is shown in SEQ ID NO:9.

Figure 4 depicts the partial cDNA sequence and partial predicted amino acid sequence of rat VR-2 (rVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 1794 of SEQ ID NO:10. The amino acid sequence corresponds to amino acids 1 to 554 of SEQ ID NO:11. The coding region without the 5' and 3' untranslated regions of the rat VR-2 (rVR-2) gene is shown in SEQ ID NO:12.

Figure 5 depicts an alignment of the hVR-1 protein (SEQ ID NO:2) with the human VR-2 protein (SEQ ID NO:5) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

Figure 6 depicts an alignment of the hVR-1 nucleotide sequence (SEQ ID NO:1) with the human VR-2 nucleotide sequence (SEQ ID NO:4) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3.

Figure 7 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the rat VR-2 protein (SEQ ID NO:11) using the CLUSTAL W (1.74) multiple sequence alignment program.

Figure 8 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the rat VR-2 protein (SEQ ID NO:11) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

Figure 9 depicts an alignment of the hVR-1 nucleotide sequence (SEQ ID NO:1) with the rat VR-1 nucleotide sequence (Accession Number:AF029310) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3.

Figure 10 depicts an alignment of the hVR-1 protein (SEQ ID NO:2) with the rat

VR-1 protein (Accession Number:AF029310) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

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Figure 11 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the human VR-2 protein (alternate form) (SEQ ID NO:8) using the CLUSTAL W (1.74) multiple sequence alignment program.

Figure 12 depicts a structural, hydrophobicity, and antigenicity analysis of the hVR-1 protein.

Figure 13 depicts the results of a search using the amino acid sequence of the hVR-1 protein against the HMM database.

Figure 14 depicts a structural, hydrophobicity, and antigenicity analysis of the hVR-2 protein.

10 Figure 15 depicts the results of a search using the amino acid sequence of the hVR-2 protein against the HMM database.

Figure 16 depicts the predicted full length amino acid sequence of the human VR-2 protein (alternate form) (SEQ ID NO:20).

Figure 17 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the predicted full length human VR-2 protein (alternate form) (SEQ ID NO:20) using the CLUSTAL W (1.74) multiple sequence alignment program.

Detailed Description of the Invention

The present invention is based, at least in part, on the discovery of nucleic acid 20 and amino acid molecules which are novel members of the Capsaicin/Vanilloid family of receptors. Described herein is the isolation of the human orthologue of rat VR-1 (rVR-1), referred to herein as hVR-1, as well as another previously unknown member of the VR family of receptors, referred herein as VR-2, and specifically as human VR-2 (hVR-2) and rat VR-2 (rVR-2) nucleic acid and protein molecules. The hVR-1, hVR-2, and rVR-2 molecules were identified based on their sequence similarity to the known rat 25 vanilloid receptor (VR-1). VR-1 is a vanilloid gated, non-selective cation channel which resembles members of the transient receptor potential (TRP) ion channel family (described in Montell et al. (1989) Neuron 2:1313-1323) that mediate the influx of extracellular calcium in response to depletion of intracellular calcium stores. The rat VR-1 cDNA contains an open reading frame of 2514 nucleotides that encodes a protein 30 of 838 amino acids. Hydrophilicity analysis has indicated that rat VR-1 contains six

- 11 -

transmembrane domains (predicted to be mostly α-helices) with an additional short hydrophobic stretch between transmembrane regions 5 and 6. The amino terminal hydrophilic segment contains a relatively proline rich region followed by three ankyrin repeat domains. The rat VR-1 is expressed in small diameter neurons within sensory ganglia. The present hVR-1 sequence is the human orthologue of rVR-1. As described in further detail *infra*, the human VR-1 is expressed in nodose, trigeminal sensory neurons, as well as in some, but not all, small dorsal root ganglion (DRG) neurons and in a few medium sized DRG neurons.

The hVR-1, hVR-2, and rVR-2 molecules of the present invention play a role in pain signaling mechanisms. As used herein, the term "pain signaling mechanisms" includes the cellular mechanisms involved in the development and regulation of pain, e.g., pain elicited by noxious chemical, mechanical, or thermal stimuli, in a subject, e.g., a mammal such as a human. In mammals, the initial detection of noxious chemical. mechanical, or thermal stimuli, a process referred to as "nociception", occurs predominantly at the peripheral terminals of specialized, small diameter primary afferent neurons, called polymodal nociceptors. These afferent neurons transmit the information to the central nervous system, evoking a perception of pain or discomfort and initiating appropriate protective reflexes. Capsaicin/Vanilloid receptors, e.g., the hVR-1, hVR-2, and rVR-2 molecules of the present invention, present on these afferent neurons, are involved in detecting these noxious chemical, mechanical, or thermal stimuli and transducing this information into membrane depolarization events. Thus, the hVR-1, hVR-2, and rVR-2 molecules by participating in pain signaling mechanisms, can modulate pain elicitation and provide novel diagnostic targets and therapeutic agents to control pain.

The hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control pain in a variety of disorders, diseases, or conditions which are characterized by a deregulated, e.g., upregulated or downregulated, pain response. For example, the hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control the exaggerated pain response elicited during various forms of tissue injury, e.g., inflammation, infection, and ischemia, usually referred to as hyperalgesia (described in, for example, Fields, H.L. (1987) Pain, New York:McGraw-

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Hill). Moreover, the hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control pain associated with muscoloskeletal disorders,

e.g., joint pain; tooth pain; headaches; pain associated with surgery, or neuropathic pain.

As the hVR-1 gene maps to a region of human chromosome 17 between WI-

5436 (7.7cR) and WI-6584 (18.9cR) (Example 6), which has been associated with myasthenia gravis, Smith-Magenis syndrome, CORD5, Cone-rod dysrtophy, and breast cancer, the hVR-1 molecule may provide novel diagnostic targets and therapeutic agents to treat, diagnose, or prognose these disorders or other disorders linked to this chromosomal region. Similarly, as the hVR-2 gene maps to a region of human chromosome 17 between AFMA043ZB5 (23.3 cR) and D17S721 (29.3cR) (Example 6) which has been associated with myasthenia gravis. Smith-Magenis syndrome, CORD5, Cone-rod dysrtophy, choroidal dystrophy, central areolar, and retinal cone dystrophy, the hVR-2 molecule may provide novel diagnostic targets and therapeutic agents to treat, diagnose, or prognose these disorders or other disorders linked to this chromosomal region.

The term "family" when referring to the protein and nucleic acid molecules of the invention is intended to mean two or more proteins or nucleic acid molecules having a common structural domain or motif and having sufficient amino acid or nucleotide sequence homology as defined herein. Such family members can be naturally or non-naturally occurring and can be from either the same or different species. For example, a family can contain a first protein of human origin, as well as other, distinct proteins of human origin or alternatively, can contain homologues of non-human origin. Members of a family may also have common functional characteristics.

For example, the family of hVR-1, hVR-2, and rVR-2 proteins comprise at least one, and preferably six "transmembrane domains." As used herein, the term "transmembrane domain" includes an amino acid sequence of about 15 amino acid residues in length which spans the plasma membrane. More preferably, a transmembrane domain includes about at least 20, 25, 30, 35, 40, or 45 amino acid residues and spans the plasma membrane. Transmembrane domains are rich in hydrophobic residues, and typically have a helical structure. In a embodiment, at least 50%, 60%, 70%, 80%, 90%, 95% or more of the amino acid residues of a

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transmembrane domain are hydrophobic, *e.g.*, leucines, isoleucines, tyrosines, or tryptophans. Transmembrane domains are described in, for example, Zagotta W.N. et al, (1996) *Annual Rev. Neurosci.* 19: 235-63, the contents of which are incorporated herein by reference. Amino acid residues 434-455, 480-495, (509-531; based on homology to the rat VR-1) or 514-531, (543-569; based on homology to the rat VR-1) or 538-555, (577-596; based on homology to the rat VR-1) or 580-599, and (656-683; based on homology to the rat VR-1) or 658-682 of hVR-1 (SEQ ID NO:2) and amino acid residues 391-410, 431-448, 459-476, 486-508, 538-556, and 621-645 of hVR-2

(SEQ ID NO:5) comprise transmembrane domains.

In another embodiment, an hVR-1, hVR-2, and rVR-2 of the present invention is identified based on the presence of a "proline rich domain" in the protein or corresponding nucleic acid molecule. As used herein, the term "proline rich domain" includes an amino acid sequence of about 4-6 amino acid residues in length having the general sequence X-Pro-X-X-Pro-X (where X can be any amino acid). Proline rich domains are usually located in a helical structure and bind through hydrophobic interactions to SH3 domains. SH3 domains recognize proline rich domains in both forward and reverse orientations. Proline rich domains are described in, for example, Sattler M. et al. (1998) Leukemia 12:637-644, the contents of which are incorporated herein by reference.

In another embodiment, an hVR-1, hVR-2, and rVR-2 of the present invention is identified based on the presence of an "ankyrin repeat domain" in the protein or corresponding nucleic acid molecule. As used herein, the term "ankyrin repeat domain" includes a protein domain having an amino acid sequence of about 30-50 amino acid residues and having a bit score for the alignment of the sequence to the ankyrin repeat domain (HMM) of at least 6. Preferably, an ankyrin repeat domain includes at least about 30-45, more preferably about 30-40 amino acid residues, or about 30-35 amino acids and has a bit score for the alignment of the sequence to the ankyrin repeat domain (HMM) of at least 3-10, more preferably 10-30, more preferably 30-50, even more preferably 50-75, 75-100, 100-200 or greater. The ankyrin repeat domain HMM has been assigned the PFAM Accession PF00023 (http://genome.wustl.edu/Pfam/.html). Ankyrin repeats are involved in protein-protein interactions and are described in, for

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example, Ketchum K.A et al. (1996) FEBS Letters 378:19-26, the contents of which are incorporated herein by reference.

To identify the presence of an ankyrin repeat domain in an hVR-1, hVR-2, and rVR-2 protein and make the determination that a protein of interest has a particular profile, the amino acid sequence of the protein is searched against a database of HMMs (e.g., the Pfam database, release 2.1) using the default parameters (http://www.sanger.ac.uk/Software/Pfam/HMM_search). A description of the Pfam database can be found in Sonhammer et al. (1997) Proteins 28(3)405-420 and a detailed description of HMMs can be found, for example, in Gribskov et al.(1990) Meth. Enzymol. 183:146-159; Gribskov et al.(1987) Proc. Natl. Acad. Sci. USA 84:4355-4358; Krogh et al.(1994) J. Mol. Biol. 235:1501-1531; and Stultz et al.(1993) Protein Sci. 2:305-314, the contents of which are incorporated herein by reference. A search was performed against the HMM database resulting in the identification of three ankyrin repeat domains in the amino acid sequence of SEQ ID NO:2 (at about residues 201-233, 248-283, and 333-361) and SEQ ID NO:5 (at about residues 162-194, 208-243, and 293-328). The results of the searches are set forth in Figures 13 and 15.

Isolated proteins of the present invention, preferably hVR-1, hVR-2, and rVR-2 proteins, have an amino acid sequence sufficiently identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11 or are encoded by a nucleotide sequence sufficiently identical to SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. As used herein, the term "sufficiently identical" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain) amino acid residues or nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences share common structural domains or motifs and/or a common functional activity. For example, amino acid or nucleotide sequences which share common structural domains have at least 30%, 40%, or 50% identity, preferably 60% identity, more preferably 70%-80%, and even more preferably 90-95% identity across the amino acid sequences of the domains and contain at least one and preferably two structural domains or motifs, are defined herein as sufficiently identical. Furthermore, amino acid or nucleotide sequences which share at least 30%, 40%,

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or 50%, preferably 60%, more preferably 70-80%, or 90-95% identity and share a common functional activity are defined herein as sufficiently identical.

As used interchangeably herein, an "hVR-1, hVR-2, and rVR-2 activity", "biological activity of hVR-1, hVR-2, and rVR-2" or "functional activity of hVR-1, hVR-2, and rVR-2", refers to an activity exerted by an hVR-1, hVR-2, and rVR-2 protein, polypeptide or nucleic acid molecule on an hVR-1, hVR-2, and rVR-2 responsive cell or on an hVR-1, hVR-2, and rVR-2 protein substrate, as determined in vivo, or in vitro, according to standard techniques. In one embodiment, an hVR-1, hVR-2, and rVR-2 activity is a direct activity, such as an association with an hVR-1, hVR-2, and rVR-2-target molecule. As used herein, a "target molecule" or "binding partner" is a molecule with which an hVR-1, hVR-2, and rVR-2 protein binds or interacts in nature, such that hVR-1, hVR-2, and rVR-2-mediated function is achieved. An hVR-1, hVR-2, and rVR-2 target molecule can be a non-hVR-1, non-hVR-2, and non-rVR-2 molecule or an hVR-1, hVR-2, and rVR-2 protein or polypeptide of the present invention. In an exemplary embodiment, an hVR-1, hVR-2, and rVR-2 target molecule is an hVR-1, hVR-2, and rVR-2 ligand, e.g., capsaicin. Alternatively, an hVR-1, hVR-2, and rVR-2 activity is an indirect activity, such as a cellular signaling activity mediated by interaction of the hVR-1, hVR-2, and rVR-2 protein with an hVR-1, hVR-2, and rVR-2 ligand.

Accordingly, another embodiment of the invention features isolated hVR-1, 20 hVR-2, and rVR-2 proteins and polypeptides having an hVR-1, hVR-2, and rVR-2 activity. Other proteins of the invention are hVR-1, hVR-2, and rVR-2 proteins having at least one, and preferably six, transmembrane domains and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Yet other proteins of the invention are hVR-1, hVR-2, and rVR-2 proteins having at least one transmembrane domain, at least one proline rich domain and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Other proteins of the 25 invention are hVR-1, hVR-2, and rVR-2 proteins having at least one transmembrane domain, at least one proline rich domain, at least one ankyrin repeat domain and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Additional proteins of the invention have at least one transmembrane domain, at least one proline rich domain, at least one 30 ankyrin repeat domain, and are, preferably, encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a

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nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

The nucleotide sequence of the full length hVR-1 cDNA and the predicted amino acid sequence of the hVR-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively.

The nucleotide sequence of the full length hVR-2 cDNA and the predicted amino acid sequence of the hVR-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively.

The nucleotide sequence of the partial hVR-2 (alternate form) cDNA and the predicted amino acid sequence of the hVR-2 (alternate form) polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively.

The nucleotide sequence of the partial rVR-2 cDNA and the predicted amino acid sequence of the rVR-2 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively.

Various aspects of the invention are described in further detail in the following subsections:

I. Isolated Nucleic Acid Molecules

One aspect of the invention pertains to isolated nucleic acid molecules that encode hVR-1, hVR-2, and rVR-2 proteins or biologically active portions thereof, as well as nucleic acid fragments sufficient for use as hybridization probes to identify hVR-1, hVR-2, and rVR-2-encoding nucleic acid molecules (e.g., hVR-1, hVR-2, and rVR-2 mRNA) and fragments for use as PCR primers for the amplification or mutation of hVR-1, hVR-2, and rVR-2 nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (e.g., cDNA or genomic DNA) and RNA molecules (e.g., mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

The term "isolated nucleic acid molecule" includes nucleic acid molecules which are separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. For example, with regards to genomic DNA, the term "isolated"

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- 17 -

includes nucleic acid molecules which are separated from the chromosome with which the genomic DNA is naturally associated. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated hVR-1, hVR-2, and rVR-2 nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, *e.g.*, a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. Using all or portion of the nucleic acid sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, as a hybridization probe, hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be isolated using standard hybridization and cloning techniques (*e.g.*, as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

Moreover, a nucleic acid molecule encompassing all or a portion of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to hVR-1, hVR-2, and rVR-2 nucleotide sequences can be prepared by standard synthetic techniques, *e.g.*, using an automated DNA synthesizer.

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In one embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:1. The sequence of SEQ ID NO:1 corresponds to the full length hVR-1 encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:4. The sequence of SEQ ID NO:4 corresponds to the full length hVR-2 encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:7. The sequence of SEQ ID NO:7 corresponds to a fragment of the hVR-2 (alternate form) encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:10. The sequence of SEQ ID NO:10 corresponds to a fragment of the rVR-2 cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion of any of these nucleotide sequences. A nucleic acid molecule which is complementary to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, is one which is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, such that it can hybridize to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 thereby forming a stable duplex.

In still another embodiment, an isolated nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least about 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous to the entire length of the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion of any of these nucleotide sequences.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, for example, a fragment which can be used as a probe or primer or a fragment encoding a portion of an hVR-1, hVR-2, and rVR-2 protein, e.g., a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein. The nucleotide sequence determined from the cloning of the hVR-1, hVR-2, and rVR-2 gene allows for the generation of probes and primers

- 19 -

designed for use in identifying and/or cloning other hVR-1, hVR-2. and rVR-2 family members, as well as hVR-1, hVR-2, and rVR-2 homologues from other species. The probe/primer typically comprises a substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12 or 15, preferably about 20 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, 75, or 100 consecutive nucleotides of a sense sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, of an anti-sense sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or of a naturally occurring allelic variant or mutant of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In an exemplary embodiment, a nucleic acid molecule of the present invention comprises a nucleotide sequence which is greater than 100-150, 150-200, 200-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, 550-600, 600-650, 650-700, 700-750, 750-800, 800-850, 850-900, 900-950, 950-1000, 1088, or more nucleotides in length and hybridizes under stringent hybridization conditions to a nucleic acid molecule of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

Probes based on the hVR-1, hVR-2, and rVR-2 nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments, the probe further comprises a label group attached thereto, *e.g.*, the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress an hVR-1, hVR-2, and rVR-2 protein, such as by measuring a level of an hVR-1, hVR-2, and rVR-2-encoding nucleic acid in a sample of cells from a subject *e.g.*, detecting hVR-1, hVR-2, and rVR-2 mRNA levels or determining whether a genomic hVR-1, hVR-2, and rVR-2 gene has been mutated or deleted.

A nucleic acid fragment encoding a "biologically active portion of an hVR-1, hVR-2, and rVR-2 protein" can be prepared by isolating a portion of the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, which encodes a polypeptide having an hVR-1, hVR-2, and rVR-2 biological activity (the biological activities of the hVR-1, hVR-2, and rVR-2 proteins are described herein), expressing the encoded portion of the hVR-1, hVR-2, and rVR-2 protein (e.g., by recombinant expression in vitro) and assessing the activity of the encoded portion of the hVR-1, hVR-2, and rVR-2 protein.

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The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, due to degeneracy of the genetic code and thus encode the same hVR-1, hVR-2, and rVR-2 proteins as those encoded by the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11.

In addition to the hVR-1, hVR-2, and rVR-2 nucleotide sequences shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the hVR-1, hVR-2, and rVR-2 proteins may exist within a population (*e.g.*, the human population). Such genetic polymorphism in the hVR-1, hVR-2, and rVR-2 genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which include an open reading frame encoding an hVR-1, hVR-2, and rVR-2 protein, preferably a mammalian hVR-1, hVR-2, and rVR-2 protein, and can further include noncoding regulatory sequences, and introns.

Allelic variants of hVR-1, hVR-2, and rVR-2 include both functional and non-functional hVR-1, hVR-2, and rVR-2 proteins. Functional allelic variants are naturally occurring amino acid sequence variants of the hVR-1, hVR-2, and rVR-2 protein that maintain the ability to bind an hVR-1, hVR-2, and rVR-2 ligand and/or modulate a pain signaling mechanism. Functional allelic variants will typically contain only conservative substitution of one or more amino acids of SEQ ID NO:2, 5, 8, or 11, or substitution, deletion or insertion of non-critical residues in non-critical regions of the protein.

Non-functional allelic variants are naturally occurring amino acid sequence variants of the hVR-1, hVR-2, and rVR-2 protein that do not have the ability to either bind an hVR-1, hVR-2, and rVR-2 ligand and/or modulate a pain signaling mechanism. Non-functional allelic variants will typically contain a non-conservative substitution, a deletion, or insertion or premature truncation of the amino acid sequence of SEQ ID

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- 21 -

NO:2, 5, 8, or 11, or a substitution, insertion or deletion in critical residues or critical regions.

The present invention further provides non-human orthologues of the hVR-2 and rVR-2 protein. Orthologues of the hVR-2 and rVR-2 protein are proteins that are isolated from non-human and non-rat organisms and possess the same hVR-2 and rVR-2 ligand binding and/or modulation of pain signaling mechanism capabilities of the hVR-2 and rVR-2 proteins. Orthologues of the hVR-2 and rVR-2 proteins can readily be identified as comprising an amino acid sequence that is substantially homologous to SEQ ID NO: 4, 6, 8 or 10.

Moreover, nucleic acid molecules encoding other hVR-1, hVR-2, and rVR-2 family members and, thus, which have a nucleotide sequence which differs from the hVR-1, hVR-2, and rVR-2 sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, are intended to be within the scope of the invention. For example, another hVR-1, hVR-2, and rVR-2 cDNA can be identified based on the nucleotide sequence of hVR-1, hVR-2, and rVR-2. Moreover, nucleic acid molecules encoding VR-2 proteins from different species, and which, thus, have a nucleotide sequence which differs from the hVR-2 and rVR-2 sequences of SEQ ID NO:4, 6, 8, or 10 are intended to be within the scope of the invention. For example, a mouse hVR-2 cDNA can be identified based on the nucleotide sequence of the human VR-2 (hVR-2) or the rat VR-2 (rVR-2).

Nucleic acid molecules corresponding to natural allelic variants and homologues of the hVR-1, hVR-2, and rVR-2 cDNAs of the invention can be isolated based on their homology to the hVR-1, hVR-2, and rVR-2 nucleic acids disclosed herein using the cDNAs disclosed herein, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. Nucleic acid molecules corresponding to natural allelic variants and homologues of the hVR-1, hVR-2, and rVR-2 cDNAs of the invention can further be isolated by mapping to the same chromosome or locus as the hVR-1, hVR-2, and rVR-2 gene.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15, 20, 25, 30 or more nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In other embodiment, the nucleic acid is at least

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- 22 -

30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, or 950 nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% identical to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least 5 about 70%, more preferably at least about 80%, even more preferably at least about 85% or 90% identical to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, nonlimiting example of stringent hybridization conditions are hybridization in 6X sodium 10 chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50°C, preferably at 55°C, more preferably at 60°C, and even more preferably at 65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In addition to naturally-occurring allelic variants of the hVR-1, hVR-2, and rVR-2 sequences that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, thereby leading to changes in the amino acid sequence of the encoded hVR-1, hVR-2, and rVR-2 proteins, without altering the functional ability of the hVR-1, hVR-2, and rVR-2 proteins. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of hVR-1, hVR-2, and rVR-2 (e.g., the sequence of SEQ ID NO:2, 5, 8, or 11) without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the hVR-1, hVR-2, and rVR-2 proteins of the present invention, are predicted to be particularly unamenable to alteration. Furthermore,

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- 23 -

additional amino acid residues that are conserved between the hVR-1, hVR-2, and rVR-2 proteins of the present invention and other members of the Capsaicin/Vanilloid receptor family are not likely to be amenable to alteration.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins that contain changes in amino acid residues that are not essential for activity. Such hVR-1, hVR-2, and rVR-2 proteins differ in amino acid sequence from SEQ ID NO:2, 5, 8, or 11, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to SEQ ID NO:2, 5, 8, or 11.

An isolated nucleic acid molecule encoding an hVR-1, hVR-2, and rVR-2 protein homologous to the protein of SEQ ID NO:2, 5, 8, or 11 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in an hVR-1, hVR-2, and rVR-2 protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of an hVR-1, hVR-

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2, and rVR-2 coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for hVR-1, hVR-2, and rVR-2 biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

In a embodiment, a mutant hVR-1, hVR-2, and rVR-2 protein can be assayed for the ability to (1) interact with a non-hVR-1, non-hVR-2, or non- rVR-2 protein molecule, e.g., a vanilloid compound such as capsaicin; (2) modulate intracellular calcium concentration; (3) activate an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; or (4) modulate a pain signaling mechanism.

In addition to the nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire hVR-1, hVR-2, and rVR-2 coding strand, or to only a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding hVR-1, hVR-2, and rVR-2. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (e.g., the coding region of hVR-1, hVR-2, and rVR-2). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding hVR-1, hVR-2, and rVR-2. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding hVR-1, hVR-2, and rVR-2 disclosed herein, antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of hVR-1, hVR-2, and rVR-2 mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding

- 25 -

or noncoding region of hVR-1, hVR-2, and rVR-2 mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of hVR-1, hVR-2, and rVR-2 mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5- oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (i.e., RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an hVR-1, hVR-2, and rVR-2 protein to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The

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- 26 -

hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention include direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, *e.g.*, by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an -anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids*. *Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue *et al.* (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) Nature 334:585-591)) can be used to catalytically cleave hVR-1, hVR-2, and rVR-2 mRNA transcripts to thereby inhibit translation of hVR-1, hVR-2, and rVR-2 mRNA. A ribozyme having specificity for an hVR-1, hVR-2, and rVR-2-encoding nucleic acid can be designed based upon the nucleotide sequence of an hVR-1, hVR-2, and rVR-2 cDNA disclosed herein (i.e., SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12). For example, a derivative of a Tetrahymena L-19 IVS

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- 27 -

RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an hVR-1, hVR-2, and rVR-2-encoding mRNA. See, e.g., Cech et al. U.S. Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, hVR-1, hVR-2, and rVR-2 mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, for example, Bartel, D. and Szostak, J.W. (1993) Science 261:1411-1418.

Alternatively, hVR-1, hVR-2, and rVR-2 gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the hVR-1, hVR-2, and rVR-2 (e.g., the hVR-1, hVR-2, and rVR-2 promoter and/or enhancers) to form triple helical structures that prevent transcription of the hVR-1, hVR-2, and rVR-2 gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. et al. (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher, L.J. (1992) *Bioassays* 14(12):807-15.

In yet another embodiment, the hVR-1, hVR-2, and rVR-2 nucleic acid molecules of the present invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acid molecules can be modified to generate peptide nucleic acids (see Hyrup B. et al. (1996) Bioorganic & Medicinal Chemistry 4 (1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup B. et al. (1996) supra; Perry-O'Keefe et al. Proc. Natl. Acad. Sci. 93: 14670-675.

PNAs of hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, for example, inducing transcription or translation arrest or inhibiting replication. PNAs of hVR-1,

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- 28 -

hVR-2, and rVR-2 nucleic acid molecules can also be used in the analysis of single base pair mutations in a gene, (e.g., by PNA-directed PCR clamping); as 'artificial restriction enzymes' when used in combination with other enzymes, (e.g., S1 nucleases (Hyrup B. (1996) supra)); or as probes or primers for DNA sequencing or hybridization (Hyrup B. et al. (1996) supra; Perry-O'Keefe supra).

In another embodiment, PNAs of hVR-1, hVR-2, and rVR-2 can be modified, (e.g., to enhance their stability or cellular uptake), by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, (e.g., RNAse H and DNA polymerases), to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup B. (1996) supra). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup B. (1996) supra and Finn P.J. et al. (1996) Nucleic Acids Res. 24 (17): 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs, e.g., 5'-(4methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used as a between the PNA and the 5' end of DNA (Mag, M. et al. (1989) Nucleic Acid Res. 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn P.J. et al. (1996) supra). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser, K.H. et al. (1975) Bioorganic Med. Chem. Lett. 5: 1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors in vivo), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) Proc. Natl. Acad. Sci. USA 86:6553-6556; Lemaitre et al. (1987) Proc. Natl. Acad. Sci. USA 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization-

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triggered cleavage agents (See, e.g., Krol et al. (1988) Bio-Techniques 6:958-976) or intercalating agents. (See, e.g., Zon (1988) Pharm. Res. 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, (e.g., a peptide, hybridization triggered cross-linking agent, transport agent, or hybridization-triggered cleavage agent).

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II. Isolated hVR-1, hVR-2, and rVR-2 Proteins and Anti-hVR-1, Anti-hVR-2, and Anti-rVR-2 Antibodies

One aspect of the invention pertains to isolated hVR-1, hVR-2, and rVR-2 proteins, and biologically active portions thereof, as well as polypeptide fragments suitable for use as immunogens to raise anti-hVR-2, anti-hVR-2, and anti-rVR-2 antibodies. In one embodiment, native hVR-1, hVR-2, and rVR-2 proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, hVR-1, hVR-2, and rVR-2 proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, an hVR-1, hVR-2, and rVR-2 protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the hVR-1, hVR-2, and rVR-2 protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of hVR-1, hVR-2, and rVR-2 protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of hVR-1, hVR-2, and rVR-2 protein having less than about 30% (by dry weight) of non-hVR-1, hVR-2, and rVR-2 protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-hVR-1, hVR-2, and rVR-2 protein, still more preferably less than about 10% of non-hVR-1, hVR-2, and rVR-2 protein, and most preferably less than about 5% non-hVR-1, non-hVR-2, and non-rVR-2 protein. When the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture

medium, *i.e.*, culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of hVR-1, hVR-2, and rVR-2 protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of hVR-1, hVR-2, and rVR-2 protein having less than about 30% (by dry weight) of chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, more preferably less than about 20% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, still more preferably less than about 10% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, and most preferably less than about 5% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals.

As used herein, a "biologically active portion" of an hVR-1, hVR-2, and rVR-2 protein includes a fragment of an hVR-1, hVR-2, and rVR-2 protein which participates in an interaction between an hVR-1, hVR-2, and rVR-2 molecule and a non-hVR-1, non-hVR-2, and non-rVR-2 molecule, respectively. Biologically active portions of an hVR-1, hVR-2, and rVR-2 protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the hVR-1, hVR-2, and rVR-2 protein, e.g., the amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11, which include less amino acids than the full length hVR-1, hVR-2, and rVR-2 proteins, and exhibit at least one activity of an hVR-1, hVR-2, and rVR-2 protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the hVR-1, hVR-2, and rVR-2 protein, e.g., binding of an hVR-1, hVR-2, and rVR-2 ligand such as a vanilloid compound, e.g., Capsaicin. A biologically active portion of an hVR-1, hVR-2, and rVR-2 protein can be a polypeptide which is, for example, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200 or more amino acids in length. Biologically active portions of an hVR-1, hVR-2, and rVR-2 protein can be used as targets for developing agents which modulate an hVR-1, hVR-2, and rVR-2 mediated activity, e.g., a pain signaling mechanism.

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- 31 -

In one embodiment, a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein comprises at least one transmembrane domain, and/or at least one proline rich domain, and/or at least one ankyrin repeat domain. It is to be understood that a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein of the present invention may contain at least one of the above-identified structural domains. A more biologically active portion of an hVR-1, hVR-2, and rVR-2 protein may contain at least two of the above-identified structural domains. Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native hVR-1, hVR-2, and rVR-2 protein.

In a embodiment, the hVR-1, hVR-2, and rVR-2 protein has an amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11. In other embodiments, the hVR-1, hVR-2, and rVR-2 protein is substantially homologous to SEQ ID NO:2, 5, 8, or 11, and retains the functional activity of the protein of SEQ ID NO:2, 5, 8, or 11, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail in subsection I above. Accordingly, in another embodiment, the hVR-1, hVR-2, and rVR-2 protein is a protein which comprises an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to SEQ ID NO:2, 5, 8, or 11.

To determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, or 90% of the length of the reference sequence (e.g., when aligning a second sequence to the hVR-1, hVR-2, and rVR-2 amino acid sequence of SEQ ID NO:2, 5, 8, or 11, having 177 amino acid residues, at least 80, preferably at least 100, more preferably at least 120, even more preferably at least 140, and even more preferably at least 150, 160 or 170 amino acid residues are aligned). The amino acid residues or nucleotides at

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corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a embodiment, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch (*J. Mol. Biol.* (48):444-453 (1970)) algorithm which has been incorporated into the GAP program in the GCG software package (available at http://www.gcg.com), using either a Blossum 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. In another embodiment, the percent identity between two amino acid or nucleotide sequences is determined using the algorithm of E. Meyers and W. Miller (CABIOS, 4:11-17 (1989)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The nucleic acid and protein sequences of the present invention can further be

25 used as a "query sequence" to perform a search against public databases to, for example, identify other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (1990) J.

Mol. Biol. 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences

30 homologous to hVR-1, hVR-2, and rVR-2 nucleic acid molecules of the invention.

BLAST protein searches can be performed with the XBLAST program, score = 50,

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- 33 -

wordlength = 3 to obtain amino acid sequences homologous to hVR-1, hVR-2, and rVR-2 protein molecules of the invention. To obtain gapped alignments for comparison purposes. Gapped BLAST can be utilized as described in Altschul *et al.*, (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (*e.g.*, XBLAST and NBLAST) can be used. See http://www.ncbi.nlm.nih.gov.

The invention also provides hVR-1, hVR-2, and rVR-2 chimeric or fusion proteins. As used herein, an hVR-1, hVR-2, and rVR-2 "chimeric protein" or "fusion protein" comprises an hVR-1, hVR-2, and rVR-2 polypeptide operatively linked to a non-hVR-1, hVR-2, and rVR-2 polypeptide. An "hVR-1, hVR-2, and rVR-2 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to hVR-1, hVR-2, and rVR-2, whereas a "non-hVR-1, non-hVR-2, and non-rVR-2 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the hVR-1, hVR-2, and rVR-2 protein, 15 e.g., a protein which is different from the hVR-1, hVR-2, and rVR-2 protein and which is derived from the same or a different organism. Within an hVR-1, hVR-2, and rVR-2 fusion protein the hVR-1, hVR-2, and rVR-2 polypeptide can correspond to all or a portion of an hVR-1, hVR-2, and rVR-2 protein. In a embodiment, an hVR-1, hVR-2, and rVR-2 fusion protein comprises at least one biologically active portion of an hVR-1, hVR-2, and rVR-2 protein. In another embodiment, an hVR-1, hVR-2, and rVR-2 20 fusion protein comprises at least two biologically active portions of an hVR-1, hVR-2, and rVR-2 protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the hVR-1, hVR-2, and rVR-2 polypeptide and the non-hVR-1, nonhVR-2, and non-rVR-2 polypeptide are fused in-frame to each other. The non-hVR-1, hVR-2, and rVR-2 polypeptide can be fused to the N-terminus or C-terminus of the hVR-1, hVR-2, and rVR-2 polypeptide.

For example, in one embodiment, the fusion protein is a GST-hVR-1, GST-hVR-2, and GST-rVR-2 fusion protein in which the hVR-1, hVR-2, and rVR-2 sequences are fused to the C-terminus of the GST sequences. Such fusion proteins can facilitate the purification of recombinant hVR-1, hVR-2, and rVR-2.

- 34 -

In another embodiment, the fusion protein is an hVR-1, hVR-2, and rVR-2 protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of hVR-1, hVR-2, and rVR-2 can be increased through use of a heterologous signal sequence.

The hVR-1, hVR-2, and rVR-2 fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject *in vivo*. The hVR-1, hVR-2, and rVR-2 fusion proteins can be used to affect the bioavailability of an hVR-1, hVR-2, and rVR-2 substrate. Use of hVR-1, hVR-2, and rVR-2 fusion proteins may be useful therapeutically for the treatment of disorders caused by, for example, (i) aberrant modification or mutation of a gene encoding an hVR-1, hVR-2, and rVR-2 protein; (ii) mis-regulation of the hVR-1, hVR-2, and rVR-2 gene; and (iii) aberrant post-translational modification of an hVR-1, hVR-2, and rVR-2 protein.

Moreover, the hVR-1, hVR-2, and rVR-2-fusion proteins of the invention can be used as immunogens to produce anti-hVR-1. anti-hVR-2, and anti-rVR-2 antibodies in a subject, to purify hVR-1, hVR-2, and rVR-2 ligands and in screening assays to identify molecules which inhibit the interaction of hVR-1, hVR-2, and rVR-2 with an hVR-1, hVR-2, and rVR-2 substrate.

Preferably, an hVR-1, hVR-2, and rVR-2 chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel *et al.* John Wiley & Sons: 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST

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polypeptide). An hVR-1, hVR-2, and rVR-2-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the hVR-1, hVR-2, and rVR-2 protein.

The present invention also pertains to variants of the hVR-1, hVR-2, and rVR-2 proteins which function as either hVR-1, hVR-2, and rVR-2 agonists (mimetics) or as hVR-1, hVR-2, and rVR-2 antagonists. Variants of the hVR-1, hVR-2, and rVR-2 proteins can be generated by mutagenesis, *e.g.*, discrete point mutation or truncation of an hVR-1, hVR-2, and rVR-2 protein. An agonist of the hVR-1, hVR-2, and rVR-2 proteins can retain substantially the same, or a subset, of the biological activities of the naturally occurring form of an hVR-1, hVR-2, and rVR-2 protein. An antagonist of an hVR-1, hVR-2, and rVR-2 protein can inhibit one or more of the activities of the naturally occurring form of the hVR-1, hVR-2, and rVR-2 protein by, for example, competitively modulating an hVR-1, hVR-2, and rVR-2-mediated activity of an hVR-1, hVR-2, and rVR-2 protein. Thus, specific biological effects can be elicited by treatment with a variant of limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the protein has fewer side effects in a subject relative to treatment with the naturally occurring form of the hVR-1, hVR-2, and rVR-2 protein.

In one embodiment, variants of an hVR-1, hVR-2, and rVR-2 protein which 20 function as either hVR-1, hVR-2, and rVR-2 agonists (mimetics) or as hVR-1, hVR-2, and rVR-2 antagonists can be identified by screening combinatorial libraries of mutants, e.g., truncation mutants, of an hVR-1, hVR-2, and rVR-2 protein for hVR-1, hVR-2, and rVR-2 protein agonist or antagonist activity. In one embodiment, a variegated library of hVR-1, hVR-2, and rVR-2 variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of 25 hVR-1, hVR-2, and rVR-2 variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential hVR-1, hVR-2, and rVR-2 sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage 30 display) containing the set of hVR-1, hVR-2, and rVR-2 sequences therein. There are a variety of methods which can be used to produce libraries of potential hVR-1, hVR-2,

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- 36 -

and rVR-2 variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential hVR-1, hVR-2, and rVR-2 sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, e.g., Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura et al. (1984) *Annu. Rev. Biochem.* 53:323; Itakura et al. (1984) *Science* 198:1056; Ike et al. (1983) *Nucleic Acid Res.* 11:477.

In addition, libraries of fragments of an hVR-1, hVR-2, and rVR-2 protein coding sequence can be used to generate a variegated population of hVR-1, hVR-2, and rVR-2 fragments for screening and subsequent selection of variants of an hVR-1, hVR-2. and rVR-2 protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of an hVR-1, hVR-2, and rVR-2 coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the hVR-1, hVR-2, and rVR-2 protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of hVR-1, hVR-2, and rVR-2 proteins. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recrusive ensemble mutagenesis

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- 37 -

(REM), a new technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify hVR-1, hVR-2, and rVR-2 variants (Arkin and Yourvan (1992) *Proc. Natl. Acad. Sci. USA* 89:7811-7815; Delgrave *et al.* (1993) *Protein Engineering* 6(3):327-331).

In one embodiment, cell based assays can be exploited to analyze a variegated hVR-1, hVR-2, and rVR-2 library. For example, a library of expression vectors can be transfected into a cell line, *e.g.*, a neuronal cell line, which ordinarily responds to a particular ligand in an hVR-1, hVR-2, and rVR-2-dependent manner. The transfected cells are then contacted with the ligand and the effect of expression of the mutant on signaling by the ligand can be detected, *e.g.*, by measuring intracellular calcium concentration, neuronal membrane depolarization, or the activity of an hVR-1, hVR-2, and rVR-2-regulated transcription factor. Plasmid DNA can then be recovered from the cells which score for inhibition, or alternatively, potentiation of signaling by the ligand, and the individual clones further characterized.

An isolated hVR-1, hVR-2, and rVR-2 protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind hVR-1, hVR-2, and rVR-2 using standard techniques for polyclonal and monoclonal antibody preparation. A full-length hVR-1, hVR-2, and rVR-2 protein can be used or, alternatively, the invention provides antigenic peptide fragments of hVR-1, hVR-2, and rVR-2 for use as immunogens. The antigenic peptide of hVR-1, hVR-2, and rVR-2 comprises at least 8 amino acid residues of the amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11 and encompasses an epitope of hVR-1, hVR-2, and rVR-2 such that an antibody raised against the peptide forms a specific immune complex with hVR-1, hVR-2, and rVR-2. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues.

Epitopes encompassed by the antigenic peptide are regions of hVR-1, hVR-2, and rVR-2 that are located on the surface of the protein, e.g., hydrophilic regions, as well as regions with high antigenicity (see, for example, Figures 12 and 14).

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An hVR-1, hVR-2, and rVR-2 immunogen typically is used to prepare antibodies by immunizing a suitable subject, (e.g., rabbit, goat, mouse or other mammal) with the immunogen. An appropriate immunogenic preparation can contain, for example, recombinantly expressed hVR-1, hVR-2, and rVR-2 protein or a chemically synthesized hVR-1, hVR-2, and rVR-2 polypeptide. The preparation can further include an adjuvant, such as Freund's complete or incomplete adjuvant, or similar immunostimulatory agent. Immunization of a suitable subject with an immunogenic hVR-1, hVR-2, and rVR-2 preparation induces a polyclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody response.

Accordingly, another aspect of the invention pertains to anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site which specifically binds (immunoreacts with) an antigen, such as hVR-1, hVR-2, and rVR-2. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind hVR-1, hVR-2, and rVR-2. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of hVR-1, hVR-2, and rVR-2. A monoclonal antibody composition thus typically displays a single binding affinity for a particular hVR-1, hVR-2, and rVR-2 protein with which it immunoreacts.

Polyclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies can be prepared as described above by immunizing a suitable subject with an hVR-1, hVR-2, and rVR-2 immunogen. The anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized hVR-1, hVR-2, and rVR-2. If desired, the antibody molecules directed against hVR-1, hVR-2, and rVR-2 can be isolated from the mammal (e.g., from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction. At an

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- 39 -

appropriate time after immunization, e.g., when the anti-hVR-1, anti-hVR-2, and antirVR-2 antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) Nature 256:495-497) (see also, Brown et al. (1981) J. Immunol. 127:539-46; Brown et al. (1980) J. Biol. Chem .255:4980-83; Yeh et al. (1976) Proc. Natl. Acad. Sci. USA 76:2927-31; and Yeh et al. (1982) Int. J. Cancer 29:269-75), the more recent human B cell hybridoma technique (Kozbor et al. (1983) Immunol Today 4:72), the EBVhybridoma technique (Cole et al. (1985), Monoclonal Antibodies and Cancer Therapy, 10 Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing monoclonal antibody hybridomas is well known (see generally R. H. Kenneth, in Monoclonal Antibodies: A New Dimension In Biological Analyses, Plenum Publishing Corp., New York, New York (1980); E. A. Lerner (1981) Yale J. Biol. Med., 54:387402; M. L. Gefter et al. (1977) Somatic Cell Genet. 3:23136). Briefly, an immortal cell line 15 (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal immunized with an hVR-1, hVR-2, and rVR-2 immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds hVR-1, hVR-2, and rVR-2.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating an anti-hVR-1, anti-20 hVR-2, and anti-rVR-2 monoclonal antibodies (see, e.g., G. Galfre et al. (1977) Nature 266:55052; Gefter et al. Somatic Cell Genet., cited supra; Lerner, Yale J. Biol. Med., cited supra; Kenneth, Monoclonal Antibodies, cited supra). Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods which also 25 would be useful. Typically, the immortal cell line (e.g., a myeloma cell line) is derived from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line. immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a 30 number of myeloma cell lines can be used as a fusion partner according to standard

- 40 -

techniques, e.g., the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind hVR-1, hVR-2, and rVR-2, e.g., using a standard ELISA assay.

10 Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with hVR-1, hVR-2, and rVR-2 to thereby isolate immunoglobulin library members that bind hVR-1, hVR-2, and rVR-2. Kits for generating and screening phage display libraries are commercially available (e.g., the 15 Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; and the Stratagene SurfZAPTM Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, Ladner et al. U.S. Patent No. 5,223,409; Kang et al. PCT International Publication No. WO 92/18619; Dower et al. 20 PCT International Publication No. WO 91/17271; Winter et al. PCT International Publication WO 92/20791; Markland et al. PCT International Publication No. WO 92/15679; Breitling et al. PCT International Publication WO 93/01288; McCafferty et al. PCT International Publication No. WO 92/01047; Garrard et al. PCT International 25 Publication No. WO 92/09690; Ladner et al. PCT International Publication No. WO 90/02809; Fuchs et al. (1991) Bio/Technology 9:1370-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; Griffiths et al. (1993) EMBO J 12:725-734; Hawkins et al. (1992) J. Mol. Biol. 226:889-896; Clarkson et al. (1991) Nature 352:624-628; Gram et al. (1992) Proc. Natl. Acad. Sci. USA 89:3576-3580; Garrad et al. (1991) Bio/Technology 9:1373-1377; Hoogenboom et al. 30

-41-

(1991) Nuc. Acid Res. 19:4133-4137; Barbas et al. (1991) Proc. Natl. Acad. Sci. USA 88:7978-7982; and McCafferty et al. Nature (1990) 348:552-554.

Additionally, recombinant anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson et al. International Application No. PCT/US86/02269; Akira, et al. European Patent Application 184,187; Taniguchi, M., European Patent Application 171,496; Morrison et al. European Patent Application 10 173,494; Neuberger et al. PCT International Publication No. WO 86/01533; Cabilly et al. U.S. Patent No. 4,816,567; Cabilly et al. European Patent Application 125,023; Better et al. (1988) Science 240:1041-1043; Liu et al. (1987) Proc. Natl. Acad. Sci. USA 84:3439-3443; Liu et al. (1987) J. Immunol. 139:3521-3526; Sun et al. (1987) Proc. Natl. Acad. Sci. USA 84:214-218; Nishimura et al. (1987) Canc. Res. 47:999-1005; Wood et al. (1985) Nature 314:446-449; and Shaw et al. (1988) J. Natl. Cancer Inst. 80:1553-1559); Morrison, S. L. (1985) Science 229:1202-1207; Oi et al. (1986) BioTechniques 4:214; Winter U.S. Patent 5,225,539; Jones et al. (1986) Nature 321:552-525; Verhoeyan et al. (1988) Science 239:1534; and Beidler et al. (1988) J. 20 Immunol. 141:4053-4060.

An anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody (*e.g.*, monoclonal antibody) can be used to isolate hVR-1, hVR-2, and rVR-2 by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can facilitate the purification of natural hVR-1, hVR-2, and rVR-2 from cells and of recombinantly produced hVR-1, hVR-2, and rVR-2 expressed in host cells. Moreover, an anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can be used to detect hVR-1, hVR-2, and rVR-2 protein (*e.g.*, in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the hVR-1, hVR-2, and rVR-2 protein. Anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can

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- 42 -

be facilitated by coupling (*i.e.*, physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

III. Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding an hVR-1, hVR-2, and rVR-2 protein (or a portion thereof). As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective

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retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression of the nucleotide sequence (e.g., in an in vitro transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cells and those which direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, and the like. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., hVR-1, hVR-2, and rVR-2 proteins, mutant forms of hVR-1, hVR-2, and rVR-2 proteins, fusion proteins, and the like).

The recombinant expression vectors of the invention can be designed for expression of hVR-1, hVR-2, and rVR-2 proteins in prokaryotic or eukaryotic cells. For example, hVR-1, hVR-2, and rVR-2 proteins can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the

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recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith, D.B. and Johnson, K.S. (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Purified fusion proteins can be utilized in hVR-1, hVR-2, and rVR-2 activity assays, (e.g., direct assays or competitive assays described in detail below), or to, for example, generate antibodies specific for hVR-1, hVR-2, and rVR-2 proteins. In a embodiment, an hVR-1, hVR-2, and rVR-2 fusion protein expressed in a retroviral expression vector of the present invention can be utilized to infect bone marrow cells which are subsequently transplanted into irradiated recipients. The pathology of the subject recipient is then examined after sufficient time has passed (e.g., six (6) weeks).

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann *et al.*, (1988) *Gene* 69:301-315) and pET 11d (Studier *et al.*, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral

- 45 -

polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada *et al.*, (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the hVR-1, hVR-2, and rVR-2 expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerivisae* include pYepSec1 (Baldari, *et al.*, (1987) *Embo J.* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.*, (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and picZ (InVitrogen Corp., San Diego, CA).

Alternatively, hVR-1, hVR-2, and rVR-2 proteins can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith et al. (1983) Mol. Cell Biol. 3:2156-2165) and the pVL series (Lucklow and Summers (1989) Virology 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, B. (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring*

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Harbor Laboratory. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-5 specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert et al. (1987) Genes Dev. 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) Adv. Immunol. 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) EMBO J. 8:729-733) and immunoglobulins (Banerji et al. (1983) Cell 10 33:729-740; Queen and Baltimore (1983) Cell 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) Proc. Natl. Acad. Sci. USA 86:5473-5477), pancreas-specific promoters (Edlund et al. (1985) Science 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 15 4,873,316 and European Application Publication No. 264,166). Developmentallyregulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) Science 249:374-379) and the α-fetoprotein promoter (Campes and Tilghman (1989) Genes Dev. 3:537-546).

within a cell line or microorganism may be modified by inserting a heterologous DNA regulatory element into the genome of a stable cell line or cloned microorganism such that the inserted regulatory element is operatively linked with the endogenous hVR-1, hVR-2, and rVR-2 gene. For example, an endogenous hVR-1, hVR-2, and rVR-2 gene which is normally "trancriptionally silent", *i.e.*, a hVR-1, hVR-2, and rVR-2 gene which is normally not expressed, or is expressed only at very low levels in a cell line or microorganism, may be activated by inserting a regulatory element which is capable of promoting the expression of a normally expressed gene product in that cell line or microorganism. Alternatively, a transcriptionally silent, endogenous hVR-1, hVR-2, and rVR-2 gene, may be activated by insertion of a promiscuous regulatory element that works across cell types.

- 47 -

A heterologous regulatory element may be inserted into a stable cell line or cloned microorganism, such that it is operatively linked with an endogenous hVR-1, hVR-2, and rVR-2 gene, using techniques, such as targeted homologous recombination, which are well known to those of skill in the art, and described *e.g.*, in Chappel, U.S. Patent No.: 5,272,071; PCT publication No. WO 91/06667, published May 16, 1991.

The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to hVR-1, hVR-2, and rVR-2 mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub, H. et al., Antisense RNA as a molecular tool for genetic analysis, Reviews - Trends in Genetics, Vol. 1(1) 1986.

Another aspect of the invention pertains to host cells into which an hVR-1, hVR-2, and rVR-2 nucleic acid molecule of the invention is introduced, e.g., an hVR-1, hVR-2, and rVR-2 nucleic acid molecule within a recombinant expression vector or an hVR-1, hVR-2, and rVR-2 nucleic acid molecule containing sequences which allow it to homologously recombine into a specific site of the host cell's genome. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not,

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- 48 -

in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, an hVR-1, hVR-2, and rVR-2 protein can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding an hVR-1, hVR-2, and rVR-2 protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) an hVR-1, hVR-2, and rVR-2 protein. Accordingly, the invention further provides methods for producing an hVR-1, hVR-2, and rVR-2 protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression

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- 49 -

vector encoding an hVR-1, hVR-2, and rVR-2 protein has been introduced) in a suitable medium such that an hVR-1, hVR-2, and rVR-2 protein is produced. In another embodiment, the method further comprises isolating an hVR-1, hVR-2, and rVR-2 protein from the medium or the host cell.

The host cells of the invention can also be used to produce non-human transgenic animals. For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which hVR-1, hVR-2, and rVR-2-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous hVR-1, hVR-2, and rVR-2 sequences have been introduced into their genome or homologous recombinant animals in which endogenous hVR-1, hVR-2, and rVR-2 sequences have been altered. Such animals are useful for studying the function and/or activity of an hVR-1, hVR-2, and rVR-2 and for identifying and/or evaluating modulators of hVR-1, hVR-2, and rVR-2 activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous hVR-1, hVR-2, and rVR-2 gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing an hVR-1, hVR-2, and rVR-2-encoding nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The hVR-1, hVR-2, and rVR-2 cDNA sequence of SEQ ID NO:1, 3, 5, 7 or 9 can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of a hVR-2 gene, such as a

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- 50 -

mouse or rat hVR-2, e.g., the rVR-2 gene, can be used as a transgene. Alternatively, an hVR-1, hVR-2, and rVR-2 gene homologue, such as another member of the Capsaicin/Vanilloid family, can be isolated based on hybridization to the hVR-1, hVR-2, and rVR-2 cDNA sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, (described further in subsection I above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to an hVR-1, hVR-2, and rVR-2 transgene to direct expression of an hVR-1, hVR-2, and rVR-2 protein to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., Manipulating the Mouse Embryo, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of an hVR-1, hVR-2, and rVR-2 transgene in its genome and/or expression of hVR-1, hVR-2, and rVR-2 mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding an hVR-1, hVR-2, and rVR-2 protein can further be bred to other transgenic animals carrying other transgenes.

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of an hVR-1, hVR-2, and rVR-2 gene into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, functionally disrupt, the hVR-1, hVR-2, and rVR-2 gene. The VR-1 or VR-2 gene can be a human gene (*e.g.*, the cDNA of SEQ ID NO:1, 3, 5, 4, 6, 7, or 9), but more preferably, is a non-human homologue of a hVR-1 and hVR-2 gene (*e.g.*, the cDNA of SEQ ID NO:10 or 12, or a cDNA isolated by stringent hybridization with the nucleotide sequence of SEQ ID NO: 1, 3, 5, 4, 6, 7, or 9). For example, a mouse VR-2 gene can be used to construct a homologous recombination nucleic acid molecule, *e.g.*, a vector, suitable for altering an endogenous VR-2 gene in the mouse genome. In a embodiment, the homologous recombination nucleic acid molecule is designed such that, upon homologous recombination, the

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- 51 -

endogenous hVR-1, hVR-2, and rVR-2 gene is functionally disrupted (i.e., no longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the homologous recombination nucleic acid molecule can be designed such that, upon homologous recombination, the endogenous hVR-1, hVR-2, and rVR-2 gene is mutated or otherwise altered but still encodes functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous hVR-1, hVR-2, and rVR-2 protein). In the homologous recombination nucleic acid molecule, the altered portion of the hVR-1, hVR-2, and rVR-2 gene is flanked at its 5' and 3' ends by additional nucleic acid sequence of the hVR-1, hVR-2, and rVR-2 gene to allow for homologous recombination to occur between the exogenous hVR-1, hVR-2, and rVR-2 10 gene carried by the homologous recombination nucleic acid molecule and an endogenous hVR-1, hVR-2, and rVR-2 gene in a cell, e.g., an embryonic stem cell. The additional flanking hVR-1, hVR-2, and rVR-2 nucleic acid sequence is of sufficient length for successful homologous recombination with the endogenous gene. Typically, 15 several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the homologous recombination nucleic acid molecule (see, e.g., Thomas, K.R. and Capecchi, M. R. (1987) Cell 51:503 for a description of homologous recombination vectors). The homologous recombination nucleic acid molecule is introduced into a cell, e.g., an embryonic stem cell line (e.g., by electroporation) and cells in which the 20 introduced hVR-1, hVR-2, and rVR-2 gene has homologously recombined with the endogenous hVR-1, hVR-2, and rVR-2 gene are selected (see e.g., Li, E. et al. (1992) Cell 69:915). The selected cells can then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see e.g., Bradley, A. in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E.J. Robertson, ed. (IRL, Oxford, 1987) pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant 25 female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination 30 nucleic acid molecules, e.g., vectors, or homologous recombinant animals are described further in Bradley, A. (1991) Current Opinion in Biotechnology 2:823-829 and in PCT

International Publication Nos.: WO 90/11354 by Le Mouellec et al.; WO 91/01140 by Smithies et al.; WO 92/0968 by Zijlstra et al.; and WO 93/04169 by Berns et al.

In another embodiment, transgenic non-humans animals can be produced which contain selected systems which allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, *e.g.*, Lakso *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.* (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. (1997) Nature 385:810-813 and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G₀ phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The recontructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

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IV. Pharmaceutical Compositions

The hVR-1, hVR-2, and rVR-2 nucleic acid molecules, fragments of hVR-1, hVR-2, and rVR-2 proteins, and anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically

- 53 -

acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the

25 extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor ELTM (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for

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example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyetheylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a fragment of an hVR-1, hVR-2, and rVR-2 protein or an anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or

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- 55 -

lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will

protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems.

Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid.

Methods for preparation of such formulations will be apparent to those skilled in the art.

The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

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- 56 -

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (*i.e.*, the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

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As defined herein, a therapeutically effective amount of protein or polypeptide (i.e., an effective dosage) ranges from about 0.001 to 30 mg/kg body weight, preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight. The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments. In a preferred example, a subject is treated with antibody, protein, or polypeptide in the range of between about 0.1 to 20 mg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of antibody, protein, or polypeptide used for treatment may increase or decrease over the course of a particular treatment. Changes in dosage may result and become apparent from the results of diagnostic assays as described herein.

The present invention encompasses agents which modulate expression or activity. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (i.e., including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds.

It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon

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- 58 -

the identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention.

Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein.

When one or more of these small molecules is to be administered to an animal (e.g., a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, gender, and diet of the subject, the time of administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Patent 5,328,470) or by stereotactic injection (see e.g., Chen et al. (1994) Proc. Natl. Acad. Sci. USA 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, e.g., retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

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- 59 -

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

V. Uses and Methods of the Invention

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The nucleic acid molecules, proteins, protein homologues, and antibodies described herein can be used in one or more of the following methods: a) screening assays; b) predictive medicine (*e.g.*, diagnostic assays, prognostic assays, monitoring clinical trials, and pharmacogenetics); and c) methods of treatment (*e.g.*, therapeutic and prophylactic). As described herein, an hVR-1, hVR-2, and rVR-2 protein of the invention has one or more of the following activities: (1) it interacts with a non-hVR-1, non-hVR-2, and non-rVR-2 protein molecule, *e.g.*, a vanilloid compound such as capsaicin; (2) it modulates intracellular calcium concentration; (3) it activates an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; and (4) it modulates a pain signaling mechanism, and, thus, can be used to, for example, (1) modulate the interaction with a non-hVR-1, non-hVR-2, and non-rVR-2 protein molecule; (2) modulate intracellular calcium concentration; (3) activate an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; and (4) modulate a pain signaling mechanism.

The isolated nucleic acid molecules of the invention can be used, for example, to express hVR-1, hVR-2, and rVR-2 protein (*e.g.*, via a recombinant expression vector in a host cell in gene therapy applications), to detect hVR-1, hVR-2, and rVR-2 mRNA (*e.g.*, in a biological sample) or a genetic alteration in an hVR-1, hVR-2, and rVR-2 gene, and to modulate hVR-1, hVR-2, and rVR-2 activity, as described further below. The hVR-1, hVR-2, and rVR-2 proteins can be used to screen for naturally occurring hVR-1, hVR-2, and rVR-2 substrates, to screen for drugs or compounds which modulate hVR-1, hVR-2, and rVR-2 activity, as well as to treat disorders characterized by insufficient or excessive production of hVR-1, hVR-2, and rVR-2 protein forms which have decreased or aberrant activity compared to hVR-1, hVR-2, and rVR-2 wild type protein (*e.g.*, pain disorders). Moreover, the anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies of the invention can be used to detect and isolate hVR-1, hVR-2, and rVR-2 proteins, regulate the

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bioavailability of hVR-1, hVR-2, and rVR-2 proteins, and modulate hVR-1, hVR-2, and rVR-2 activity.

A. Screening Assays:

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The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, peptides, peptidomimetics, small molecules or other drugs) which bind to hVR-1, hVR-2, and rVR-2 proteins, have a stimulatory or inhibitory effect on, for example, hVR-1, hVR-2, and rVR-2 expression or hVR-1, hVR-2, and rVR-2 activity, or have a stimulatory or inhibitory effect on, for example, the expression or activity of hVR-1, hVR-2, and rVR-2 substrate.

In one embodiment, the invention provides assays for screening candidate or test compounds which are substrates of an hVR-1, hVR-2, and rVR-2 protein or polypeptide or biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of an hVR-1, hVR-2, and rVR-2 protein or polypeptide or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:6909; Erb et al. (1994) Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al. (1994). J. Med. Chem. 37:2678; Cho et al. (1993) Science 261:1303; Carrell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2061; and in Gallop et al. (1994) J. Med. Chem. 37:1233.

- 61 -

Libraries of compounds may be presented in solution (e.g., Houghten (1992) Biotechniques 13:412-421), or on beads (Lam (1991) Nature 354:82-84), chips (Fodor (1993) Nature 364:555-556), bacteria (Ladner USP 5,223,409), spores (Ladner USP '409), plasmids (Cull et al. (1992) Proc Natl Acad Sci USA 89:1865-1869) or on phage (Scott and Smith (1990) Science 249:386-390); (Devlin (1990) Science 249:404-406); (Cwirla et al. (1990) Proc. Natl. Acad. Sci. 87:6378-6382); (Felici (1991) J. Mol. Biol. 222:301-310); (Ladner supra.).

In one embodiment, an assay is a cell-based assay in which a cell, e.g., a neuronal cell, which expresses an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity is determined. Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity can be accomplished by monitoring, for example, intracellular calcium concentration or membrane depolarization by, e.g., patch-clamp recordings in whole-cell, inside-out, and outside-out configurations (as described in, for example, Tominaga M. et al. (1998) Neuron 21:531-543). Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity can further be accomplished by monitoring the activity of an hVR-1, hVR-2, and rVR-2-regulated transcription factor. The cell, for example, can be of mammalian origin, e.g., a neuronal cell.

The ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 binding to a substrate or to bind to hVR-1, hVR-2, and rVR-2 can also be determined.

Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 binding to a substrate can be accomplished, for example, by coupling the hVR-1, hVR-2, and rVR-2 substrate with a radioisotope or enzymatic label such that binding of the hVR-1, hVR-2, and rVR-2 substrate to hVR-1, hVR-2, and rVR-2 can be determined by detecting the labeled hVR-1, hVR-2, and rVR-2 substrate in a complex. Determining the ability of the test compound to bind hVR-1, hVR-2, and rVR-2 can be accomplished, for example, by coupling the compound with a radioisotope or enzymatic label such that binding of the compound to hVR-1, hVR-2, and rVR-2 can be determined by detecting the labeled hVR-1, hVR-2, and rVR-2 compound in a complex. For example, compounds (e.g., hVR-1, hVR-2, and rVR-2 substrates) can be labeled with 1251, 358.

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- 62 -

¹⁴C, or ³H, either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a compound (e.g., an hVR-1, hVR-2, and rVR-2 substrate) to interact with hVR-1, hVR-2, and rVR-2 without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of a compound with hVR-1, hVR-2, and rVR-2 without the labeling of either the compound or the hVR-1, hVR-2, and rVR-2. McConnell, H. M. et al. (1992) Science 257:1906-1912. As used herein, a "microphysiometer" (e.g., Cytosensor) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between a compound and hVR-1, hVR-2, and rVR-2.

In yet another embodiment, an assay of the present invention is a cell-free assay in which an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is determined. biologically active portions of the hVR-1, hVR-2, and rVR-2 proteins to be used in 20 assays of the present invention include fragments which participate in interactions with non-hVR-1, non-hVR-2, and non-rVR-2 molecules, e.g., fragments with high surface probability scores. Binding of the test compound to the hVR-1, hVR-2, and rVR-2 protein can be determined either directly or indirectly as described above. In a embodiment, the assay includes contacting the hVR-1, hVR-2, and rVR-2 protein or 25 biologically active portion thereof with a known compound which binds hVR-1, hVR-2, and rVR-2 to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with an hVR-1, hVR-2, and rVR-2 protein, wherein determining the ability of the test compound to interact with an hVR-1, hVR-2, and rVR-2 protein comprises determining the ability of the test 30

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- 63 -

compound to preferentially bind to hVR-1, hVR-2, and rVR-2 or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate (e.g., stimulate or inhibit) the activity of the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is determined. Determining the ability of the test compound to modulate the activity of an hVR-1, hVR-2, and rVR-2 protein can be accomplished, for example, by determining the ability of the hVR-1, hVR-2, and rVR-2 protein to bind to an hVR-1, 10 hVR-2, and rVR-2 target molecule, e.g., a vanilloid compound such as capsaicin, by one of the methods described above for determining direct binding. Determining the ability of the hVR-1, hVR-2, and rVR-2 protein to bind to an hVR-1, hVR-2, and rVR-2 target molecule can also be accomplished using a technology such as real-time Biomolecular Interaction Analysis (BIA). Sjolander, S. and Urbaniczky, C. (1991) Anal. Chem. 15 63:2338-2345 and Szabo et al. (1995) Curr. Opin. Struct. Biol. 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (e.g., BIAcore). Changes in the optical phenomenon of surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

In an alternative embodiment, determining the ability of the test compound to modulate the activity of an hVR-1, hVR-2, and rVR-2 protein can be accomplished by determining the ability of the hVR-1, hVR-2, and rVR-2 protein to further modulate the activity of a downstream effector of an hVR-1, hVR-2, and rVR-2 target molecule. For example, the activity of the effector molecule on an appropriate target can be determined or the binding of the effector to an appropriate target can be determined as previously described.

In yet another embodiment, the cell-free assay involves contacting an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof with a known compound which binds the hVR-1, hVR-2, and rVR-2 protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the hVR-1, hVR-2, and rVR-2 protein, wherein

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- 64 -

determining the ability of the test compound to interact with the hVR-1, hVR-2, and rVR-2 protein comprises determining the ability of the hVR-1, hVR-2, and rVR-2 protein to preferentially bind to or modulate the activity of an hVR-1, hVR-2, and rVR-2 target molecule.

The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of isolated proteins (*e.g.*, hVR-1, hVR-2, and rVR-2 proteins or biologically active portions thereof). In the case of cell-free assays in which a membrane-bound form of an isolated protein is used it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the isolated protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPS), 3-[(3-cholamidopropyl)dimethylamminio]-2-hydroxy-1-propane sulfonate (CHAPSO), or N-dodecyl=N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either hVR-1, hVR-2, and rVR-2 or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to an hVR-1, hVR-2, and rVR-2 protein, or interaction of an hVR-1, hVR-2, and rVR-2 protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and microcentrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase/ hVR-1, hVR-2, and rVR-2 fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or hVR-1, hVR-2, and rVR-2 protein, and the mixture incubated under

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- 65 -

conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of hVR-1, hVR-2, and rVR-2 binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either an hVR-1, hVR-2, and rVR-2 protein or an hVR-1, hVR-2, and rVR-2 target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated hVR-1, hVR-2, and rVR-2 protein 10 or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with hVR-1, hVR-2, and rVR-2 protein or target molecules but which do not interfere with binding of the hVR-1, hVR-2, and rVR-2 protein to its target molecule can be derivatized to the wells of the plate, and unbound target or hVR-1, hVR-2, and rVR-2 protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the hVR-1, hVR-2, and rVR-2 protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the hVR-1, hVR-2, and rVR-2 protein or target molecule.

In another embodiment, modulators of hVR-1, hVR-2, and rVR-2 expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the cell is determined. The level of expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the presence of the candidate compound is compared to the level of expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of hVR-1, hVR-2, and rVR-2 expression based on this comparison. For example, when expression of hVR-1, hVR-2, and rVR-2 mRNA or protein is greater (statistically significantly greater) in the presence

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- 66 -

of the candidate compound than in its absence, the candidate compound is identified as a stimulator of hVR-1, hVR-2, and rVR-2 mRNA or protein expression. Alternatively, when expression of hVR-1, hVR-2, and rVR-2 mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of hVR-1, hVR-2, and rVR-2 mRNA or protein expression. The level of hVR-1, hVR-2, and rVR-2 mRNA or protein expression in the cells can be determined by methods described herein for detecting hVR-1, hVR-2, and rVR-2 mRNA or protein.

In yet another aspect of the invention, the hVR-1, hVR-2, and rVR-2 proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. 10 Patent No. 5,283,317; Zervos et al. (1993) Cell 72:223-232; Madura et al. (1993) J. Biol. Chem. 268:12046-12054; Bartel et al. (1993) Biotechniques 14:920-924; Iwabuchi et al. (1993) Oncogene 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with hVR-1, hVR-2, and rVR-2 ("hVR-1-binding 15 proteins", "hVR-2-binding proteins", and "rVR-2-binding proteins" or "hVR-1-bp", "hVR-2-bp", and "rVR-2-bp") and are involved in hVR-1, hVR-2, and rVR-2 activity. Such hVR-1, hVR-2, and rVR-2-binding proteins are also likely to be involved in the propagation of signals by the hVR-1, hVR-2, and rVR-2 proteins or hVR-1, hVR-2, and rVR-2 targets as, for example, downstream elements of an hVR-1, hVR-2, and rVR-2mediated signaling pathway, e.g., a pain signaling pathway. Alternatively, such hVR-1, 20 hVR-2, and rVR-2-binding proteins are likely to be hVR-1, hVR-2, and rVR-2 inhibitors.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for an hVR-1, hVR-2, and rVR-2 protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, in vivo, forming an hVR-1, hVR-2, and rVR-2-dependent complex, the DNA-binding and

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- 67 -

activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the hVR-1, hVR-2, and rVR-2 protein.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (e.g., an hVR-1, hVR-2, and rVR-2 modulating agent, an antisense hVR-1, hVR-2, and rVR-2 nucleic acid molecule, an hVR-1, hVR-2, and rVR-2-specific antibody, or an hVR-1, hVR-2, and rVR-2-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

B. Detection Assays

Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue typing); and (iii) aid in forensic identification of a biological sample. These applications are described in the subsections below.

1. Chromosome Mapping

Once the sequence (or a portion of the sequence) of a gene has been isolated, this sequence can be used to map the location of the gene on a chromosome. This process is called chromosome mapping. Accordingly, portions or fragments of the hVR-1, hVR-2,

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and rVR-2 nucleotide sequences, described herein, can be used to map the location of the hVR-1, hVR-2, and rVR-2 genes on a chromosome. The mapping of the hVR-1, hVR-2, and rVR-2 sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease.

Briefly, hVR-1, hVR-2, and rVR-2 genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the hVR-1, hVR-2, and rVR-2 nucleotide sequences. Computer analysis of the hVR-1, hVR-2, and rVR-2 sequences can be used to predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the hVR-1, hVR-2, and rVR-2 sequences will yield an amplified fragment.

Somatic cell hybrids are prepared by fusing somatic cells from different mammals (e.g., human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of individual genes to specific human chromosomes. (D'Eustachio P. et al. (1983) Science 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycler. Using the hVR-1, hVR-2, and rVR-2 nucleotide sequences to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can similarly be used to map an hVR-1, hVR-2, and rVR-2 sequence to its chromosome include *in situ* hybridization (described in Fan, Y. *et al.* (1990) *Proc. Natl. Acad. Sci.*

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- 69 -

USA, 87:6223-27), pre-screening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a DNA sequence to a metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a DNA sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good results at a reasonable amount of time. For a review of this technique, see Verma *et al.*, Human Chromosomes: A Manual of Basic Techniques (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, Mendelian Inheritance in Man, available on-line through Johns Hopkins University Welch Medical Library). The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland, J. et al. (1987) Nature, 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the hVR-1, hVR-2, and rVR-2 gene, can be determined. If a mutation is observed in some or all of the affected individuals but not

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- 70 -

in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

2. Tissue Typing

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The hVR-1, hVR-2, and rVR-2 sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an alternative technique which determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The hVR-1, hVR-2, and rVR-2 nucleotide sequences of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding

- 71 -

regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals.

If a panel of reagents from hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

3. Use of Partial hVR-1, hVR-2, and rVR-2 Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means for positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, *e.g.*, hair or skin, or body fluids, *e.g.*, blood, saliva, or semen found at a crime scene. The amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

The sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e. another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Examples of polynucleotide reagents include the hVR-1, hVR-2, and rVR-2 nucleotide sequences or portions thereof, e.g., fragments derived from SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 11 having a length of at least 20 bases, preferably at least 30 bases.

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- 72 -

The hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein can further be used to provide polynucleotide reagents, e.g., labeled or labelable probes which can be used in, for example, an *in situ* hybridization technique, to identify a specific tissue, e.g., brain tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such hVR-1, hVR-2, and rVR-2 probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, e.g., hVR-1, hVR-2, and rVR-2 primers or probes can be used to screen tissue culture for contamination (i.e. screen for the presence of a mixture of different types of cells in a culture).

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C. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining hVR-1, hVR-2, and rVR-2 protein and/or nucleic acid expression as well as hVR-1, hVR-2, and rVR-2 activity, in the context of a biological sample (e.g., blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. The invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with hVR-1, hVR-2, and rVR-2 protein, nucleic acid expression or activity. For example, mutations in an hVR-1, hVR-2, and rVR-2 gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby phophylactically treat an individual prior to the onset of a disorder characterized by or associated with hVR-1, hVR-2, and rVR-2 protein, nucleic acid expression or activity.

Another aspect of the invention pertains to monitoring the influence of agents (e.g., drugs, compounds) on the expression or activity of hVR-1, hVR-2, and rVR-2 in clinical trials.

These and other agents are described in further detail in the following sections.

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting hVR-1, hVR-2, and rVR-2 protein or nucleic acid (*e.g.*, mRNA, genomic DNA) that encodes hVR-1, hVR-2, and rVR-2 protein such that the presence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid is detected in the biological sample. A agent for detecting hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA. The nucleic acid probe can be, for example, a full-length hVR-1, hVR-2, and rVR-2 nucleic acid, such as the nucleic acid of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

An agent for detecting hVR-1, hVR-2, and rVR-2 protein is an antibody capable of binding to hVR-1, hVR-2, and rVR-2 protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')2) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect hVR-1, hVR-2, and rVR-2 mRNA, protein, or genomic DNA in a biological sample in vitro as well as in vivo. For example, in vitro techniques for detection of hVR-1, hVR-2, and rVR-2 mRNA include Northern hybridizations and in situ hybridizations. In vitro

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techniques for detection of hVR-1, hVR-2, and rVR-2 protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of hVR-1, hVR-2, and rVR-2 genomic DNA include Southern hybridizations. Furthermore, *in vivo* techniques for detection of hVR-1, hVR-2, and rVR-2 protein include introducing into a subject a labeled anti-hVR-1, hVR-2, and rVR-2 antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A biological sample is a serum sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA, such that the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA in the control sample with the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting the presence of hVR-1, hVR-2, and rVR-2 in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting hVR-1, hVR-2, and rVR-2 protein or mRNA in a biological sample; means for determining the amount of hVR-1, hVR-2, and rVR-2 in the sample; and means for comparing the amount of hVR-1, hVR-2, and rVR-2 in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect hVR-1, hVR-2, and rVR-2 protein or nucleic acid.

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- 75 -

2. Prognostic Assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. As used herein, the term "aberrant" includes an hVR-1, hVR-2, and rVR-2 expression or activity which deviates from the wild type hVR-1, hVR-2, and rVR-2 expression or activity. Aberrant expression or activity includes increased or decreased expression or activity, as well as expression or activity which does not follow the wild type developmental pattern of expression or the subcellular pattern of expression. For example, aberrant hVR-1, hVR-2, and rVR-2 expression or activity is intended to include the cases in which a mutation in the hVR-1, hVR-2, and rVR-2 gene causes the hVR-1, hVR-2, and rVR-2 gene to be underexpressed or over-expressed and situations in which such mutations result in a nonfunctional hVR-1, hVR-2, and rVR-2 protein or a protein which does not function in a wild-type fashion, e.g., a protein which does not interact with an hVR-1, hVR-2, and rVR-2 ligand or one which interacts with a non-hVR-1, non-hVR-2, and non-rVR-2 ligand.

The assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing a disorder associated with a misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a pain disorder. Alternatively, the prognostic assays can be utilized to identify a subject having or at risk for developing a disorder associated with a misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a pain disorder. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity in which a test sample is obtained from a subject and hVR-1, hVR-2, and rVR-2 protein or nucleic acid (e.g., mRNA or genomic DNA) is detected, wherein the presence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest. For example, a test sample can be a biological fluid (e.g., serum), cell sample, or tissue.

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- 76 -

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. For example, such methods can be used to determine whether a subject can be effectively treated with an agent for a pain disorder. Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity in which a test sample is obtained and hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity is detected (*e.g.*, wherein the abundance of hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity).

The methods of the invention can also be used to detect genetic alterations in an hVR-1, hVR-2, and rVR-2 gene, thereby determining if a subject with the altered gene is 15 at risk for a disorder characterized by misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a neurodegenerative disorder. In embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding an hVR-1, hVR-2, and rVR-2-protein, or the 20 mis-expression of the hVR-1, hVR-2, and rVR-2 gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of 1) a deletion of one or more nucleotides from an hVR-1, hVR-2, and rVR-2 gene; 2) an addition of one or more nucleotides to an hVR-1, hVR-2, and rVR-2 gene; 3) a substitution of one or more nucleotides of an hVR-1, hVR-2, and rVR-2 gene, 4) a chromosomal 25 rearrangement of an hVR-1, hVR-2, and rVR-2 gene; 5) an alteration in the level of a messenger RNA transcript of an hVR-1, hVR-2, and rVR-2 gene, 6) aberrant modification of an hVR-1, hVR-2, and rVR-2 gene, such as of the methylation pattern of the genomic DNA, 7) the presence of a non-wild type splicing pattern of a messenger RNA transcript of an hVR-1, hVR-2, and rVR-2 gene, 8) a non-wild type level of an 30 hVR-1, hVR-2, and rVR-2-protein, 9) allelic loss of an hVR-1, hVR-2, and rVR-2 gene,

- 77 -

and 10) inappropriate post-translational modification of an hVR-1, hVR-2, and rVR-2-protein. As described herein, there are a large number of assays known in the art which can be used for detecting alterations in an hVR-1, hVR-2, and rVR-2 gene. A biological sample is a tissue or serum sample isolated by conventional means from a subject.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al. (1988) Science 241:1077-1080; and Nakazawa et al. (1994) Proc. Natl. Acad. Sci. USA 91:360-364), the latter of which can be particularly useful for detecting point mutations in the hVR-1, hVR-2, and rVR-2-gene (see Abravaya et al. (1995) Nucleic Acids Res .23:675-682). This method can include the steps of collecting a sample of cells from a subject, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to an hVR-1, hVR-2, and rVR-2 gene under conditions such that hybridization and amplification of the hVR-1, hVR-2, and rVR-2-gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. et al., (1990) Proc. Natl. Acad. Sci. USA 87:1874-1878), transcriptional amplification system (Kwoh, D.Y. et al., (1989) Proc. Natl. Acad. Sci. USA 86:1173-1177), Q-Beta Replicase (Lizardi, P.M. et al. (1988) Bio-Technology 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in an hVR-1, hVR-2, and rVR-2 gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally),

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- 78 -

digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Patent No. 5,498,531) can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in hVR-1, hVR-2, and rVR-2 can be identified by hybridizing a sample and control nucleic acids, e.g., DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin, M.T. et al. (1996) Human Mutation 7: 244-255; Kozal, M.J. et al. (1996) Nature Medicine 2: 753-759). For example, genetic mutations in hVR-1, hVR-2, and rVR-2 can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. et al. supra. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping 15 probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene. 20

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the hVR-1, hVR-2, and rVR-2 gene and detect mutations by comparing the sequence of the sample hVR-1, hVR-2, and rVR-2 with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on techniques developed by Maxam and Gilbert ((1977) *Proc. Natl. Acad. Sci. USA* 74:560) or Sanger ((1977) *Proc. Natl. Acad. Sci. USA* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, *e.g.*, PCT International Publication No. WO 94/16101; Cohen *et al.* (1996) *Adv. Chromatogr.* 36:127-162; and Griffin *et al.* (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

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- 79 -

Other methods for detecting mutations in the hVR-1, hVR-2, and rVR-2 gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers et al. (1985) Science 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wildtype hVR-1, hVR-2, and rVR-2 sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent which cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton et al. (1988) Proc. Natl Acad Sci USA 85:4397; Saleeba et al. (1992) Methods Enzymol. 217:286-295. In a embodiment, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in hVR-1, hVR-2, and rVR-2 cDNAs obtained from samples of cells. For example, the mutY enzyme of E. coli cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu et al. (1994) Carcinogenesis 15:1657-1662). According to an exemplary embodiment, a probe based on an hVR-1, hVR-2, and rVR-2 sequence, e.g., a wild-type hVR-1, hVR-2, and rVR-2 sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

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- 80 -

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in hVR-1, hVR-2, and rVR-2 genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (orita et al. (1989) Proc Natl. Acad. Sci USA: 86:2766, see also Cotton (1993) Mutat. Res. 285:125-144; and Hayashi (1992) Genet. Anal. Tech. Appl. 9:73-79). Single-stranded DNA fragments of sample and control hVR-1, hVR-2, and rVR-2 nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In a embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen et al. (1991) Trends Genet 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers et al. (1985) Nature 313:495). When DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) Biophys Chem 265:12753).

Examples of other techniques for detecting point mutations include, but are not
25 limited to, selective oligonucleotide hybridization, selective amplification, or selective
primer extension. For example, oligonucleotide primers may be prepared in which the
known mutation is placed centrally and then hybridized to target DNA under conditions
which permit hybridization only if a perfect match is found (Saiki et al. (1986) Nature
324:163); Saiki et al. (1989) Proc. Natl Acad. Sci USA 86:6230). Such allele specific
30 oligonucleotides are hybridized to PCR amplified target DNA or a number of different

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- 81 -

mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology which depends on selective PCR amplification may be used in conjunction with the instant invention.

Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs et al. (1989) Nucleic Acids Res. 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner (1993) Tibtech 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini et al. (1992) Mol. Cell Probes 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) Proc. Natl. Acad. Sci USA 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing prepackaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving an hVR-1, hVR-2, and rVR-2 gene.

Furthermore, any cell type or tissue in which hVR-1, hVR-2, and rVR-2 is expressed may be utilized in the prognostic assays described herein.

25 3. Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (e.g., drugs) on the expression or activity of an hVR-1, hVR-2, and rVR-2 protein can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or upregulate hVR-1, hVR-2, and rVR-2 activity, can be monitored in clinical trials of subjects exhibiting decreased hVR-1, hVR-2, and rVR-2

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gene expression, protein levels, or downregulated hVR-1, hVR-2, and rVR-2 activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or downregulate hVR-1, hVR-2, and rVR-2 activity, can be monitored in clinical trials of subjects exhibiting increased hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or upregulated hVR-1, hVR-2, and rVR-2 activity. In such clinical trials, the expression or activity of an hVR-1, hVR-2, and rVR-2 gene, and preferably, other genes that have been implicated in, for example, an hVR-1, hVR-2, and rVR-2-associated disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes, including hVR-1, hVR-2, and rVR-2, that are modulated in cells by treatment with an agent (e.g., compound, drug or small molecule) which modulates hVR-1, hVR-2, and rVR-2 activity (e.g., identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on hVR-1, hVR-2, and rVR-2-associated disorders (e.g., pain disorders), for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of hVR-1, hVR-2, and rVR-2 and other genes implicated in the hVR-1, hVR-2, and rVR-2-associated disorder, respectively. The levels of gene expression (e.g., a gene expression pattern) can be quantified by northern blot analysis or RT-PCR, as described herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of hVR-1, hVR-2, and rVR-2 or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be determined before, and at various points during treatment of the individual with the agent.

In a embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) including the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of an hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the preadministration sample; (iii) obtaining one or more post-

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- 83 -

administration samples from the subject; (iv) detecting the level of expression or activity of the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the pre-administration sample with the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of hVR-1, hVR-2, and rVR-2 to higher levels than detected, *i.e.*, to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of hVR-1, hVR-2, and rVR-2 to lower levels than detected, *i.e.* to decrease the effectiveness of the agent. According to such an embodiment, hVR-1, hVR-2, and rVR-2 expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

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D. Methods of Treatment:

The present invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. With regards to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics. "Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market. More specifically, the term refers the study of how a patient's genes determine his or her response to a drug (e.g., a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with either the hVR-1, hVR-2, and rVR-2 molecules of the present invention or hVR-1, hVR-2, and rVR-2 modulators according to that individual's drug response genotype. Pharmacogenomics allows a clinician or physician to target prophylactic or

- 84 -

PCT/US99/26701

therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug-related side effects.

1. Prophylactic Methods

In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with an aberrant hVR-1, hVR-2, and rVR-2 expression or activity, by administering to the subject an hVR-1, hVR-2, and rVR-2 or an agent which modulates hVR-1, hVR-2, and rVR-2 expression or at least one hVR-1, hVR-2, and rVR-2 activity. Subjects at risk for a disease which is caused or contributed to by aberrant hVR-1, hVR-2, and rVR-2 expression or activity can be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the hVR-1, hVR-2, and rVR-2 aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of hVR-1, hVR-2, and rVR-2 aberrancy, for example, an hVR-1, hVR-2, and rVR-2, hVR-1, hVR-2, and rVR-2 agonist or hVR-1, hVR-2, and rVR-2 antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

20 2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating hVR-1, hVR-2, and rVR-2 expression or activity for therapeutic purposes. Accordingly, in an exemplary embodiment, the modulatory method of the invention involves contacting a cell with an hVR-1, hVR-2, and rVR-2 or agent that modulates one or more of the activities of hVR-1, hVR-2, and rVR-2 protein activity associated with the cell. An agent that modulates hVR-1, hVR-2, and rVR-2 protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of an hVR-1, hVR-2, and rVR-2 protein (e.g., an hVR-1, hVR-2, and rVR-2 substrate), an hVR-1, hVR-2, and rVR-2 antibody, an hVR-1, hVR-2, and rVR-2 agonist or antagonist, a peptidomimetic of an hVR-1, hVR-2, and rVR-2 agonist or antagonist, or other small molecule. In one embodiment, the agent stimulates one or more hVR-1,

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hVR-2, and rVR-2 activities. Examples of such stimulatory agents include active hVR-1, hVR-2, and rVR-2 protein and a nucleic acid molecule encoding hVR-1, hVR-2, and rVR-2 that has been introduced into the cell. In another embodiment, the agent inhibits one or more hVR-1, hVR-2, and rVR-2 activities. Examples of such inhibitory agents include antisense hVR-1, hVR-2, and rVR-2 nucleic acid molecules, anti-hVR-1, hVR-2, and rVR-2 antibodies, and hVR-1, hVR-2, and rVR-2 inhibitors. These modulatory methods can be performed in vitro (e.g., by culturing the cell with the agent) or, alternatively, in vivo (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of an hVR-1, hVR-2, and rVR-2 protein or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) hVR-1, hVR-2, and rVR-2 expression or activity. In another embodiment, the method involves administering an hVR-1, hVR-2, and rVR-2 protein or nucleic acid molecule as therapy to compensate for reduced or aberrant hVR-1, hVR-2, and rVR-2 expression or activity.

Stimulation of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which hVR-1, hVR-2, and rVR-2 is abnormally downregulated and/or in which increased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect. For example, stimulation of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which an hVR-1, hVR-2, and rVR-2 is downregulated and/or in which increased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect. Likewise, inhibition of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which hVR-1, hVR-2, and rVR-2 is abnormally upregulated and/or in which decreased hVR-1, hVR-2, and rVR-2 is abnormally upregulated and/or in which decreased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect.

3. Pharmacogenomics

The hVR-1, hVR-2, and rVR-2 molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on hVR-1, hVR-2, and rVR-2 activity (e.g., hVR-1, hVR-2, and rVR-2 gene expression) as identified by a screening assay described herein can be administered to individuals to treat

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- 86 -

(prophylactically or therapeutically) hVR-1, hVR-2, and rVR-2-associated disorders (e.g., pain disorders) associated with aberrant hVR-1, hVR-2, and rVR-2 activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator as well as tailoring the dosage and/or therapeutic regimen of treatment with an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, for example, Eichelbaum, M. et al. (1996) Clin. Exp. Pharmacol. Physiol. 23(10-11):983-985 and Linder, M.W. et al. (1997) Clin. Chem. 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (e.g., a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants.) Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically

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- 87 -

significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a "SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict drug response. According to this method, if a gene that encodes a drugs target is known (e.g., an hVR-1, hVR-2, and rVR-2 protein of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a 20 major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (e.g., N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive 25 metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently 30 experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as

- 88 -

demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (e.g., an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator of the present invention) can give an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator, such as a modulator identified by one of the exemplary screening assays described herein.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the Figures and the Sequence Listing are incorporated herein by reference.

EXAMPLES

25 EXAMPLE 1: IDENTIFICATION AND CHARACTERIZATION OF hVR-1, hVR-2, and rVR-2 cDNA

In this example, the identification and characterization of the genes encoding hVR-1 (clone Fchrb87a6), hVR-2 (clone flh21e11), hVR-2 alternate form (clone frhob12c4), and rVR-2 (clone flrxb147g11) are described.

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Isolation of the hVR-1, hVR-2, and the rVR-2 cDNA

The invention is based, at least in part, on the discovery of two human genes and one rat gene encoding novel members of the Capsaicin/Vanilloid receptor family, referred to herein as hVR-1, hVR-2, and rVR-2, respectively. These clones were identified from a human heart library and a rat dorsal root ganglion (DRG) library, based on sequence homology to the known rat VR-1 (Accession Number AF029310). The sequence of the two human clones and the rat clone was determined and found to contain open reading frames.

The nucleotide sequence of the full length hVR-1 cDNA and the predicted amino acid sequence of the hVR-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively.

The nucleotide sequence of the full length hVR-2 cDNA and the predicted amino acid sequence of the hVR-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively.

The nucleotide sequence of the partial hVR-2 (alternate form) cDNA and the predicted amino acid sequence of the hVR-2 (alternate form) polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively.

The amino acid sequence of the predicted full length human VR-2 protein (alternate form) is shown in Figure 16 and in SEQ ID NO:20.

The nucleotide sequence of the partial rVR-2 cDNA and the predicted amino acid sequence of the rVR-2 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively.

Analysis of the hVR-1, hVR-2, and rVR-2 Molecules

The hVR-1 protein (SEQ ID NO:2) was aligned with the human VR-2 protein (SEQ ID NO:5) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 46.348% identity and 55.378% similarity between the two sequences (see Figure 5).

The hVR-1 nucleotide sequence (SEQ ID NO:1) was aligned with the human

VR-2 nucleotide sequence (SEQ ID NO:4) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3. The

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results showed a 55.316% identity and 55.316% similarity between the two sequences (see Figure 6).

The hVR-2 protein (SEQ ID NO:5) was aligned with the rat VR-2 protein (SEQ ID NO:11) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 7), as well as using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 79.167% identity and 81.703% similarity between the two sequences (see Figure 8).

The hVR-1 nucleotide sequence (SEQ ID NO:1) was aligned with the rat VR-1 nucleotide sequence (Accession Number:AF029310) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3. The results showed a 82.125% identity and 82.125% similarity between the two sequences (see Figure 9).

The hVR-1 protein (SEQ ID NO:2) was aligned with the rat VR-1 protein (Accession Number:AF029310) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 86.022% identity and 89.247% similarity between the two sequences (see Figure 10).

The hVR-2 protein (SEQ ID NO:5) was aligned with the human VR-2 protein (alternate form) (SEQ ID NO:8) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 11).

Finally, the hVR-2 protein (SEQ ID NO:5) was aligned with the predicted full length human VR-2 protein (alternate form) (SEQ ID NO:20) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 17).

A search was performed against the HMM database resulting in the identification of three ankyrin repeat domains in the amino acid sequence of hVR-1 (SEQ ID NO:2) at about residues 201-233, 248-283, and 333-361, and in the amino acid sequence of hVR-2 (SEQ ID NO:5) at about residues 162-194, 208-243, and 293-328. The results of the searches are set forth in Figures 13 and 15, respectively.

Hydropathy plots have identified 6 transmembrane domains in the hVR-1 and the hVR-2 proteins (see Figures 12 and 14, respectively).

- 91 -

A series of searches have revealed that the hVR-1 protein matches the ProDom entry 141801 for the vanilloid receptor subtype and the ProDom entry 145518 for the vanilloid receptor subtype.

Moreover, a search was performed against the Prosite database resulting in the identification of four N-glycosylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 171-174, 192-195, 604-607, and 749-752), three cGMP-dependent protein kinase phosphorylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 2-5, 368-371, and 499-502), a series of protein kinase C and Casein kinase II phosphorylation sites in the amino acid sequence of SEQ ID NO:5, two tyrosine kinase phosphorylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 368-375 and 622-628), and two myristoylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 169-174 and 765-770).

Tissue Distribution of hVR-1 and hVR-2 mRNA

This Example describes the tissue distribution of hVR-1 and hVR-2 mRNA as determined by *in situ* hybridization.

For *in situ* analysis, tissues, such as brain regions and whole brain, obtained from human and monkey were first frozen on dry ice. Ten-micrometer-thick coronal sections of the tissues were postfixed with 4% formaldehyde in DEPC treated 1X phosphate-buffered saline at room temperature for 10 minutes before being rinsed twice in DEPC 1X phosphate-buffered saline and once in 0.1 M triethanolamine-HCl (pH 8.0). Following incubation in 0.25% acetic anhydride-0.1 M triethanolamine-HCl for 10 minutes, sections were rinsed in DEPC 2X SSC (1X SSC is 0.15M NaCl plus 0.015M sodium citrate). Tissue was then dehydrated through a series of ethanol washes, incubated in 100% chloroform for 5 minutes, and then rinsed in 100% ethanol for 1 minute and 95% ethanol for 1 minute and allowed to air dry.

Hybridizations were performed with ³⁵S-radiolabeled (5 X 10⁷ cpm/ml) cRNA probes. Probes were incubated in the presence of a solution containing 600 mM NaCl, 10 mM Tris (pH 7.5), 1 mM EDTA, 0.01% sheared salmon sperm DNA, 0.01% yeast tRNA, 0.05% yeast total RNA type X1, 1 X Denhardt's solution, 50% formamide, 10%

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- 92 -

dextran sulfate, 100 mM dithiothreitol, 0.1% sodium dodecyl sulfate (SDS), and 0.1% sodium thiosulfate for 18 hours at 55°C.

After hybridization, slides were washed with 2 X SSC. Sections were then sequentially incubated at 37°C in TNE (a solution containing 10 mM Tris-HCl (pH 7.6), 500 mM NaCl, and 1 mM EDTA), for 10 minutes, in TNE with 10µg of RNase A per ml for 30 minutes, and finally in TNE for 10 minutes. Slides were then rinsed with 2 X SSC at room temperature, washed with 2 X SSC at 50°C for 1 hour, washed with 0.2 X SSC at 55°C for 1 hour, and 0.2 X SSC at 60°C for 1 hour. Sections were then dehydrated rapidly through serial ethanol-0.3 M sodium acetate concentrations before being air dried and exposed to Kodak Biomax MR scientific imaging film for 24 hours and subsequently dipped in NB-2 photoemulsion and exposed at 4°C for 7 days before being developed and counter stained.

The data indicate that the hVR-1 molecule is not expressed in human nor monkey brain. The hVR-1 molecule is expressed in nodose, trigeminal sensory neurons, but is not expressed in sympathetic neurons. Within the nodose sensory neurons and trigeminal sensory neurons, expression was seen in distinct sub-populations. Moreover, hVR1 is expressed in some, but not all, small dorsal root ganglion (DRG) neurons and in a few medium sized DRG neurons. The hVR-1 molecule is partially co-expressed with the neuropeptide CGRP and with substance P which are present in nociceptive neurons.

The data further indicate that the VR-2 molecule is expressed in both human and monkey brain, primarily in cortical neurons. The VR2 molecule is also expressed in other brain regions, for example, the thalamus, striatum, hippocampus, hypothalamus, midbrain, medula and brain stem. In addition, the VR-2 molecule is expressed in parasympathetic neurons of the monkey heart (atrium), nodose sensory neurons, trigeminal (TRG) sensory neurons, dorsal root ganglion sensory neurons, sympathetic neurons, and motor neurons of the spinal cord. The VR2 molecule is widely expressed in TRG and DRG neurons, being present in most small and medium sized neurons and also in a few of the large neurons. VR2, like VR-1, partially co-localizes with CGRP and substance P.

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Trigeminal sensory neurons are recognized pain centers while sympathetic neurons have been shown to be involved in neuropathic pain.

EXAMPLE 2: EXPRESSION OF RECOMBINANT hVR-1, hVR-2, AND rVR-2 PROTEIN IN BACTERIAL CELLS

In this example, hVR-1, hVR-2, and rVR-2 is expressed as a recombinant glutathione-S-transferase (GST) fusion polypeptide in *E. coli* and the fusion polypeptide is isolated and characterized. Specifically, hVR-1, hVR-2, and rVR-2 is fused to GST and this fusion polypeptide is expressed in *E. coli*, *e.g.*, strain PEB199. Expression of the GST-hVR-1, GST-hVR-2, and GST-rVR-2 fusion protein in PEB199 is induced with IPTG. The recombinant fusion polypeptide is purified from crude bacterial lysates of the induced PEB199 strain by affinity chromatography on glutathione beads. Using polyacrylamide gel electrophoretic analysis of the polypeptide purified from the bacterial lysates, the molecular weight of the resultant fusion polypeptide is determined.

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EXAMPLE 3: EXPRESSION OF RECOMBINANT hVR-1, hVR-2, AND rVR-2 PROTEIN IN COS CELLS

To express the hVR-1, hVR-2, and rVR-2 gene in COS cells, the pcDNA/Amp vector from Invitrogen Corporation (San Diego, CA) is used. This vector contains an SV40 origin of replication, an ampicillin resistance gene, an *E. coli* replication origin, a CMV promoter followed by a polylinker region, and an SV40 intron and polyadenylation site. A DNA fragment encoding the entire hVR-1, hVR-2, and rVR-2 protein and an HA tag (Wilson *et al.* (1984) *Cell* 37:767) or a FLAG tag fused in-frame to its 3' end of the fragment is cloned into the polylinker region of the vector, thereby placing the expression of the recombinant protein under the control of the CMV promoter.

To construct the plasmid, the hVR-1, hVR-2, and rVR-2 DNA sequence is amplified by PCR using two primers. The 5' primer contains the restriction site of interest followed by approximately twenty nucleotides of the hVR-1, hVR-2, and rVR-2 coding sequence starting from the initiation codon; the 3' end sequence contains complementary sequences to the other restriction site of interest, a translation stop

codon, the HA tag or FLAG tag and the last 20 nucleotides of the hVR-1, hVR-2, and rVR-2 coding sequence. The PCR amplified fragment and the pCDNA/Amp vector are digested with the appropriate restriction enzymes and the vector is dephosphorylated using the CIAP enzyme (New England Biolabs, Beverly, MA). Preferably the two restriction sites chosen are different so that the hVR-1, hVR-2, and rVR-2 gene is inserted in the correct orientation. The ligation mixture is transformed into *E. coli* cells (strains HB101, DH5a, SURE, available from Stratagene Cloning Systems, La Jolla, CA, can be used), the transformed culture is plated on ampicillin media plates, and resistant colonies are selected. Plasmid DNA is isolated from transformants and examined by restriction analysis for the presence of the correct fragment.

COS cells are subsequently transfected with the hVR-1, hVR-2, and rVR-2pcDNA/Amp plasmid DNA using the calcium phosphate or calcium chloride coprecipitation methods, DEAE-dextran-mediated transfection, lipofection, or electroporation. Other suitable methods for transfecting host cells can be found in Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning: A Laboratory Manual. 15 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989. The expression of the hVR-1, hVR-2, and rVR-2 polypeptide is detected by radiolabelling (35S-methionine or 35S-cysteine available from NEN, Boston, MA, can be used) and immunoprecipitation (Harlow, E. and Lane, D. Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring 20 Harbor, NY, 1988) using an HA specific monoclonal antibody. Briefly, the cells are labelled for 8 hours with ³⁵S-methionine (or ³⁵S-cysteine). The culture media are then collected and the cells are lysed using detergents (RIPA buffer, 150 mM NaCl, 1% NP-40, 0.1% SDS, 0.5% DOC, 50 mM Tris, pH 7.5). Both the cell lysate and the culture media are precipitated with an HA specific monoclonal antibody. Precipitated 25 polypeptides are then analyzed by SDS-PAGE.

Alternatively, DNA containing the hVR-1, hVR-2, and rVR-2 coding sequence is cloned directly into the polylinker of the pCDNA/Amp vector using the appropriate restriction sites. The resulting plasmid is transfected into COS cells in the manner described above, and the expression of the hVR-1, hVR-2, and rVR-2 polypeptide is

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- 95 -

detected by radiolabelling and immunoprecipitation using an hVR-1, hVR-2, and rVR-2 specific monoclonal antibody.

EXAMPLE 4: ELECTROPHYSIOLOGICAL STUDIES OF VR2

Human VR2 was functionally characterized in both HEK293 cells and *Xenopus* oocytes using electrophysiological methods. VR2 (in the pcDNA3.1 vector purchased by Invitrogen) was transiently expressed in HEK293 cells (ATCC) and recordings were performed 48 hours after transfection of cells using the whole-cell patch-clamp method (described in Bertil Hille, Ionin Channels of excitable membranes, 1992; Hammill *et al.* (1981) *Pluger Arch.* 391:85-100). The results indicate that heat stimulation (>50 °C) induces a rapid inactivating inward current (1-2 nA). Heat-evoked currents of VR2 displayed profound desensitization and could be reversibly blocked by the VR1 inhibitor capsazepin (at a 10 μM concentration). In contrast to rat VR1, Capsaicin (at a 1-10 μM concentration), resiniferatoxin (at a 0.1-3 μM concentration), and low pH (5.0-6.0) do not induce any currents from VR2. Binding studies of [³H]-resiniferatoxin (NEN) to both human VR1 and VR2 in membranes isolated from HEK293 cell homogenates also indicate that resiniferatoxin (at a 0.1-10 nM concentration) has no specific binding to VR2 while it binds to human VR1 with high affinities.

For the oocyte studies, human VR2 was subcloned into an oocyte expression

vector containing 5'- and 3'-UTR of *Xenopus* β-globin (Chiara *et al.* (1999) *Biochemistry* 38(20)6689-6698). *In vitro* transcription was carried out as described in

Chiara *et al.* (*supra*) and cRNA (10-100 ng) was then injected into the oocytes. VR2

function was characterized in the oocytes 48 hours after cRNA injection using a standard two-electrode voltage-clamp. Consistent with the data from the HEK293 studies, VR2

can only be activated by heat stimulation (48-50 °C) but not by vanilloid receptor agonists, capsaicin, or resiniferatoxin. The vanilloid receptor antagonist capsazepine (at a 1-10 μM concentration) blocks the heat response of VR2 reversibly.

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EXAMPLE 5: GENERATION OF ANTI-hVR-2 ANTIBODIES AND hVR-2 PROTEIN LOCALIZATION BY IMMUNOSTAINING

Polyclonal antisera were raised in rabbits against the following three peptides derived from the human VR2 amino acid sequence, using the techniques described in Ed Harlow and David Lane (1988) "Antibodies; A Laboratory Manual" Cold Spring harbor Laboratory Press.

Antibody PEPTIDE 1: AFHCKSPHRHRMVVLE (SEQ ID NO:13)

Antibody PEPTIDE 2: RPEAPTGPNATESVQPMEGQEDEGN (SEQ ID NO:14)

Antibody PEPTIDE 3: SVLEMENGYWWCRKKQRAG (SEQ ID NO:15)

10 Antisera were subsequently affinity purified using the peptide immunogen.

The polyclonal antisera were tested for immunostaining of both monkey and rat dorsal root ganglion sensory neurons. Peptides 1 and 3 gave specific staining of subpopulations of sensory neurons that was competed with the corresponding peptide. This pattern of expression was very similar to the one observed using a VR-2 riboprobe.

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EXAMPLE 6: CHROMOSOMAL LOCALIZATION OF hVR-1 AND hVR-2

To chromosomally map the hVR-1 gene, primers were designed based on the sequence of hVR-1 (clone Fchrb87a6) (amplifying a 177 bp product from a human control cell line DNA and multiple faint larger products from a control Hamster cell line DNA by PCR). These primers were used to amplify 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL).

The hVR-1 primers used in the PCR mapping studies were: forward - TAGGAGACCCCGTTGCCACG (SEQ ID NO:16) and reverse -

GATTCACTTGGGGACAGTGACG (SEQ ID NO:17) and the PCR reactions were performed as follows: 5 μl Template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR Buffer, 1.2μl Pharmacia dNTP mix 2.5 mM, 1.15μl Forward primer 6.6μM, 1.15μl Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq .05U/μl (Hot Start), using an amplification profile of: 95°C for 10 minutes followed by 35 Cycles of 94°C for 40 seconds, 55°C for 40 seconds, 72°C for 40 seconds, and 72°C for 5 minutes. The PCR

- 97 -

products were run on 2% agarose gels, post-stainedwith SYBR Gold (1:10,000 dilution in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

10 RH linkage analysis was performed using the Map Manager QTb28 software package.

hVR1 was found to map to the p arm of human chromosome 17, 18.9 cR₃₀₀₀ telomeric to the Whitehead Institute framework marker WI-6584, and 7.7 cR₃₀₀₀ centromeric of the Whitehead framework marker WI-5436. LOD scores for linkage were 14.5 for WI-6584 and 19.3 for WI-5436. This region corresponds to the cytogenetic location 17p12-13. This region is syntenic to mouse chromosome 11.

To chromosomally map the hVR-2 gene, primers were designed from 5' UTR sequence of human VR2 (clone Flh21e11) (amplifying a 166 bp product from a human control cell line DNA and 2 much larger faint bands from a control Hamster cell line DNA by PCR). These primers were used to amplify 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL).

The hVR-2 primers used in the PCR mapping studies were: forward TTAAGCTCCCGTTCTCACCG (SEQ ID NO:18) and reverse GCTGCGGGAGGAAGTGAAGC (SEQ ID NO:19) and the PCR reactions were
performed as follows: 5μl Template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR
Buffer, 1.2μl Pharmacia dNTP mix 2.5mM, 1.15μl Forward primer 6.6μM, 1.15μl
Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq .05U/μl (Hot Start), using an
amplification profile of 95°C for 10 minutes, followed by 35 Cycles of 94°C for 40
seconds, 55°C for 40 seconds, 72°C for 40 seconds, and 72°C for 5 minutes. The PCR
products were run on 2% agarose gels, post-stainedwith SYBR Gold (1:10,000 dilution
in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

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The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

RH linkage analysis was performed using the Map Manager QTb28 software package.

hVR2 was found to map to the p arm of human chromosome 17, 29.3cR cR₃₀₀₀ telomeric to the Whitehead Institute framework marker D17S721, and 23.3 cR₃₀₀₀ centromeric of the Whitehead framework marker AFMA043ZB5. LOD scores for linkage were 11.9 for D17S721 and 13.6 for AFMA043ZB5. This region corresponds to the cytogenetic location 17p11-12. This region is syntenic to mouse chromosome 11.

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Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

What is claimed:

- 1. An isolated nucleic acid molecule selected from the group consisting of:
 - (a) a nucleic acid molecule comprising the nucleotide sequence set
- forth in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof; and
 - (b) a nucleic acid molecule consisting of the nucleotide sequence set forth in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof.
- 2. An isolated nucleic acid molecule which encodes a polypeptide selected 10 from the group consisting of:
 - (a) a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11; and
 - (b) a polypeptide consisting of the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11.

- 3. An isolated nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11.
- 4. An isolated nucleic acid molecule selected from the group consisting of:
 - a) a nucleic acid molecule comprising a nucleotide sequence which is at least 83% identical to the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof;
- b) a nucleic acid molecule comprising a fragment of at least 20 25 nucleotides of a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof;
 - c) a nucleic acid molecule which encodes a polypeptide comprising an amino acid sequence at least about 87% identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11; and

d) a nucleic acid molecule which encodes a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15 contiguous amino acid residues of the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.

- 5. An isolated nucleic acid molecule comprising the nucleic acid molecule of any one of claims 1, 2, 3, or 4, and a nucleotide sequence encoding a heterologous polypeptide.
- 10 6. A vector comprising the nucleic acid molecule of any one of claims 1, 2, 3, or 4.
 - 7. The vector of claim 6, which is an expression vector.
- 15 8. A host cell transfected with the expression vector of claim 7.
 - 9. A method of expressing a polypeptide comprising culturing the host cell of claim 8 in an appropriate culture medium to, thereby, express the polypeptide.

- 10. An isolated polypeptide selected from the group consisting of:
- a) a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15 contiguous amino acids of SEQ ID NO:2, 5, 8, or 11;
- b) a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the polypeptide is encoded by a nucleic acid molecule which hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions;
- c) a polypeptide which is encoded by a nucleic acid molecule comprising a nucleotide sequence which is at least 83% identical to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12; and
 - d) a polypeptide comprising an amino acid sequence which is at least 87% identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.
- 15 11. The isolated polypeptide of claim 10 comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.
 - 12. The polypeptide of claim 10, further comprising heterologous amino acid sequences.
 - 13. An antibody which selectively binds to a polypeptide of claim 10.
 - 14. A method for detecting the presence of a polypeptide of claim 10 in a sample comprising:
- 25 a) contacting the sample with a compound which selectively binds to the polypeptide; and
 - b) determining whether the compound binds to the polypeptide in the sample to thereby detect the presence of a polypeptide of claim 10 in the sample.
- The method of claim 14, wherein the compound which binds to the polypeptide is an antibody.

- 16. A kit comprising a compound which selectively binds to a polypeptide of claim 10 and instructions for use.
- 5 17. A method for detecting the presence of a nucleic acid molecule of any one of claims 1, 2, 3, or 4 in a sample comprising:
 - a) contacting the sample with a nucleic acid probe or primer which selectively hybridizes to the nucleic acid molecule; and
- b) determining whether the nucleic acid probe or primer binds to a nucleic acid molecule in the sample to thereby detect the presence of a nucleic acid molecule of any one of claims 1, 2, 3, or 4 in the sample.
 - 18. The method of claim 17, wherein the sample comprises mRNA molecules and is contacted with a nucleic acid probe.

19. A kit comprising a compound which selectively hybridizes to a nucleic acid molecule of any one of claims 1, 2, 3, or 4 and instructions for use.

- 20. A method for identifying a compound which binds to a polypeptide of claim 10 comprising:
 - a) contacting the polypeptide, or a cell expressing the polypeptide with a test compound; and
 - b) determining whether the polypeptide binds to the test compound.
- 25 21. The method of claim 20, wherein the binding of the test compound to the polypeptide is detected by a method selected from the group consisting of:
 - a) detection of binding by direct detection of test compound/polypeptide binding;
 - b) detection of binding using a competition binding assay; and
- 30 c) detection of binding using an assay for hVR-1, hVR-2, or rVR-2 activity.

22. A method for modulating the activity of a polypeptide of claim 10 comprising contacting the polypeptide or a cell expressing the polypeptide with a compound which binds to the polypeptide in a sufficient concentration to modulate the activity of the polypeptide.

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- 23. A method for identifying a compound which modulates the activity of a polypeptide of claim 10 comprising:
 - a) contacting a polypeptide of claim 10 with a test compound; and
- b) determining the effect of the test compound on the activity of the polypeptide to thereby identify a compound which modulates the activity of the polypeptide.
 - 24. A method for treating a subject having a disorder characterized by aberrant hVR-1 or hVR-2 protein activity or nucleic acid expression comprising administering to the subject a hVR-1 or hVR-2 modulator such that treatment of the subject occurs.
 - 25. The method of claim 24, wherein the hVR-1 or hVR-2 modulator is a small molecule.

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26. The method of claim 24, wherein the disorder is a pain disorder.

1/37

humanVRl gene with translation of open reading frame Input file Fchrb87a6.seq; Output File Fchrb87a6.tra Sequence length 3909

GTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAA CGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGG GCAGTGAGCGCAACGCACTTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGC GCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTG TTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGTGAGCGGATAACAATTTCACACAGGAAACAGCTATGACCATGATT ${\tt ACGCCAAGCTCTAATACGACTCACTATAGGGAAAGCTGGTACGCCTGCAGGTACCGGTCCGGAATTCCCGGGTCGACCC}$ ACGCGTCCGAAAACACACCTCTCTGCTGTGGGAAGACTGTGCAATGGCACAGCCGCAGAGCTTGGTTTGGGAGGTTGAA GTGCTCTGGGGAGAATTCGTAGATCATCCTCAGAAAAGCCTTGCCCTGGTGTTCTACCAGAAAAACGTCTCCCAATCAC CCAGAAAAGCTGTCCACAGTAGTCCCCCCTTATCCACGGGTGTCACTTTCCATGGGTTCAGTTATTTGCGGTCAACCAC GGTCTGCCAATATTAAATGGAAAATTCTTCAAACAGTTCCCAAGTTTTCCCTTGTGCATTGTTCTGAGCAGTGTGATGA AGAGTCTCTGCCGTGCCATCTGGGATGCAAACCGTCCCTGTGTCCCCCACGTCCAGGCCGTAGATGCTCCCCGCCGGTC AGTCACTTAGTCGTCAGATCGCCCGTCCTGGTATCACAGTGCTTCTGTTCAGGTTGCACACTGGGCCACAGAGGATCCA K K W S S T D L G A D

18 GCAAGG ATG AAG AAA TGG AGC AGC ACA GAC TTG GGG ACA GCT GCG GAC CCA CTC CAA AAG P D L G D P N S R P P GAC ACC TGC CCA GAC CCC CTG GAT GGA GAC CCT AAC TCC AGG CCA CCT CCA GCC AAG CCC 114 K R T F R L K G D S Ε 58 Ε CAG CTC CCC ACG GCC AAG AGC CGC ACC CGG CTC TTT GGG AAG GGT GAC TCG GAG GAT 174 E Ε L TTC CCG GTG GAT TGC CCC CAC GAG GAA GGT GAG TTG GAC TCC TGC CCG ACC ATC ACA GTC 234 Ι P Ι R G D G Ρ T G Α R L L 98 AGC CCT GTT ATC ACC ATC CAG AGG CCA GGA GAC GGC CCC ACC GGT GCC AGG CTG CTG TCC 294

Fig. 1

2/37

Ε KTL R L Y D CAG GAC TCT GTC GCC GCC AGC ACC GAG AAG ACC CTC AGG CTC TAT GAT CGC AGG AGT ATC 354 S L F N С 0 D L Ε L TTT GAA GCC GTT GCT CAG AAT AAC TGC CAG GAT CTG GAG AGC CTG CTG CTC TTC CTG CAG 414 158 F K D P T E Ε G K N AAG AGC AAG AAG CAC CTC ACA GAC AAC GAG TTC AAA GAC CCT GAG ACA GGG AAG ACC TGT 178 I P L LLK A M L N L H D G O N T T L CTG CTG AAA GCC ATG CTC AAC CTG CAC GAC GGA CAG AAC ACC ACC ATC CCC CTG CTC CTG 198 K E L V N A S Y D S L GAG ATC GCG CGG CAA ACG GAC AGC CTG AAG GAG CTT GTC AAC GCC AGC TAC ACG GAC AGC 218 A L H I A I Ε R R N TAC TAC AAG GGC CAG ACA GCA CTG CAC ATC GCC ATC GAG AGA CGC AAC ATG GCC CTG GTG 654 238 V Q D H N G A Α A A ACC CTC CTG GTG GAG AAC GGA GCA GAC GTC CAG GCT GCG GCC CAT GGG GAC TTC TTT AAG 714 258 T K GRPG F Y F G E L P L S AAA ACC AAA GGG CGG CCT GGA TTC TAC TTC GGT GAA CTG CCC CTG TCC CTG GCC GCG TGC 774 I V K F L L O N S W 278 ACC AAC CAG CTG GGC ATC GTG AAG TTC CTG CTG CAG AAC TCC TGG CAG ACG GCC GAC ATC 834 298 V L H A L AGC GCC AGG GAC TCG GTG GGC AAC ACG GTG CTG CAC GCC CTG GTG GAG GTG GCC GAC AAC 894 V M Y N L 318 D K F T S E Ι M L ACG GCC GAC AAC ACG AAG TTT GTG ACG AGC ATG TAC AAT GAG ATT CTG ATG CTG GGG GCC LHPTLKL K K TPL 338 E E L T N G M AAA CTG CAC CCG ACG CTG AAG CTG GAG GAG CTC ACC AAC AAG AAG GGA ATG ACG CCG CTG 1014 T G K I G V L Α Y Ι L 0 R GCT CTG GCA GCT GGG ACC GGG AAG ATC GGG GTC TTG GCC TAT ATT CTC CAG CGG GAG ATC 1074 Н S R K E L F CAG GAG CCC GAG TGC AGG CAC CTG TCC AGG AAG TTC ACC GAG TGG GCC TAC GGG CCC GTG 1134 Y D L S C I D C E K N 398 CAC TCC TCG CTG TAC GAC CTG TCC TGC ATC GAC ACC TGC GAG AAG AAC TCG GTG CTG GAG 1194 418 E P N Н D GTG ATC GCC TAC AGC AGC AGC GAG ACC CCT AAT CGC CAC GAC ATG CTC TTG GTG GAG CCG 1254

Fig. 1 continued

SUBSTITUTE SHEET (RULE 26)

3/37

N R L L 0 D K W D R F V K R I F Y F N 438 CTG AAC CGA CTC CTG CAG GAC AAG TGG GAC AGA TTC GTC AAG CGC ATC TTC TAC TTC AAC 1314 VYC L Y M I I F T M A A Y Y 458 R TTC CTG GTC TAC TGC CTG TAC ATG ATC ATC TTC ACC ATG GCT GCC TAC TAC AGG CCC GTG 1374 p F K M Ε K I G D Y F R 478 GAT GGC TTG CCT CCC TTT AAG ATG GAA ARA ATT GGA GAC TAT TTC CGA GTT ACT GGA GAG 1434 G G F Y F F R G Ι 0 Y F L ATC CTG TCT GTG TTA GGA GGA GTC TAC TTC TTT TTC CGA GGG ATT CAG TAT TTC CTG CAG 1494 R P S M K F V D S Y S Ε М F L 518 AGG CGG CCG TCG ATG AAG ACC CTG TTT GTG GAC AGC TAC AGT GAG ATG CTT TTC TTT CTG 1554 F M L Α T V v LYFS H L K Ε Y 538 CAG TCA CTG TTC ATG CTG GCC ACC GTG GTG CTG TAC TTC AGC CAC CTC AAG GAG TAT GTG 1614 F S L A L G W T N M L Y Y Т GCT TCC ATG GTA TTC TCC CTG GCC TTG GGC TGG ACC AAC ATG CTC TAC TAC ACC CGC GGT 1674 Y Α M Ι Ε K M L R TTC CAG CAG ATG GGC ATC TAT GCC GTC ATG ATA GAG AAG ATG ATC CTG AGA GAC CTG TGC 1734 M F Y Ι V F F G L F S Α V V 598 CGT TTC ATG TTT GTC TAC ATC GTC TTC TTG TTC GGG TTT TCC ACA GCG GTG GTG ACG CTG 1794 K N D S L P S E S Т S H R W 618 ATT GAA GAC GGG AAG AAT GAC TCC CTG CCG TCT GAG TCC ACG TCG CAC AGG TGG CGG GGG 1854 D S S Y N S L Y S T C L E 638 CCT GCC TGC AGG CCC CCC GAT AGC TCC TAC AAC AGC CTG TAC TCC ACC TGC CTG GAG CTG 1914 M G D L Е F Ε N Y TTC AAG TTC ACC ATC GGC ATG GGC GAC CTG GAG TTC ACT GAG AAC TAT GAC TTC AAG GCT 1974 V F I Ι L L L Y Α V I L T Y Ι L L L N 678 GTC TTC ATC ATC CTG CTG CTG GCC TAT GTA ATT CTC ACC TAC ATC CTC CTG CTC AAC ATG 2034 M Ε Т ٧ N K Ι K 698 CTC ATC GCC CTC ATG GGT GAG ACT GTC AAC AAG ATC GCA CAG GAG AGC AAG AAC ATC TGG 2094 Ι Ι L D T Ε K S F L K 718 AAG CTG CAG AGA GCC ATC ACC ATC CTG GAC ACG GAG AAG AGC TTC CTT AAG TGC ATG AGG 2154

Fig. 1 continued

SUBSTITUTE SHEET (RULE 26)

K	Α	F	R	S	G	K	L	L	Q	V	G	Y	T	P	D	G	K	D	D	738
																				2214
																				758
TAC	CGG	TGG	TGC	TTC	AGG	GTG	GAC	GAG	GTG	AAC	TGG	ACC	ACC	TGG	AAC	ACC	AAC	GTG	GGC	2274
I	I	N	E	D	P	G	N	C	E	G	v	K	R	T	L	S	F	S	L	778
ATC	ATC	AAC	GAA	GAC	CCG	GGC	AAC	TGT	GAG	GGC	GTC	AAG	CGC	ACC	CTG	AGC	TTC	TCC	CTG	2334
R	S	S	R	v	S	G	R	н	W	ĸ	N	F	A	L	V	P	L	L	R	798
																				2394
E	A	S	A	R	D	R	0	S	A	0	P	E	E	v	Y	L	R	0	F	818
																				2454
S	G	S	L	K	P	E	D	A	E	v	F	K	S	P	A	A	S	G	E	838
																				2514
ĸ	•																			840
AAG	TGA																			2520

Fig. 1 continued

Full-length human VR2

Input file Flh21e11.seq; Output File Flh21e11.tra Sequence length 2809

S P TGCACAGAGGTCCTGGCTGGACCGAGCAGCCTCCTCCTAGG ATG ACC TCA CCC TCC AGC TCT CCA 24 V R L Ε T L D G G 0 Ε D G S E Α D R 28 GTT TTC AGG TTG GAG ACA TTA GAT GGA GGC CAA GAA GAT GGC TCT GAG GCG GAC AGA GGA G L P P M E S Q F Q G 48 AAG CTG GAT TTT GGG AGC GGG CTG CCT CCC ATG GAG TCA CAG TTC CAG GGC GAG GAC CGG 144 Ι R N L N Y R K G G 68 T AAA TTC GCC CCT CAG ATA AGA GTC AAC CTC AAC TAC CGA AAG GGA ACA GGT GCC AGT CAG 204 N D R D R L F N A R 88 CCG GAT CCA AAC CGA TTT GAC CGA GAT CGG CTC TTC AAT GCG GTC TCC CGG GST GTC CCC 264 Ε D L Α G L P Y S E L R T S R Y L 108 T D GAG GAT CTG GCT GGA CTT CCA GAG TAC CTG AGC AAG ACC AGC AAG TAC CTC ACC GAC TCG 324 Ε S G K T C L 128 GAA TAC ACA GAG GGC TCC ACA GGT AAG ACG TGC CTG ATG AAG GCT GTG CTG AAC CTT AAG 384 I L P L L 0 D R 148 GAC GGA GTC AAT GCC TGC ATT CTG CCA CTG CTG CAG ATC GAC AGG GAC TCT GGC AAT CCT 444 L V 0 C Т Α D D Y Y R G H Α 168 CAG CCC CTG GTA AAT GCC CAG TGC ACA GAT GAC TAT TAC CGA GGC CAC AGC GCT CTG CAC 504 Ι Ε Ι R S L 0 C V K L L V Ε N G Α 188 N ATC GCC ATT GAG AAG AGG AGT CTG CAG TGT GTG AAG CTC CTG GTG GAG AAT GGG GCC AAT 564 R G R F F 0 K G G T C F 208 Y GTG CAT GCC CGG GCC TGC GGC CGC TTC TTC CAG AAG GGC CAA GGG ACT TGC TTT TAT TTC 624 K 228 ת GGT GAG CTA CCC CTC TCT TTG GCC GCT TGC ACC AAG CAG TGG GAT GTG GTA AGC TAC CTC

LQATDS T V 248 PHQPAS QGN CTG GAG AAC CCA CAC CAG CCC GCC AGC CTG CAG GCC ACT GAC TCC CAG GGC AAC ACA GTC 744 Ι S S A Ε N I 268 V M D N CTG CAT GCC CTA GTG ATG ATC TCG GAC AAC TCA GCT GAG AAC ATT GCA CTG GTG ACC ACC 804 288 R C P G A L ATG TAT GAT GGG CTC CTC CAA GCT GGG GCC CGC CTC TGC CCT ACC GTG CAG CTT GAG GAC 864 LQDLTPLK K I 308 N L A A K ATC GCC AAC CTG CAG GAT CTC ACG CCT CTG AAG CTG GCC GCC AAG GAG GGC AAG ATC GAG 924 328 F S H I L Q R E G L S ATT TTC AGG CAC ATC CTG CAG CGG GAG TTT TCA GGA CTG AGC CAC CTT TCC CGA AAG TTC 984 348 R S L Y D L ACC GAG TGG TGC TAT GGG CCT GTC CGG GTG TCG CTG TAT GAC CTG GCT TCT GTG GAC AGC 1044 368 E Ι Ι Α F H C K S P H R TGT GAG GAG AAC TCA GTG CTG GAG ATC ATT GCC TTT CAT TGC AAG AGC CCG CAC CGA CAC 1104 MVVL E P LNKLLO LI 388 Α K W D L CGA ATG GTC GTT TTG GAG CCC CTG AAC AAA CTG CTG CAG GCG AAA TGG GAT CTG CTC ATC 1164 408 N L N L Ι Y М Ι F C CCC AAG TTC TTC TTA AAC TTC CTG TGT AAT CTG ATC TAC ATG TTC ATC TTC ACC GCT GTT 1224 L T L K K A A P Н K Α GCC TAC CAT CAG CCT ACC CTG AAG AAG CAG GCC GCC CCT CAC CTG AAA GCG GAG GTT GGA 1284 G Η I L I L L G Ι Y AAC TCC ATG CTG CTG ACG GGC CAC ATC CTT ATC CTG CTA GGG GGG ATC TAC CTC CTC GTG 1344 Y F R R H V F Ι W Ι S F Ι W GGC CAG CTG TGG TAC TTC TGG CGG CGC CAC GTG TTC ATC TGG ATC TCG TTC ATA GAC AGC 1404 F F 0 A L Ŀ V V S TAC TTT GAA ATC CTC TTC CTG TTC CAG GCC CTG CTC ACA GTG GTG TCC CAG GTG CTG TGT 1464 Α TTC CTG GCC ATC GAG TGG TAC CTG CCC CTG CTT GTG TCT GCG CTG GTG CTG GGC TGG CTG 1524 I Y Y Y T R G F 0 H T G S V Ι 0 528 AAC CTG CTT TAC TAT ACA CGT GGC TTC CAG CAC ACA GGC ATC TAC AGT GTC ATG ATC CAG 1584

Fig. 2 continued

	V GTC	I ATC	L CTG	R CGG	D GAC	L CTG	L CTG	R CGC	F TTC	L CTT	L CTG	I ATC	Y TAC	L TTA	V GTC	F TTC	L CTT	F TTC	G GGC	548 1644
F TTC	A GCT	V GTA	A GCC	L CTG	V GTG	S AGC	L CTG	S AGC	Q CAG	E GAG	A GCT	W TGG	R CGC	P CCC	E GAA	A GCT	P CCT		G GGC	568 1704
P CCC	N AAT		T ACA		S TCA	V GTG	Q CAG	P CCC	M ATG	E GAG	G GGA	Q CAG	E GAG	D GAC	E GAG	G GGC	N AAC		A GCC	588 1764
Q CAG	_	R AGG	_	I ATC	_	E GAA							K AAA				G GGC	M ATG	G GGC	608 1824
E GAG	L CTG		F TTC		E GAG								V GTG					L CTG		628 1884
_	V GTG	L CTG			Y TAC				L CTC							M ATG	S AGC	E GAG	T ACC	648 1944
V GTC		_	V GTC	A GCC	T ACT	D GAC	S AGC	W TGG	S AGC	I ATC	W TGG	K AAG	L CTG	Q CAG	K AAA	A GCC		S TCT	V GTC	668 2004
L CTG	_		E GAG	N AAT	G GGC	Y TAT	W TGG	W TGG	C TGC	R AGG	K AAG	K AAG	Q CAG	R CGG	A GCA	G GGT	V GTG	M ATG	L CTG	688 2064
			T ACT						P CCG				W TGG			R AGG	V GTG	E GAG	E GAG	708 2124
V GTG	N AAC	W TGG	A GCT	S TCA	W TGG	E GAG	Q CAG	T ACG	L CTG	P CCT	T ACG	L CTG	C TGT	E GAG	D GAC			G GGG	A GCA	728 2184
G GGT			R CGA						V GTC				P CCT	P CCC	K AAG	E GAG	D GAT	E GAG		748 2244
G GGT	A GCC	S TCT	E GAG	E GAA	N AAC	Y TAT	V GTG	P CCC	V GTC	Q CAG	L CTC	L CTC	Q CAG	S TCC	N AAC					765 2295
TGGC	CCAG	ATGC	AGCA	GGAG	GCCA	GAGG	ACAG	AGCA	GAGG	ATCI	TTCC	AACC	ACA1	CTGC	TGGC	CTCTG	GGGI	CCCA	GTG	

Fig. 2 continued

8/37

Partial human VR2 alternate form

Input file frhob12c4.seg; Output File frhob12c4.tra
Sequence length 1489

19 T C G Q G F Y F G Ε L F F O K GC GGC CGC TTC TTC CAG AAG GGC CAA GGG ACT TGC TTT TAT TTC GGT GAG CTA CCC CTC 57 39 T K W D V V S Y L E C Q L N A TCT TTG GCC GCT TGC ACC AAG CAG TGG GAT GTG GTA AGC TAC CTC CTG GAG AAC CCA CAC 117 59 L D S Q G N T V L H CAG CCC GCC AGC CTG CAG GCC ACT GAC TCC CAG GGC AAC ACA GTC CTG CAT GCC CTA GTG 177 S Y D G 79 Ι S D N S E I A L V T M M A N ATG ATC TCG GAC AAC TCA GCT GAG AAC ATT GCA CTG GTG ACC AGC ATG TAT GAT GGG CTC 237 99 V E D C P T 0 L R Α R L 297 CTC CAA GCT GGG GCC CGC CTC TGC CCT ACC GTG CAG CTT GAG GAC ATC CGC AAC CTG CAG Ε G K I 119 L K L Α Α K GAT CTC ACG CCT CTG AAG CTG GCC GCC AAG GAG GGC AAG ATC GAG ATT TTC AGG CAC ATC 357 139 S L S H L S R K Ε CTG CAG CGG GAG TTT TCA GGA CTG AGC CAC CTT TCC CGA AAG TTC ACC GAG TGG TGC TAT 417 159 ٧ S C Е G P V R V S L Y S D E N D L Α GGG CCT GTC CGG GTG TCG CTG TAT GAC CTG GCT TCT GTG GAC AGC TGT GAG GAG AAC TCA 477 179 L Α F H C K S P H R H R M V Τ Ι GTG CTG GAG ATC ATT GCC TTT CAT TGC AAG AGC CCG CAC CGA CAC CGA ATG GTC GTT TTG 537 199 D L Ι P K K L L Α K W L GAG CCC CTG AAC AAA CTG CTG CAG GCG AAA TGG GAT CTG CTC ATC CCC AAG TTC TTA 597 219 F A Y N L Ι Y M I F T Α AAC TTC CTG TGT AAT CTG ATC TAC ATG TTC ATC TTC ACC GCT GTT GCC TAC CAT CAG CCT 657 S 239 L K K 0 A P H L K A Ε V G N M A ACC CTG AAG AAG CAG GCC GCC CCT CAC CTG AAA GCG GAG GTT GGA AAC TCC ATG CTG CTG 717 G 259 Ι L Ι L L G Ι Y L L V G ACG GGC CAC ATC CTT ATC CTG CTA GGG GGG ATC TAC CTC CTC GTG GGC CAG CTG TGG TAC 777 279 W F Ι TTC TGG CGG CGC CAC GTG TTC ATC TGG ATC TCG TTC ATA GAC AGC TAC TTT GAA ATC CTC 837

A L L V V S V L 299 TTC CTG TTC CAG GCC CTG CTC ACA GTG GTG TCC CAG GTG CTG TGT TTC CTG GCC ATC GAG L V S Α V L L G TGG TAC CTG CCC CTG CTT GTG TCT GCG CTG GTG CTG GGC TGG CTG AAC CTG CTT TAC TAT H G Ι Y S V M K 339 ACA CGT GGC TTC CAG CAC ACA GGC ATC TAC AGT GTC ATG ATC CAG AAG AAA GCC ATC TCT 1017 L E N G Y W C W R K K R GTC CTG GAG ATG GAG AAT GGC TAT TGG TGG TGC AGG AAG AAG CAG CGG GCA GGT GTG ATG 1077 K P D G S P D E R C W F R V 379 CTG ACC GTT GGC ACT AAG CCA GAT GGC AGC CCG GAT GAG CGC TGG TGC TTC AGG GTG GAG 1137 E T L P T L C E D P 399 GAG GTG AAC TGG GCT TCA TGG GAG CAG ACG CTG CCT ACG CTG TGT GAG GAC CCG TCA GGG 1197 R L Р P Α K GCA GGT GTC CCT CGA ACT CTC GAG AAC CCT GTC CTG GCT TCC CCT CCC AAG GAG GAT GAG 1257 A S E E N Y V P V L L 437 GAT GGT GCC TCT GAG GAA AAC TAT GTG CCC GTC CAG CTC CTC CAG TCC AAC TGA 1311 TGGCCCAGATGCAGCAGGGGCCAGAGGACAGAGCAGAGGATCTTTCCAACCACATCTGCTGGCTCTGGGGTCCCAGTG GCGGACGCGTGGGTCGAC

Fig. 3 continued

10/37

Partial rat VR2

Input file Flrxb147g11.seq; Output File Flrxb147g11.tra Sequence length 1794

C K 19 S L Α T A A G TCG ACC CAC GCG TCC GCT CTT TCT CTG GCT GCG TGC ACC AAG CAG TGG GAT GTG GTG 57 39 P H P S Ε N A L Α T ACC TAC CTC CTG GAG AAC CCA CAC CAG CCG GCC AGC CTG GAG GCC ACC GAC TCC CTG GGC 117 59 Ε H A L M Ι Α D N S P Α AAC ACA GTC CTG CAT GCT CTG GTA ATG ATT GCA GAT AAC TCG CCT GAG AAC AGT GCC CTG 177 79 Ι Y D G L 0 M G R L L Α GTG ATC CAC ATG TAC GAC GGG CTT CTA CAA ATG GGG GCG CGC CTC TGC CCC ACT GTG CAG 237 99 G P L Α N H L T K L CTT GAG GAA ATC TCC AAC CAC CAA GGC CTC ACA CCC CTG AAA CTA GCC GCC AAG GAA GGC 297 Ε F G P 119 R H Ι L R S Y AAA ATC GAG ATT TTC AGG CAC ATT CTG CAG CGG GAA TTC TCA GGA CCG TAC CAG CCC CTT 357 139 S R K F Ε W C Y G P V R V S L Y D L TCC CGA AAG TTT ACT GAG TGG TGT TAC GGT CCT GTG CGG GTA TCG CTG TAC GAC CTG TCC 417 V C 159 Ε K N S L E Ι Ι A F H K TCT GTG GAC AGC TGG GAA AAG AAC TCG GTG CTG GAG ATC ATC GCT TTT CAT TGC AAG AGC 477 179 R M V V L E P L N K L L CCG AAC CGG CAC CGC ATG GTG GTT TTA GAA CCA CTG AAC AAG CTT CTG CAG GAG AAA TGG 537 199 N F Α C Y L GAT CGG CTC GTC TCA AGA TTC TTC TTC AAC TTC GCC TGC TAC TTG GTC TAC ATG TTC ATC 597 219 F Y H P S L D 0 P Α Ι P TTC ACC GTC GTT GCC TAC CAC CAG CCT TCC CTG GAT CAG CCA GCC ATC CCC TCA TCA AAA 657 239 Ε L G I L G М Ŀ H Ι L GCG ACT TTT GGG GAA TCC ATG CTG CTG CTG GGC CAC ATT CTG ATC CTG CTT GGG GGT ATT 717 Y W R F Ι 259 L W R R L TAC CTC TTA CTG GGC CAG CTG TGG TAC TTT TGG CGG CGG CGC CTG TTT ATC TGG ATC TCA 777 279 Y Ε L F L L 0 L Τ Α L TTC ATG GAC AGC TAC TTT GAA ATC CTC TTT CTC CTT CAG GCT CTG CTC ACA GTG CTG TCC 837

Q	V	L	R	F	M	E	T	E	W	Y	L	P	L	L	V	L	S	L	V	299
CAG	GTG	CTG	CGC	TTC	ATG	GAG	ACT	GAA	TGG	TAC	CTA	CCC	CTG	CTA	GTG	TTA		CTA	GTG	897
L	G	W	L	N	L	L	Y	Y	T	R	G	F	Q	H	T	G	I	Y	S	319
CTG	GGC	TGG	CTG	AAC	CTG	CTT	TAC	TAC	ACA	CGG	GGC	TTT	CAG	CAC	ACA	GGC	ATC	TAC	AGT	957
V	M	I	Q	K	V	I	L	R	D	L	L	R	F	L	L	V	Y	L	V	339
GTC	ATG	ATC	CAG	AAG	GTC	ATC	CTT	CGA	GAC	CTG	CTC	CGT	TTC	CTG	CTG	GTC	TAC	CTG	GTC	1017
F	L	F	G	F	A	V	A	L	V	S	L	S	R	E	A	R	S	P	K	359
TTC	CTT	TTC	GGC	TTT	GCT	GTA	GCC	CTA	GTA	AGC	TTG	AGC	AGA	GAG	GCC	CGA	AGT	CCC	Aaa	1077
A	_	E	D	N	N	S	T	V	T	E	Q	P	T	V	G	Q	E	E	E	379
GCC		GAA	GAT	AAC	AAC	TCC	ACA	GTG	ACG	GAA	CAG	CCC	ACG	GTG	GGC	CAG	GAG	GAG	GAG	1137
P	A	P	Y	R	S	I	L	D	A	S	L	E	L	F	K	F	T	I	g	399
CCA	GCT	CCA	TAT	CGG	AGC	ATT	CTG	GAT	GCC	TCC	CTA	GAG	CTG	TTC	AAG	TTC	ACC	ATT	GGT	1197
M	G	E	L	A	F	Q	E	Q	L	R	F	R	G	V	V	L	L	L	L	419
ATG	GGG	GAG	CTG	GCT	TTC	CAG	GAA	CAG	CTG	CGT	TTT	CGT	GGG	GTG	GTC	CTG	CTG	TTG	CTG	1257
L TTG		Y TAC			L CTC	T ACC	Y TAC	V GTC	L CTG	L CTG	L CTC	N AAC	M ATG	L CTC	I ATT	A GCT	L CTC	M ATG	S AGC	439 1317
e	T	V	N	H	V	A	D	N	S	W	S	I	W	K	L	Q	K	A	I	459
gaa	ACT	GTC	AAC	CAC	GTT	GCT	GAC	AAC	AGC	TGG	AGC	ATC	TGG	AAG	TTG	CAG	AAA	GCC	ATC	1377
S	V	L	E	M	E	N	g	Y	W	W	C	R	R	K	K	H	R	E	G	479
TCT	GTC	TTG	GAG	ATG	GAG	AAT	GGT	TAC	TGG	TGG	TGC	CGG	AGG	AAG	AAA	CAT	CGT	GAA	GGG	1437
R AGG	L	T.	v	17	~	_	_	_											_	400
	CTG	CTG	AAA	GTC	GGC	ACC	R AGG	G GGG	D GAT	G GGT	T ACC	P CCT	D GAT	E GAG	R CGC	W TGG	C TGC	F TTC	AGG	499 1497
v	E	CTG E	AAA V	GTC N	GGC W	ACC A GCT	AGG A	GGG W	GAT E	GGT K	ACC T	CCT	GAT P	GAG T	CGC	TGG S	TGC E	TTC	AGG	
V GTG S	E GAG G	E GAA P	AAA V GTA G	GTC N AAT I	GGC W TGG	ACC A GCT G	AGG A GCT N	GGG W TGG K	GAT E GAG K	GGT K AAG N	ACC T ACT	CCT L CTT	P CCC	GAG T ACC K	CGC L TTA	TGG S TCT G	TGC E GAG K	TTC D GAT N	AGG	1497 519 1557

 ${\tt TGGCCCAGATGCAGCAGGCTGGCAGGATGGAGTAGGGAATCTTCCCAGCCACACCAGAGGCTACTGAATTTTGGTG}$

Fig. 4 continued

12/37 GAP of: humanvr2.pep check: 5746 from: 1 to: 764 humanVR2 Flh21e11 to: humanvrl.pep check: 6877 from: 1 to: 839 humanVR1 _Fbh18547pat - fchrb87a6, 3909 bases, 4554 checksum. Symbol comparison table: /ddm_local/gcg/gcg 9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430 Gap Weight: 12 Average Match: 2.912 Length Weight: 4 Average Mismatch: -2.003 Quality: 1530 Length: 850 Ratio: 2.003 Gaps: Percent Similarity: 55.378 Percent Identity: 46.348 Match display thresholds for the alignment(s): = IDENTITY = 2. = 1 humanvr2.pep x humanvr1.pepMTSPSSSPVF 10 1 MKKWSSTDLGTAADPLQKDTCPDPLDGDPNSRPPPAKPQLPTAKSRTRLF 50 11 RLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY 60 : | . | | : . : : 51 GKGDSEEAFPVDCPHEEGELDSCPTI.TVSPVITIQRPGDGPTGARLLSQ 99 61 RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEY 110 :|| :| ||.. :|| | :| |. |:|||.|: 100 DSVAASTEKTLRLYDRRSIFEAVAONNCODLESLLLFLOKSKKHLTDNEF 149 111 TEGSTGKTCLMKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDY 160 150 KDPETGKTCLLKAMLNLHDGQNTTIPLLLEIARQTDSLKELVNASYTDSY 199

Fig. 5

161 YRGHSALHIAIEKRSLQCVKLLVENGANVHARACGRFFQKGQG.TCFYFG 209

200 YKGQTALHIAIERRNMALVTLLVENGADVQAAAHGDFFKKTKGRPGFYFG 249

210	ELPLSLAACTKOWDVVSYLLENPHOPASLQATDSQGNTVLHALVMISDNS	259
250	ELPLSLAACTNQLGIVKFLLQNSWQTADISARDSVGNTVLHALVEVADNT	299
260	AENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKLAAKEGKIEI	309
300	: .: : :: . :	349
310	FRHILOREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEI :	357
350	LAYILQREIQEPECRHLSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEV	399
358	IAF.HCKSPHRHRMVVLEPLNKLLQAKWDLLIPK.FFLNFLCNLIYMFIF	405
400	: . : :: : :	
	TAVAYHQPTLKKQAAPHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFW :. :. : :. : :. : :. : TMAAYYRPVDGLPPFKMEKIGDYFRVTGEILSVLGGVYFFFRGIQYFL	455
450		
	RRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEWYLPLLVSALVL	
	QŘRPSMKTLÝVĎSÝSÉMLÝFLÓSĽFMLATVÝĽYÝSHLKEÝVASMÝFSĽAĽ	547 555
548		
556	LSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLELFKFTI	605
	:: . :	
606	GMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETVNSVATD	
644	: : . :: :	693
656	SWSIWKLQKAISVLEMENGYWWC.RKKQRAGVMLTVGTKPDGSPDERWCF	704
694	.	743
705	RVEEVNWASWEQTLPTLCEDPSGA.GVPRTLENPVLASPPKEDEDGASEE	753
	: . . : RVDEVNWTTWNTNVGIINEDPGNCEGVKRTLSFSLRSSRVSGRHWK	789
754	NYVPVQLLQSN	764
790	NFALVPLLREASARDRQSAQPEEVYLRQFSGSLKPEDAEVFKSPAASGEK	839

Fig. 5 continued

SUBSTITUTE SHEET (RULE 26)

14/37

GAP of: humanvr2.seq check: 8853 from: 1 to: 2809

humanVR2 21e11a, 2809 bases, 8853 checksum.

to: humanvrl.seq check: 4554 from: 1 to: 3909

humanVR1 Fbh18547pat - Import - complete

Symbol comparison table:

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Quality: 14359 Length: 3934
Ratio: 5.112 Gaps: 15
Percent Similarity: 55.316 Percent Identity: 55.316

Match display thresholds for the alignment(s):

= IDENTITY

: = 5

. = 1

humanvr2.seq x humanvr1.seq

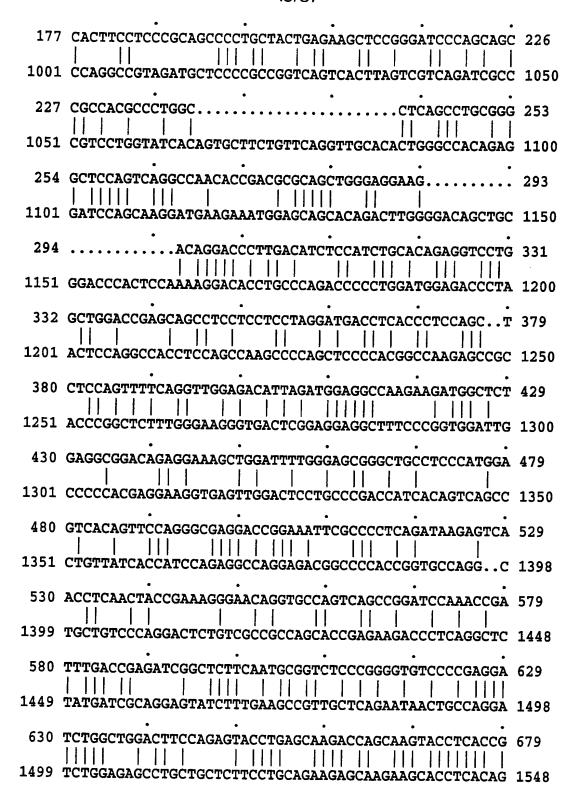


Fig. 6 continued

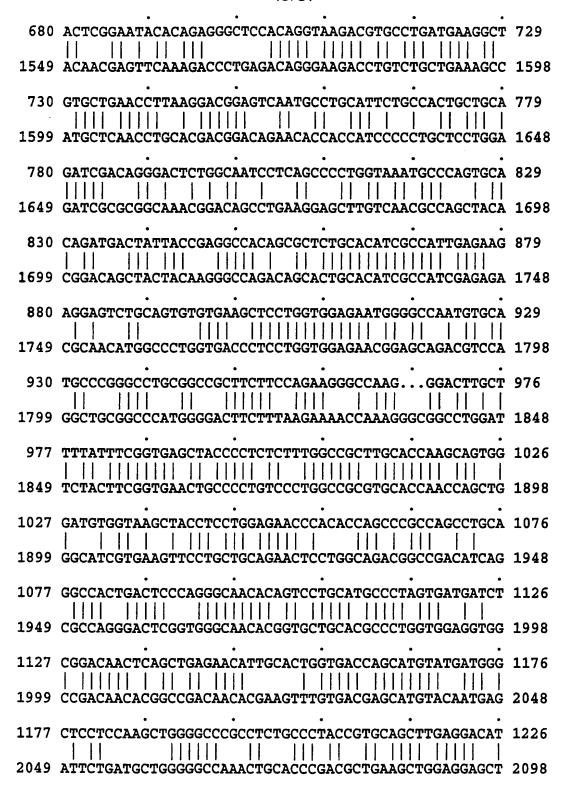


Fig. 6 continued

1227	CCGCAACCTGCAGGATCTCACGCCTCTGAAGCTGGCCGCCAAGGAGGGCA	1276
2099		2148
1277	AGATCGAGATTTTCAGGCACATCCTGCAGCGGGAGTTTTCAGGA	1320
2149	AGATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAG	2198
1321	CTGAGCCACCTTTCCCGAAAGTTCACCGAGTGGTGCTATGGGCCTGTCCG	1370
2199	TGCAGGCACCTGTCCAGGAAGTTCACCGAGTGGGCCCTACGGGCCCGTGCA	2248
1371	GGTGTCGCTGTATGACCTGGCTTCTGTGGACAGCTGTGAGGAGAACTCAG	1420
2249	CTCCTCGCTGTACGACCTGTCCTGCATCGACACCTGCGAGAAGAACTCGG	2298
1421	TGCTGGAGATCATTGCCTTTCATTGCAAGAGCCCGCACCGACACCGA	1467
2299	TGCTGGAGGTGATCGCCTACAGCAGCAGCGAGACCCCTAATCGCCACGAC	2348
1468	ATGGTCGTTTTGGAGCCCCTGAACAACTGCTGCAGGCGAAATGGGA	1514
2349	ATGCTCTTGGTGGAGCCGCTGAACCGACTCCTGCAGGACAAGTGGGACAG	2398
1515	TCTGCTCATCCCCAAGTTCTTCTTAAACTTCCTGTGTAATCTGATCTACA	1564
2399	ATTCGTCAAGCGCATCTTCTACTTCAACTTCCTGGTCTACTGCCTGTACA	2448
1565	TGTTCATCTTCACCGCTGTTGCCTACCATCAGCCTACCCTGAAGAAGCAG	1614
2449	TGATCATCTTCACCATGGCTGCCTACTACAGGCCCGTGGATGGCTT	2494
1615	GCCGCCCTCACCTGAAAGCGGAGGTTGGAAACTCCATGCTGCTGACGGG	1664
2495	GCCTCCCTTTAAGATGGAAAAAATTGGAGACTATTTCCGAGTTACTGG	2542
1665	CCACATCCTTATCCTGCTAGGGGGGATCTACCTCCTCGTGGGCCAGCTGT	1714
2543		2592
1715	GGTACTTCTGGCGGCGCCACGTGTTCATCTGGATCTCGTTCATAGACAGC	1764
2593	AGTATTTCCTGCAGAGGCGGCCGTCGATGAAGACCCTGTTTGTGGACACC	2642

Fig. 6 continued

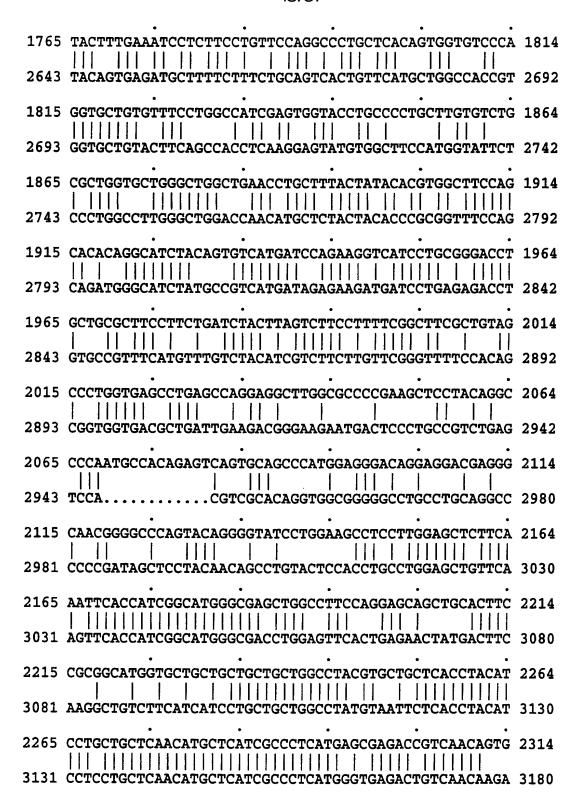


Fig. 6 continued

2315	TCGCCACTGACAGCTGGAGCATCTGGAAGCTGCAGARAGCCATCTCTGTC	2364
3181	TCGCACAGGAGACAACATCTGGAAGCTGCAGAGAGCCATCACCATC	3230
2365	CTGGAGATGGAGAATGGCTATTGGTGGTGCAGGAAGAAGCAGCGGGC	2411
3231	CTGGACACGGAGAAGAGCTTCCTTAAGTGCATGAGGAAGGCCTTCCGCTC	3280
2412	AGGTGTGATGCTGACCGTTGGCACTAAGCCAGATGGCAGCCCGGATGAGC	2461
3281	AGGCAAGCTGCTGCAGGTGGGGTACACCCTGATGGCAAGGACGACTACC	3330
24 62	GCTGGTGCTTCAGGGTGAGGGGGGGGGGGGGGGGGGGGG	2511
3331	GGTGGTGCTTCAGGGTGGACGAGGTGAACTGGACCACCTGGAACACCAAC	3380
2512	CTGCCTACGCTGTGTGAGGACCCGTCAGGGGCAGGTGTCCCTCGAAC	2558
	GTGGGCATCATCAACGAAGACCCGGGCAACTGTGAGGGCGTCAAGCGCAC	
	TCTCGAGAACCCTGTCCTGGCTTCCCCTCCCAAGGAGGATGAGGAT	
	CCTGAGCTTCTCCCTGCGGTCAAGCAGAGTTTCAGGCAGACACTGGAAGA	
	GGTGCCTCTGAGGAAAACTATGTGCCCGTCCAGCTCCAGTCCAACTG	
	ACTTTGCCCTGGTCCCCCTTTTAAGAGAGGCAAGTGCTCGAGATAGGCAG	
	ATGGCCCAGATGCAGCAGGAGGCCAGAGGACAGAGCAGAGGATCTTTCCA	
	ACCACATCTGCTGGGGTCCCAGTGAATTCTGGTGGCAAATATAT	
	ATTTTCACTAACTCAAAAAAAAAAAAAAAAAAAAAAAAA	
2805	AAAAA	2809
3676	CCCCGTTGCCACGGGGGCTGCTGAGGGAACACCAGTGCTCTGTCAGCAG	3725

Fig. 6 continued

SUBSTITUTE SHEET (RULE 26)

20/37

CLUSTAL W (1.74) multiple sequence alignment

humanVR2 rat VR2	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY
humanVR2 rat VR2	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2 rat VR2	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK
humanVR2 rat VR2	LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQASTHASALSLAACTKQWDVVTYLLENPHQPASLEA .************************************
humanVR2 rat VR2	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSLGNTVLHALVMIADNSPENSALVIHMYDGLLQMGARLCPTVQLEEISNHQGLTPLKL *** ********************************
humanVR2 rat VR2	AAKEGKIEIFRHILQREFSG-LSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIA AAKEGKIEIFRHILQREFSGPYQPLSRKFTEWCYGPVRVSLYDLSSVDSWEKNSVLEIIA ***********************************
humanVR2 rat VR2	FHCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQA FHCKSPNRHRMVVLEPLNKLLQEKWDRLVSRFFFNFACYLVYMFIFTVVAYHQPSLDQPA ******:******************************
humanVR2 rat VR2	APHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQAL IPSSKATFGESMLLLGHILILLGGIYLLLGQLWYFWRRRLFIWISFMDSYFEILFLLQAL * ** .*:*** ***************************
humanVR2 rat VR2	LTVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHIGIYSVMIQKVILRDLLRFL LTVLSQVLRFMETEWYLPLLVLSLVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFL ***:*** *: ***************************
humanVR2 rat VR2	LIYLVFLFGFAVALVSLSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLE LVYLVFLFGFAVALVSLSREARSPKAPEDNNSTVTEQPTVGQEEEPAPYRSILDASLE *:***********************************
humanVR2 rat VR2	LFKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETVNSVATDSWSI LFKFTIGMGELAFQEQLRFRGVVLLLLLAYVLLTYVLLLNMLIALMSETVNHVADNSWSI ***********************************
humanVR2 rat VR2	WKLQKAISVLEMENGYWWCR-KKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTL WKLQKAISVLEMENGYWWCRRKKHREGRLLKVGTRGDGTPDERWCFRVEEVNKAAWEKTL
humanVR2 rat VR2	PTLCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN PTLSEDPSGPGITGNKKNPTSK-PGKNSASEEDHLPLQVLQSP ***.****.*::***

GAP of: ratvr2.pep check: 9190 from: 1 to: 554

ratVR2 Flrxb147g11

to: humanvr2.pep check: 5746 from: 1 to: 764

humanVR2 Flh21e11

Symbol comparison table: /usr/local/gog_9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430

Gap Weight: 12 Average Match: 2.912 Length Weight: 4 Average Mismatch: -2.003

Quality: 2182 Length: 766
Ratio: 3.939 Gaps: 4
Percent Similarity: 81.703 Percent Identity: 79.167

Match display thresholds for the alignment(s):

= IDENTITY

: = 2 . = 1

ratvr2.pep x humanvr2.pep

Fig. 8

SUBSTITUTE SHEET (RULE 26)

	• • • • • •	
245	QLWYFWRRRLFIWISFMDSYFEILFLLQALLTVLSQVLRFMETEWYLPLL	294
450	QLWYFWRRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEWYLPLL	499
	• • • • • •	
295	VLSLVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLLVYLVFLFGF	344
500	VSALVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLLIYLVFLFGF	549
	• • • • • • • • • • • • • • • • • • • •	
345	AVALVSLSREARSPKAPEDNNSTVTEQPTVGQEEEPAPYRSILDASLE	392
550	AVALVSLSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLE	599
	• • • • • • • • • • • • • • • • • • • •	
393	LFKFTIGMGELAFQEQLRFRGVVLLLLLAYVLLTYVLLLNMLIALMSETV	442
600	LFKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETV	649
442		
443	NHVADNSWSIWKLQKAISVLEMENGYWWCRRKKHREGRLLKVGTRGDGTP	492
C F A		
650	NSVATDSWSIWKLQKAISVLEMENGYWWC.RKKQRAGVMLTVGTKPDGSP	698
400		
493	DERWCFRVEEVNWAAWEKTLPTLSEDPSGPGITGNKKNPTSKPGKN	538
COO		
699	DERWCFRVEEVNWASWEQTLPTLCEDPSGAGVPRTLENPVLASPPKEDED	748
E 2 0		
55 9	SASEEDHLPLQVLQSP 554	
749		
/49	GASEENYVPVQLLQSN 764	

Fig. 8 continued

GAP of: humanvr1.seq check: 4554 from: 1 to: 3909 humanVR1 Fbh18547pat - Import - complete to: ratvr1.seg check: 7921 from: 1 to: 2847 ratVR1.seq AF029310 in GenBank Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/nwsgapdna.cmp CompCheck: 8760 Gap Weight: 50 Average Match: 10.000 Length Weight: 3 Average Mismatch: 0.000 Quality: 22717 Length: 3914 Ratio: 7.979 10 Gaps: Percent Similarity: 82.125 Percent Identity: 82.125 Match display thresholds for the alignment(s): = IDENTITY : = 5 . = 1 humanvrl . seg x ratvrl . seg 1001 CCAGGCCGTAGATGCTCCCCGCCGGTCAGTCACTTAGTCGTCAGATCGCC 1050CAGCTCCAAGGCACTTGCTCC 21 1051 CGTCCTGGTATCACAGTGCTTCTGTTCAGGTTGCACACTGGGCCACAGAG 1100 22 ATTTGGGGTGTGCCTGCACCT...AGCTGGTTGCAAATTGGGCCACAGAG 68 1101 GATCCAGCAAGGATGAAGAAATGGAGCAGCACAGACTTGGGGGACAGCTGC 1150 69 GATCTGGAAAGGATGGAACAACGGGCTAGCTTAGACTCAGAGGAGTCTGA 118

Fig. 9

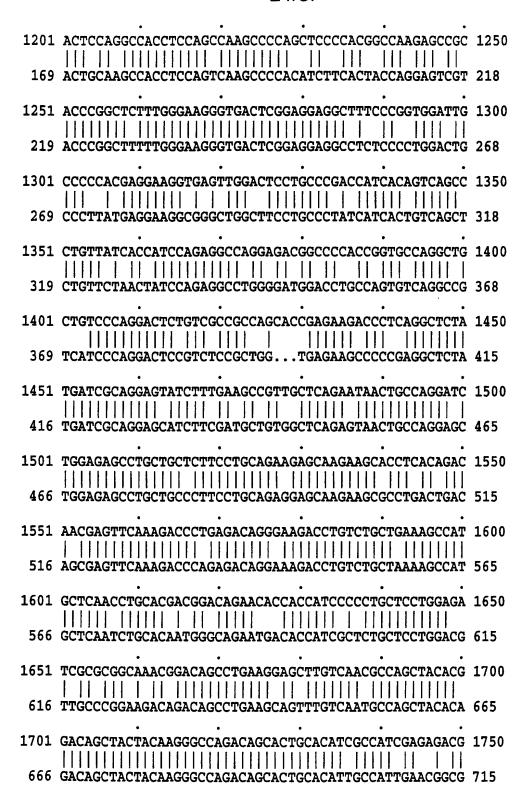


Fig. 9 continued

1751	CAACATGGCCCTGGTGACCCTCCTGGTGGAGAACGGAGCAGACGTCCAGG	1800
716		765
1801	CTGCGGCCCATGGGGACTTCTTTAAGAAAACCAAAGGGCGGCCTGGATTC	1850
766		815
1851	TACTTCGGTGAACTGCCCCTGTCCCTGGCCGCGTGCACCAACCA	1900
816	TACTTTGGTGAGCTGCCCTGTCCCTGGCTGCGTGCACCAACCA	865
1901	CATCGTGAAGTTCCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAGCG	1950
866	CATTGTGAAGTTCCTGCAGAACTCCTGGCAGACATCAGCG	915
1951	CCAGGACTCGGTGGCAACACGGTGCTGCACGCCCTGGTGGAGGTGGCC	2000
	CCCGGGACTCAGGGGCAACACGGTGCTTCATGCCCTGGTGGAGGTGGCA	965
		2050
	GATAACACAGTTGACAACACCAAGTTCGTGACAAGCATGTACAACGAGAT	1015
	TCTGATGCTGGGGGCCAAACTGCACCCGACGCTGAAGCTGGAGGAGCTCA	
	CTTGATCCTGGGGGCCAAACTCCACCCCACGCTGAAGCTGGAAGAGATCA	
	CCAACAAGAAGGGAATGACGCCGCTGGCTCTGGCAGCTGGGACCGGGAAG	2150
	ATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAGTG	
	ATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAGTG	
	CAGGCACCTGTCCAGGAAGTTCACCGAGTGGGCCCTACGGGCCCGTGCACT	
	CCGACACCTATCCAGGAAGTTCACCGAATGGGCCTATGGGCCAGTGCACT	
2251	CCTCGCTGTACGACCTGTCCTGCATCGACACCTGCGAGAAGAACTCGGTG	2300
L216	CCTCCCTTTATGACCTGTCCTGCATTGACACCTGTGAAAAGAACTCGGTT	1265

Fig. 9 continued

2301		2350
1266	CTGGAGGTGATCGCTTACAGCAGCAGTGAGACCCCTAACCGTCATGACAT	1315
2351	GCTCTTGGTGGAGCCGCTGAACCGACTCCTGCAGGACAAGTGGGACAGAT	2400
1316	GCTTCTCGTGGAACCCTTGAACCGACTCCTACAGGACAAGTGGGACAGAT	1365
2401	TCGTCAAGCGCATCTTCTACTTCAACTTCCTGGTCTACTGCCTGTACATG	2450
1366	TTGTCAAGCGCATCTTCTACTTCAACTTCTTCGTCTACTGCTTGTATATG	1415
2451	ATCATCTTCACCATGGCTGCCTACTACAGGCCCGTGGATGGCTTGCCTCC	2500
1416	ATCATCTTCACCGCGGCTGCCTACTATCGGCCTGTGGAAGGCTTGCCCCC	1465
2501	CTTTAAGATGGAAAAATTGGAGACTATTTCCGAGTTACTGGAGAGA	2547
1466	CTATAAGCTGAAAAACACCGTTGGGGACTATTTCCGAGTCACCGGAGAGA	1515
2548	TCCTGTCTGTGTTAGGAGGAGTCTACTTCTTTTTCCGAGGGATTCAGTAT	2597
1516	TCTTGTCTGTGTCAGGAGGAGTCTACTTCTTCTTCCGAGGGATTCAATAT	1565
2598	TTCCTGCAGAGGCGGCCGTCGATGAAGACCCTGTTTGTGGACAGCTACAG	2647
1566	TTCCTGCAGAGGCGACCATCCCTCAAGAGTTTGTTTGTGGACAGCTACAG	1615
2648	TGAGATGCTTTTCTTCTGCAGTCACTGTTCATGCTGGCCACCGTGGTGC	2697
1616		1665
2698	TGTACTTCAGCCACCTCAAGGAGTATGTGGCTTCCATGGTATTCTCCCTG	2747
1666	TGTACTTCAGCCAACGCAAGGAGTATGTGGCTTCCATGGTGTTCTCCCTG	1715
2748	GCCTTGGGCTGGACCAACATGCTCTACTACACCCGCGGTTTCCAGCAGAT	2797
1716	GCCATGGGCTGGACCAACATGCTCTACTATACCCGAGGATTCCAGCAGAT	1765
2798	GGGCATCTATGCCGTCATGATAGAGAAGATGATCCTGAGAGACCTGTGCC	2847
1766	GGCCATCTATCCTCTCTCATCATTCACACACACACCCTCTCTCCCC	1815

Fig. 9 continued

2848	GTTTCATGTTTGTCTACATCGTCTTCTTGTTCGGGTTTTCCACAGCGGTG	2897
1816	GGTTTATGTTCGTCTACCTCGTGTTCTTGTTTGGATTTTCCACAGCTGTG	1865
2898	GTGACGCTGATTGAAGACGGGAAGAATGACTCCCTGCCGTCTGAGTCCAC	2947
1866	GTGACACTGATTGAGGATGGGAAGAATAACTCTCTGCCTATGGAGTCCAC	1915
2948	GTCGCACAGGTGGCGGGGCCTGCCTGCAGGCCCCCGATAGCTCCTACA	2997
	ACCACACAAGTGCCGGGGGTCTGCCTGCAAGCCAGGTAACTCTTACA	
	ACAGCCTGTACTCCACCTGCCTGGAGCTGTTCAAGTTCACCATCGGCATG	
	ACAGCCTGTATTCCACATGTCTGGAGCTGTTCAAGTTCACCATCGGCATG	
	GGCGACCTGGAGTTCACTGAGAACTATGACTTCAAGGCTGTCTTCATCAT	
	GGCGACCTGGAGTTCACTGAGAACTACGACTTCAAGGCTGTCTTCATCAT CCTGCTGCTGGCCTATGTAATTCTCACCTACATCCTCCTGCTCAACATGC	
	CCTGTTACTGGCCTATGTGATTCTCACCTACATCCTCCTGCTCAACATGC	
3148	TCATCGCCCTCATGGGTGAGACTGTCAACAAGATCGCACAGGAGAGCAAG	3197
2113	TCATTGCTCTCATGGGTGAGACCGTCAACAAGATTGCACAAGAGAGCAAG	2162
3198	AACATCTGGAAGCTGCAGAGAGCCATCACCATCCTGGACACGGAGAAGAG	3247
2163	AACATCTGGAAGCTGCAGAGAGCCATCACCATCCTGGATACAGAGAAGAG	2212
3248	CTTCCTTAAGTGCATGAGGAAGGCCTTCCGCTCAGGCAAGCTGCTGCAGG	3297
2213	CTTCCTGAAGTGCATGAGGAAGGCCTTCCGCTCTGGCAAGCTGCTGCAGG	2262
	TGGGGTACACACCTGATGGCAAGGACGACTACCGGTGGTGCTTCAGGGTG	
	TGGGGTTCACTCCTGACGGCAAGGATGACTACCGGTGGTGTTTCAGGGTG	
	GACGAGGTGAACTGGACCACCTGGAACACCAACGTGGGCATCATCAACGA	3397
2313	GACGAGGTAAACTGGACTACCTGGAACACCAATGTGCCCTATCATCATCATCA	2262

Fig. 9 continued

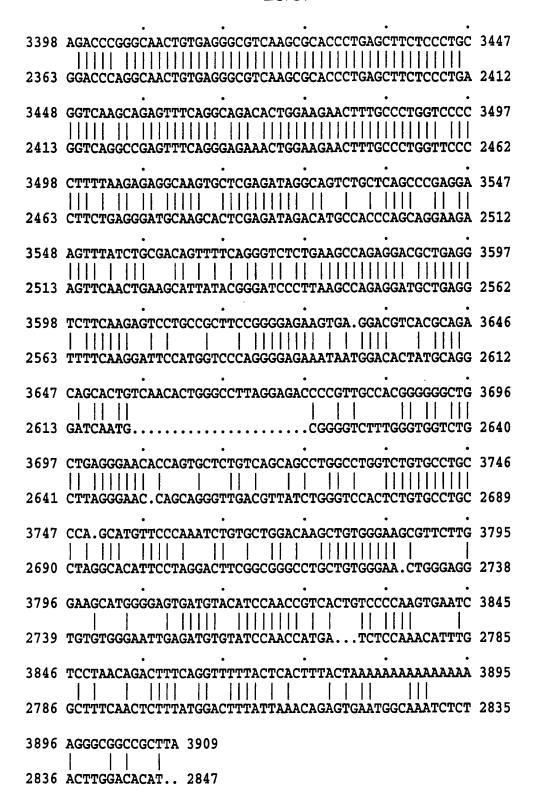


Fig. 9 continued

GAP of: humanvrl.pep check: 6877 from: 1 to: 839 humanVR1 _Fbh18547pat - fchrb87a6, 3909 bases, 4554 checksum. to: ratvrl.pep check: 5764 from: 1 to: 838 ratVR1 | AF029310 Rattus norvegicus vanilloid receptor subtype 1 mRNA, complete cds. Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430 Gap Weight: 12 Average Match: 2.912 Length Weight: 4 Average Mismatch: -2.003 Quality: 3734 Length: 840 Ratio: 4.456 Gaps: Percent Similarity: 89.247 Percent Identity: 86.022 Match display thresholds for the alignment(s): = IDENTITY humanvr1.pep x ratvr1.pep 1 MKKWSSTDLGTAADPLOKDTCPDPLDGDPNSRPPPAKPOLPTAKSRTRLF 50 1 MEQRASLDSEESESPPQENSCLDPPDRDPNCKPPPVKPHIFTTRSRTRLF 50 51 GKGDSEEAFPVDCPHEEGELDSCPTITVSPVITIQRPGDGPTGARLLSQD 100 51 GKGDSEEASPLDCPYEEGGLASCPIITVSSVLTIQRPGDGPASVRPSSQD 100 101 SVAASTEKTLRLYDRRSIFEAVAONNCQDLESLLLFLQKSKKHLTDNEFK 150 101 SVSAG.EKPPRLYDRRSIFDAVAQSNCQELESLLPFLQRSKKRLTDSEFK 149 151 DPETGKTCLLKAMLNLHDGQNTTIPLLLEIARQTDSLKELVNASYTDSYY 200 150 DPETGKTCLLKAMLNLHNGONDTIALLLDVARKTDSLKOFVNASYTDSYY 199 201 KGQTALHIAIERRNMALVTLLVENGADVQAAAHGDFFKKTKGRPGFYFGE 250 200 KGQTALHIAIERRNMTLVTLLVENGADVQAAANGDFFKKTKGRPGFYFGE 249

251	LPLSLAACTNQLGIVKFLLQNSWQTADISARDSVGNTVLHALVEVADNTA	300
250	LPLSLAACTNQLAIVKFLLQNSWQPADISARDSVGNTVLHALVEVADNTV	299
301	DNTKFVTSMYNEILMLGAKLHPTLKLEELTNKKGMTPLALAAGTGKIGVL	350
300		349
351	AYILQREIQEPECRHLSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI	400
350	AYILQREIHEPECRHLSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI	399
401	AYSSSETPNRHDMLLVEPLNRLLQDKWDRFVKRIFYFNFLVYCLYMIIFT	450
	AYSSSETPNRHDMLLVEPLNRLLQDKWDRFVKRIFYFNFFVYCLYMIIFT	
	MAAYYRPVDGLPPFKMEK.IGDYFRVTGEILSVLGGVYFFFRGIQYFLQR	
	AAAYYRPVEGLPPYKLKNTVGDYFRVTGEILSVSGGVYFFFRGIQYFLQR	
	RPSMKTLFVDSYSEMLFFLQSLFMLATVVLYFSHLKEYVASMVFSLALGW	
500	RPSLKSLFVDSYSEILFFVQSLFMLVSVVLYFSQRKEYVASMVFSLAMGW	549
550	TNMLYYTRGFQQMGIYAVMIEKMILRDLCRFMFVYIVFLFGFSTAVVTLI	599
550	TNMLYYTRGFQQMGIYAVMIEKMILRDLCRFMFVYLVFLFGFSTAVVTLI	599
600	EDGKNDSLPSESTSHRWRGPACRPPDSSYNSLYSTCLELFKFTIGMGDLE	649
600	EDGKNNSLPMESTPHKCRGSACK.PGNSYNSLYSTCLELFKFTIGMGDLE	648
650	FTENYDFKAVFIILLLAYVILTYILLLNMLIALMGETVNKIAQESKNIWK	699
649	FTENYDFKAVFIILLLAYVILTYILLLNMLIALMGETVNKIAQESKNIWK	698
700	LQRAITILDTEKSFLKCMRKAFRSGKLLQVGYTPDGKDDYRWCFRVDEVN	749
699	LQRAITILDTEKSFLKCMRKAFRSGKLLQVGFTPDGKDDYRWCFRVDEVN	748
750	WTTWNTNVGIINEDPGNCEGVKRTLSFSLRSSRVSGRHWKNFALVPLLRE	799
749	WTTWNTNVGIINEDPGNCEGVKRTLSFSLRSGRVSGRNWKNFALVPLLRD	798
800	ASARDRQSAQPEEVYLRQFSGSLKPEDAEVFKSPAASGEK 839	
799	ASTRORHATQQEEVQLKHYTGSLKPEDAEVFKDSMVPGEK 838	

Fig. 10 continued

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CLUSTAL W (1.74) multiple sequence alignment

humanVR2.alt human VR2	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY
humanVR2.alt human VR2	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2.alt human VR2	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK
humanVR2.alt human VR2	GRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA
humanVR2.alt human VR2	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL
humanVR2.alt human VR2	AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF
humanVR2.alt human VR2	HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFDNFLCNLIYMFIFTAVAYHQPTLKKQAA HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA
humanVR2.alt human VR2	PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL PHLKAEVGNSMLLTGHYLILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL
humanVR2.alt human VR2	TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQTVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLL
humanVR2.alt human VR2	IYLVFLFGFAVALVSLSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLEL
humanVR2.alt human VR2	FKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETVNSVATDSWSIW
humanVR2.alt human VR2	KKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPT KLQKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPT :************************************
humanVR2.alt human VR2	LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN ************************************

Fig. 11

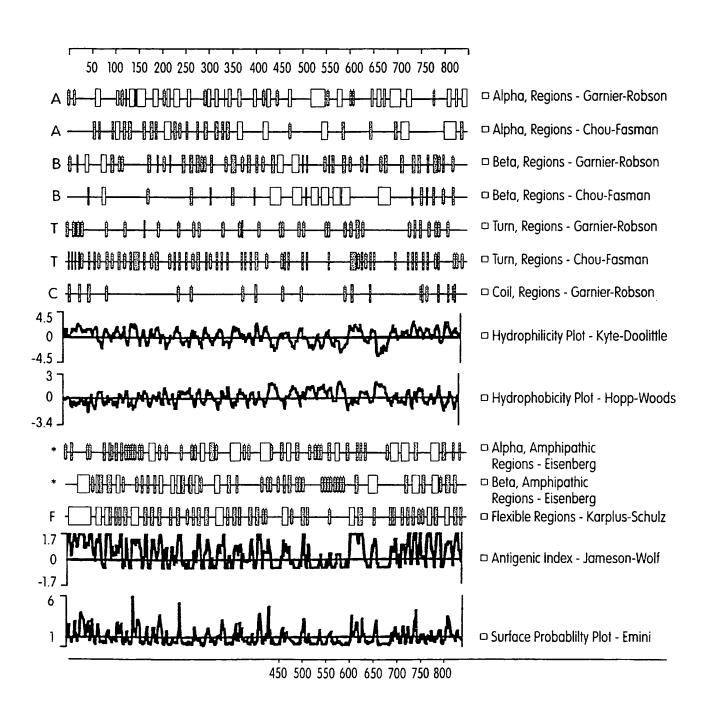


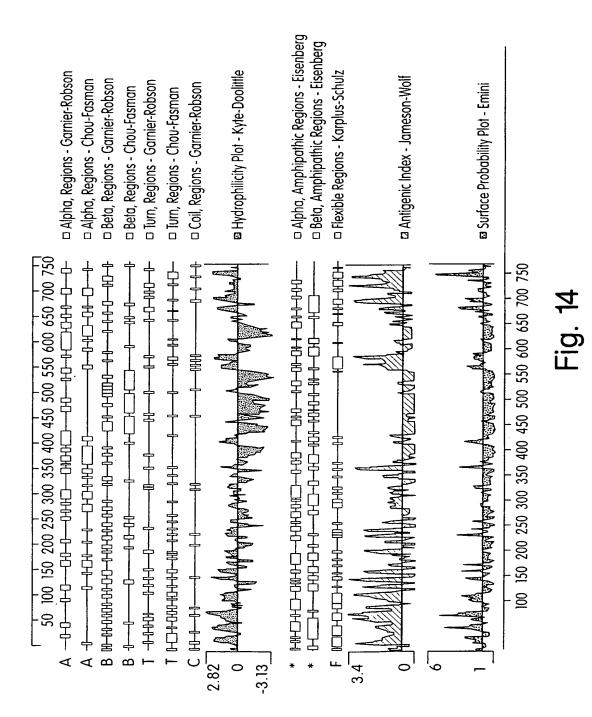
Fig. 12

33/37

Protein Family / Domain Matches, HMMer version 2

Searching for complete domains hmmpfam - search a single seg against HMM database HMMER 2.1.1 (Dec 1998) Copyright (C) 1992-1998 Washington University School of Medicine HMMER is freely distributed under the GNU General Public License (GPL). HMM file: /prod/ddm/seganal/PFAM/pfam4.2/Pfam Sequence file: /usr/ns-home/docs/seqanal/orfanal/oa-script 18670 seq Query: hVR-1 Scores for sequence family classification (score includes all domains): Model Description Score ------51.5 ank Ank repeat 1.9e-11 3 Parsed for domains: Model Domain seq-f seq-t hmm-f hmm-t score E-value ank 1/3 201 233 .. 1 33 [] ank 2/3 248 283 .. 1 33 [] ank 3/3 333 361 .. 1 33 [] 34.4 2.6e-06 13.2 2 3.4 26 Alignments of top-scoring domains: ank: domain 1 of 3, from 201 to 233: score 34.4, E = 2.6e-06 *->nGnTPLHlAarygnvevvklLLehGAdvnartk<-* +G+T+LH+A + n+ +v 1L+e+GAdv a+ hVR-1 201 KGQTALHIAIERRNMALVTLLVENGADVQAAAH 233 ank: domain 2 of 3, from 248 to 283: score 13.2, E = 2 *->nGnTPLHlAarygnvevvklLLe...hGAdvnartk<-* G PL 1Aa ++++ +vk+LL+++ + Ad+ ar+ hVR-1 248 FGELPLSLAACTNQLGIVKFLLQnswQTADISARDS 283 ank: domain 3 of 3, from 333 to 361: score 3.4, E = 26*->nGnTPLHlAarygnvevvklLLehGAdvnartk<-* +G TPL 1Aa +g++ v ++ L+ ++ hVR-1 333 KGMTPLALAAGTGKIGVLAYILO----REIOEP 361

34/37



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35/37

Protein Family / Domain Matches, HMMer version 2 Searching for complete domains hmmpfam - search a single seq against HMM database HMMER 2.1.1 (Dec 1998) Copyright (C) 1992-1998 Washington University School of Medicine HMMER is freely distributed under the GNU General Public License (GPL). HMM file: /prod/ddm/seqanal/PFAM/pfam4.2/Pfam Sequence file: /tmp/orfanal.5/g.aa HMM file: Ouery: Flh21e11 Scores for sequence family classification (score includes all domains): Model Description Score E-value -----PF00023 Ank repeat ank 53.7 4e-12 3 Parsed for domains: Model Domain seq-f seq-t hmm-f hmm-t score E-value ------
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 243
 1
 33 []
 6.4

 ank
 3/3
 293
 328
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 38.3 1.7e-07 6.4 4.3 2.1 Alignments of top-scoring domains: ank: domain 1 of 3, from 162 to 194: score 38.3, E = 1.7e-07 *->nGnTPLHlAarygnvevvklLLehGAdvnartk<-* +G+++LH+A ++ ++++vklL+e+GA+v+ar Flh21e11 162 RGHSALHIAIEKRSLQCVKLLVENGANVHARAC 194 ank: domain 2 of 3, from 208 to 243: score 6.4, E = 4.3 *->nGnTPLHlAarygnvavvklLLe...hGAdvnartk<-* G PL lAa + +++vv +LLe++++ A+ a++ Flh21e11 208 FGELPLSLAACTKQWDVVSYGLEnphQPASLQATDS 243 ank: domain 3 of 3, from 293 to 328: score 8.8, E = 2.1 *->nGnTPLHLAarygnvevvklLLe...hGAdvnart<<-* + +TPL lAa++g++e+ + L+++ G + +r Flh21e11 293 QDLTPLKLAAKEQKLEIFRHILOrafSGLSHLSRK 328

Fig. 15

LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQATSQGNTVLHALVM RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK **QREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAFHCKSPHRHRMVVLE IVGTKPDGSPDERWCFRVEEVNWASWEQTLPTLCEDPSGAGVPRTLENPVLASPPKEDED** MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY ISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKLAAKEGKIEIFRHIL PLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAAPHLKAEVGNSMLLT GHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEW YLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKKAISVLEMENGYWWCRKKQRAGVML >hvR2.altFL (full-length predicted) GASEENYVPVQLLQSN

CLUSTAL W (1.74	1) multiple sequence alignment
humanVR2 hVR2.altFL	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY
humanVR2 hVR2.altFL	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2 hVR2.altFL	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK ************************************
humanVR2 hVR2.altFL	LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA ************************************
humanVR2 hVR2.altFL	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL
humanVR2 hVR2.altFL	AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF ***********************************
humanVR2 hVR2.altFL	HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA **********************************
humanVR2 hVR2.altFL	PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL
humanVR2 hVR2.altFL	TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLL TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQK
humanVR2 hVR2.altFL	IYLVFLFGFAVALVSLSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLEL
humanVR2 hVR2.altFL	FKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETVNSVATDSWSIW
humanVR2 hVR2.altFL	KLQKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPTKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPT ***********************************
humanVR2 hVR2.altFL	LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN ************************************

Fig. 17

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											agc Ser					2333

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His	Pro	Thr	Leu	Lys 325	Leu	Glu	Glu	Leu	Thr 330	Asn	Lys	Lys	Gly	Met 335	Thr
Pro	Leu	Ala	Leu 340	Ala	Ala	Gly	Thr	Gly 345	Lys	Ile	Gly	Val	Leu 350	Ala	Tyr
Ile	Leu	Gln 355	Arg	Glu	Ile	Gln	Glu 360	Pro	Glu	Cys	Arg	His 365	Leu	Ser	Arg
Lys	Phe 370	Thr	Glu	Trp	Ala	Tyr 375	Gly	Pro	Val	His	Ser 380	Ser	Leu	Tyr	Asp
Leu 385	Ser	Cys	Ile	Asp	Thr 390	Cys	Glu	Lys	Asn	Ser 395	Val	Leu	Glu	Val	Ile 400
Ala	Tyr	Ser	Ser	Ser 405	Glu	Thr	Pro	Asn	Arg 410	His	Asp	Met	Leu	Leu 415	Val
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Phe	Thr 450	Met	Ala	Ala	Tyr	Tyr 455	Arg	Pro	Val	Asp	Gly 460	Leu	Pro	Pro	Phe
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Gln	Met	Gly	, Ile	Tyr 565		Val	. Met	Ile	Glu 570	_	Met	Ile	e Leu	Arg 575	_
Leu	Cys	Arg	Phe 580		Phe	Val	Tyr	Ile 585		. Phe	. Leu	Phe	Gly 590	Phe	Sex
Thr	Ala	Val 595		Thr	Leu	Ile	Glu 600	-	Gly	Lys	: Asn	Asp 605		Leu	Pro
Ser	Glu 610	Ser	Thr	Ser	His	Arg 615		Arg	Gly	Pro	Ala 620		Arg	Pro	Pro
Asp 625	Ser	Ser	Tyr	Asn	Ser 630	Leu	Tyr	Ser	Thr	Cys 635		Glu	Leu	Phe	Lys 640
Phe	Thr	Ile	Gly	Met 645	Gly	Asp	Leu	Glu	Phe 650		Glu	Asn	Tyr	Asp 655	Phe
Lys	Ala	Val	Phe 660	Ile	Ile	Leu	Leu	Leu 665	Ala	Tyr	Val	Ile	Leu 670	Thr	Tyr
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Lys	Ile 690	Ala	Gln	Glu	Ser	Lys 695	Asn	Ile	Trp	Lys	Leu 700	Gln	Arg	Ala	Ile
Thr 705	Ile	Leu	Asp	Thr	Glu 710	Lys	Ser	Phe	Leu	Lys 715	Cys	Met	Arg	Lys	Ala 720
Phe	Arg	Ser	Gly	Lys 725	Leu	Leu	Gln	Val	Gly 730	Tyr	Thr	Pro	Asp	Gly 735	Lys
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gcc Ala	atc Ile 210	Glu	aga Arg	cgc Arg	aac Asn	atg Met 215	Ala	ctg Leu	gtg Val	g acc	cto Leu 220	Leu	gtç Val	g gag . Glu	aac Asn	672
gga Gly 225	gca Ala	gac Asp	gtc Val	cag Gln	gct Ala 230	Ala	gcc Ala	cat His	ggg Gly	gac Asp 235	Phe	ttt Phe	aag Lys	aaa Lys	acc Thr 240	720
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gcc Ala	tac Tyr	agc Ser	Ser	agc Ser 405	gag Glu	acc Thr	cct Pro	aat Asn	cgc Arg 410	cac His	gac Asp	atg Met	ctc Leu	ttg Leu 415	gtg Val	1248

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aca Thr	gcg Ala	gtg Val 595	gtg Val	acg Thr	ctg Leu	att Ile	gaa Glu 600	gac Asp	Gly ggg	aag Lys	aat Asn	gac Asp 605	tcc Ser	ctg Leu	ccg Pro	1824
					cac His											1872
gat Asp 625	agc Ser	tcc Ser	tac Tyr	aac Asn	agc Ser 630	ctg Leu	tac Tyr	tcc Ser	acc Thr	tgc Cys 635	ctg Leu	gag Glu	ctg Leu	ttc Phe	aag Lys 640	1920
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Phe T	Thr	Ile	Gly	Met 645	Gly	Asp	Leu	Glu	Phe 650		Glu	Asn	Tyr	Asp 655		
aag o Lys A																2016
atc o		_			_			_		_				_		2064
aag a Lys I		_	_	-	_	_				_	_	_	_	_		2112
acc a Thr I 705		_	-	_		_	_			_	_	_		_	_	2160
ttc c Phe A																2208
gac g Asp A																2256
tgg a Trp A	Asn															2304
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		Arg					Gly					Phe		ttc Phe	984
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					aca Thr										1128
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					cgc Arg										1224
					ctc Leu 295										1272
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					ctg Leu										1416
					gcc Ala										1464
					ccc Pro 375										1512
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	agc Ser															2376	
	ggc Gly															2424	
	gtt Val 690															2472	
	gtg Val															2520	
	tgt Cys															2568	
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As ₁		Le د	Il د	e Pr	5 Lys		e Phe	e Lei	ı Asr	9 Phe		ı Cys	s Ası	n Leu	1 Ile 400
Туз	Met	: Phe	e Ile	e Phe 40!		Ala	a Val	L Ala	Tyr 410		Glr.	Pro	Thi	415	Lys
Lys	Glr	n Ala	a Ala 420		o His	Let	Lys	425		ı Val	Gly	' Asn	Ser 430		Leu
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Leu 625	Leu	Leu	Ala	Tyr	Val 630	Leu	Leu	Thr	Tyr	Ile 635	Leu	Leu	Leu	Asn	Met 640
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Arg Val Glu Glu Val Asn Trp Ala Ser Trp Glu Gln Thr Leu Pro Thr 705 710 715 720

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						gcc Ala		His					His		cac	1104
cga Arg	atg Met 370	Val	gtt Val	ttg Leu	gag Glu	ccc Pro 375	ctg Leu	aac Asn	aaa Lys	ctg Leu	ctg Leu 380	Gln	gcg Ala	aaa Lys	tgg Trp	1152
						ttc Phe										1200
						gct Ala										1248
						ctg Leu										1296
						atc Ile										1344
						tgg Trp 455										1392
						gaa Glu										1440
						ctg Leu										1488
						ctg Leu										1536
						cac His										1584
						ctg Leu 535										1632
						gta Val										1680
			Glu			aca Thr										1728
						gac Asp										1776

- 22 -

			580					585					590			
			Āla					Phe							ggc Gly	1824
												Met			ctg Leu	1872
_	_	_	gcc Ala	Tyr	-	_					_	_			atg Met 640	1920
		**	ctc Leu	_	_			-		_	-	_		_	_	1968
			tgg Trp 660													2016
			tgg Trp												-	2064
			act Thr									-		_		2112
			gag Glu				-				_	_	_		_	2160
_	_		gac Asp	_			_		_		-					2208
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aac Asn	tca Ser 65	Ala	gag Glu	aac Asn	att Ile	gca Ala 70	Leu	gtg Val	acc Thr	agc Ser	atg Met 75	Tyr	gat Asp	ggg	ctc Leu	239
	Gln	gct Ala									Gln					287
		ctg Leu														335
		gag Glu														383
		ctt Leu 130														431
		ctg Leu														479
	Leu	gag Glu														527
_	-	gtt Val	_													575
		atc Ile														623
		atc Ile 210														671
		gcc Ala														719

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cag Gln	ctg Leu	tgg Trp	tac Tyr	ttc Phe 260	: Trp	ogg Arg	g cgc Arg	cac His	gtg Val 265	. Phe	atc lle	tgç Trp	g ato	tcg Ser 270	ttc Phe	815
ata Ile	gac Asp	agc Ser	tac Tyr 275	Phe	gaa Glu	atc	ctc Leu	ttc Phe 280	Leu	ttc Phe	cag Gln	gcc Ala	ctg Leu 285	Leu	aca Thr	863
gtg Val	gtg Val	tcc Ser 290	cag Gln	gtg Val	ctg Leu	tgt Cys	ttc Phe 295	Leu	gcc Ala	atc Ile	gag Glu	tgg Trp 300	Tyr	ctg Leu	ccc Pro	911
ctg Leu	ctt Leu 305	gtg Val	tct Ser	gcg Ala	ctg Leu	gtg Val 310	ctg Leu	ggc Gly	tgg Trp	ctg Leu	aac Asn 315	ctg Leu	ctt Leu	tac Tyr	tat Tyr	959
aca Thr 320	cgt Arg	ggc Gly	ttc Phe	cag Gln	cac His 325	aca Thr	Gly	atc Ile	tac Tyr	agt Ser 330	gtc Val	atg Met	atc Ile	cag Gln	aag Lys 335	1007
aaa Lys	gcc Ala	atc Ile	tct Ser	gtc Val 340	ctg Leu	gag Glu	atg Met	gag Glu	aat Asn 345	ggc Gly	tat Tyr	tgg Trp	tgg Trp	tgc Cys 350	agg Arg	1055
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gct Ala	tca Ser 385	tgg Trp	gag Glu	cag Gln	acg Thr	ctg Leu 390	cct Pro	acg Thr	ctg Leu	tgt Cys	gag Glu 395	gac Asp	ccg Pro	tca Ser	G] À ààà	1199
gca (Ala (400	ggt Gly	gtc Val	cct Pro	cga Arg	act Thr 405	ctc Leu	gag Glu	aac Asn	Pro	gtc Val 410	ctg Leu	gct Ala	tcc Ser	cct Pro	ccc Pro 415	1247
aag (Lys (gag Glu	gat Asp	Glu	gat Asp 420	ggt Gly	gcc Ala	tct Ser	Glu	gaa Glu 425	aac Asn	tat Tyr	gtg Val	Pro	gtc Val 430	cag Gln	1295
ctc d Leu I	ctc Leu (Gln	tcc Ser 435	aac Asn	tgat	ggcc	ca g	atgc	agca	g ga	ggcc	agag	gac	agag	cag	1350
aggat	ctt	tc c	aacc	acat	c tg	ctgg	ctct	ggg	gtcc	cag	tgaa	ttct	gg t	ggca	aatat	1410
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1489

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Leu Trp Tyr Phe Trp Arg Arg His Val Phe Ile Trp Ile Ser Phe Ile

			260)				265	5				270)		
Asp	Ser	Tyr 275		e Glu	ı Ile	e Leu	280		n Ph∈	e Glr	n Ala	a Leu 285		ı Thr	Val	
Val	Ser 290		Val	. Leu	Суз	295		ı Ala	ıle	e Glu	300		Leu	Pro	Leu	
Leu 305		Ser	Ala	Leu	Val 310	Leu)	Gly	7 Trp	Leu	Asr 315		ı Lev	Туг	Tyr	Thr 320	
Arg	Gly	Phe	Gln	His 325	Thr	Gly	, Ile	Tyr	Ser 330		. Met	: Ile	Gln	Lys 335		
Ala	Ile	Ser	Val 340	Leu	Glu	Met	Glu	Asn 345		Tyr	Trp	Trp	Cys 350		Lys	,
Lys	Gln	Arg 355	Ala	Gly	Val	Met	Leu 360		Val	Gly	Thr	Lys 365		Asp	Gly	
Ser	Pro 370	Asp	Glu	Arg	Trp	Cys 375	Phe	Arg	Val	Glu	Glu 380		Asn	Trp	Ala	
Ser 385	Trp	Glu	Gln	Thr	Leu 390	Pro	Thr	Leu	Cys	Glu 395	Asp	Pro	Ser	Gly	Ala 400	
Gly	Val	Pro	Arg	Thr 405	Leu	Glu	Asn	Pro	Val 410	Leu	Ala	Ser	Pro	Pro 415	Lys	
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cta Leu																96
tac Tyr	ctc Leu	ctg Leu 35	gag Glu	aac Asn	cca Pro	cac His	cag Gln 40	ccc Pro	gcc Ala	agc Ser	ctg Leu	cag Gln 45	gcc Ala	act Thr	gac Asp	14

		Gly					ı His					t Ile			c aac o Asn	192
	Ala					a Lev					t Ty				c ctc Leu 80	240
					J Let					l Glr					cgc Arg	288
				Let					Let					Gly	aag Lys	336
								Gln					Gly		agc Ser	384
cac His	ctt Leu 130	tcc Ser	cga Arg	aag Lys	ttc Phe	acc Thr 135	gag Glu	tgg Trp	tgc Cys	tat Tyr	999 Gly 140	Pro	gtc Val	cgg Arg	gtg Val	432
tcg Ser 145	ctg Leu	tat Tyr	gac Asp	ctg Leu	gct Ala 150	tct Ser	gtg Val	gac Asp	agc Ser	tgt Cys 155	Glu	gag Glu	aac Asn	tca Ser	gtg Val 160	480
ctg Leu	gag Glu	atc Ile	att Ile	gcc Ala 165	ttt Phe	cat His	tgc Cys	aag Lys	agc Ser 170	ccg Pro	cac His	cga Arg	cac His	cga Arg 175	atg Met	528
		ttg Leu														576
		ccc Pro 195														624
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gcc Ala 225	gcc Ala	cct Pro	cac His	ctg Leu	aaa Lys 230	gcg Ala	gag Glu	gtt Val	gga Gly	aac Asn 235	tcc Ser	atg Met	ctg Leu	ctg Leu	acg Thr 240	720
ggc Gly	cac His	atc Ile	ctt Leu	atc Ile 245	ctg Leu	cta Leu	Gly ggg	ggg Gly	atc Ile 250	tac Tyr	ctc Leu	ctc Leu	gtg Val	ggc Gly 255	cag Gln	768
ctg Leu	tgg Trp	Tyr	ttc Phe 260	tgg Trp	cgg Arg	cgc Arg	cac His	gtg Val 265	ttc Phe	atc Ile	tgg Trp	atc Ile	tcg Ser 270	ttc Phe	ata Ile	816
gac a Asp	agc Ser	tac Tyr	ttt Phe	gaa Glu	atc Ile	ctc Leu	ttc Phe	ctg Leu	ttc Phe	cag Gln	gcc Ala	ctg Leu	ctc Leu	aca Thr	gtg Val	864

- 28 -

		275					280				285				
												ctg Leu			912
												tac Tyr			960
												cag Gln			1008
-			_	_		_						tgc Cys 350		_	1056
	_		_			-	_		_		_	cca Pro	_		1104
												aac Asn			1152
		_	_	-	_		_	-	-	 -	_	tca Ser		_	1200
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	-		Asp		_			Thr	_	-	•	_	Leu	_	atg Met	145
	-	Asp		_			Asn	_	-	-		Ile		_	tac Tyr	193
-	-				_			_		_	Pro			_	ctt Leu 80	241
								ctc Leu								289
								agg Arg 105								337
								cga Arg								385
_								gac Asp					_	_		433
-	-		_		_		_	atc Ile				_	-	-	_	481
								gaa Glu								529
_			_			-		aga Arg 185						_	_	577
						Ile		acc Thr								625
cc	ctg	gat	cag	сса	gcc	atc	ccc	tca	tca	aaa	gcg	act	ttt	ggg	gaa	673

WO 00/29577

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tcc Ser 225	Met	g ctg Leu	cto Lev	g cto 1 Lei	g ggc 1 Gly 230	, His	att Ile	ctg Leu	g ato	cto E Leu 235	ı Let	ggg Gly	g ggt y Gly	ati Ile	tac Tyr 240	721
ctc Leu	tta Leu	ctg Leu	ggc	cag Glr 245	Leu	tgo Trp	tac Tyr	ttt Phe	tgc Trp 250	Arg	g cgo	g cgo g Aro	cto g Leu	ttt Phe 255	atc Elle	769
tgg Trp	ato	tca Ser	tto Phe 260	Met	gac Asp	ago Ser	tac Tyr	ttt Phe 265	Glu	ato Ile	cto Lev	ttt Phe	cto Leu 270	Leu	cag Gln	817
gct Ala	ctg Leu	ctc Leu 275	aca Thr	gtg Val	ctg Leu	tcc Ser	cag Gln 280	gtg Val	ctg Leu	cgc Arg	ttc Phe	atg Met 285	Glu	act Thr	gaa Glu	865
tgg Trp	tac Tyr 290	Leu	ccc Pro	ctg Leu	cta Leu	gtg Val 295	tta Leu	tcc Ser	cta Leu	gtg Val	ctg Leu 300	Gly	tgg Trp	ctg Leu	aac Asn	913
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tac Tyr	ctg Leu	gtc Val	ttc Phe 340	ctt Leu	ttc Phe	ggc Gly	ttt Phe	gct Ala 345	gta Val	gcc Ala	cta Leu	gta Val	agc Ser 350	ttg Leu	agc Ser	1057
aga Arg	gag Glu	gcc Ala 355	cga Arg	agt Ser	ccc Pro	aaa Lys	gcc Ala 360	cct Pro	gaa Glu	gat Asp	aac Asn	aac Asn 365	tcc Ser	aca Thr	gtg Val	1105
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agc Ser 385	att Ile	ctg Leu	gat Asp	gcc Ala	tcc Ser 390	cta Leu	gag Glu	ctg Leu	ttc Phe	aag Lys 395	ttc Phe	acc Thr	att Ile	ggt Gly	atg Met 400	1201
Gly	gag Glu	ctg Leu	gct Ala	ttc Phe 405	cag Gln	gaa Glu	cag Gln	Leu	cgt Arg 410	ttt Phe	cgt Arg	Gly ggg	gtg Val	gtc Val 415	ctg Leu	1249
ctg Leu	ttg Leu	Leu	ttg Leu 420	gcc Ala	tac Tyr	gtc Val	ctt Leu	ctc Leu 425	acc Thr	tac Tyr	gtc Val	ctg Leu	ctg Leu 430	ctc Leu	aac Asn	1297
atg (Leu	att Ile 435	gct Ala	ctc Leu	atg Met	agc Ser	gaa Glu 440	act Thr	gtc Val	aac Asn	cac His	gtt Val 445	gct Ala	gac Asp	aac Asn	1345

					aag Lys											1393
	Asn				tgg Trp 470	_			_			-	_			1441
					acc Thr											1489
					gaa Glu											1537
					gat Asp											1585
					aaa Lys											1633
	_			-	gtc Val 550		_			tgat	ggco	ca ç	gatgo	cagca	ag	1683
cag	gctg	jca g	gato	ggagt	a go	gaat	ctto	c cca	agcca	acac	caga	ıggct	ac t	gaat	tttgg	1743
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Lys	Glu	Gly	100		e Glu	ı Ile	Phe	Arg 105		s Ile	e Leu	ı Glr	110		ı Phe
Ser	Gly	Pro 115		Glr	Pro	Leu	Ser 120		Lys	s Phe	Thr	Glu 125		Cys	туі
Gly	Pro 130		Arg	y Val	. Ser	Leu 135	_	Asp	Leu	ser	Ser 140	Val	. Asp	Ser	Trp
Glu 145	Lys	Asn	Ser	Val	Leu 150		Ile	Ile	Ala	Phe 155		Cys	: Lys	Ser	Pro 160
Asn	Arg	His	Arg	Met 165		Val	Leu	Glu	Pro 170		Asn	Lys	Leu	Leu 175	
Glu	Lys	Trp	Asp 180		Leu	Val	Ser	Arg 185	Phe	Phe	Phe	Asn	Phe 190		Cys
туг	Leu	Val 195		Met	Phe	Ile	Phe 200	Thr	Val	Val	Ala	Tyr 205		Gln	Pro
Ser	Leu 210	Asp	Gln	Pro	Ala	Ile 215	Pro	Ser	Ser	Lys	Ala 220	Thr	Phe	Gly	Glu
Ser 225	Met	Leu	Leu	Leu	Gly 230	His	Ile	Leu	Ile	Leu 235	Leu	Gly	Gly	Ile	Tyr 240
Leu	Leu	Leu	Gly	Gln 245	Leu	Trp	Tyr	Phe	Trp 250	Arg	Arg	Arg	Leu	Phe 255	Ile
Trp	Ile	Ser	Phe 260	Met	Asp	Ser	Tyr	Phe 265	Glu	Ile	Leu	Phe	Leu 270	Leu	Gln
Ala	Leu	Leu 275	Thr	Val	Leu	Ser	Gln 280	Val	Leu	Arg	Phe	Met 285	Glu	Thr	Glu
Trp	Tyr 290	Leu	Pro	Leu	Leu	Val 295	Leu	Ser	Leu	Val	Leu 300	Gly	Trp	Leu	Asn
Leu 305	Leu	Tyr	Tyr	Thr	Arg 310	Gly	Phe	Gln	His	Thr 315	Gly	Ile	Tyr	Ser	Val 320
Met	Ile	Gln	Lys	Val 325	Ile	Leu	Arg	Asp	Leu 330	Leu	Arg	Phe	Leu	Leu 335	Val
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Arg	Glu	Ala 355	Arg	Ser	Pro	Lys	Ala 360	Pro	Glu	Asp	Asn	Asn 365	Ser	Thr	Val
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Gly	Glu	Leu	Ala	Phe 405	Gln	Glu	Gln	Leu	Arg 410	Phe	Arg	Gly	Val	Val 415	Leu

Leu	Leu	Leu	Leu 420		Tyr	Val	. Leu	425		Туг	r Va]	Leu	430		Asn	
Met	Leu	Ile 435		Leu	Met	Ser	Glu 440		· Val	. Asr	n His	Val 445		Asp	Asn	
Ser	Trp 450		Ile	Trp	Lys	Leu 455		Lys	: Ala	ıle	Ser 460		Leu	Glu	Met	
Glu 465		Gly	Tyr	Trp	Trp 470	-	Arg	Arg	Lys	475		Arg	Glu	Gly	Arg 480	
Leu	Leu	Lys	Val	Gly 485	Thr	Arg	Gly	Asp	Gly 490		Pro	Asp	Glu	Arg 495	Trp	
Cys	Phe	Arg	Val 500	Glu	Glu	Val	Asn	Trp 505		Ala	Trp	Glu	Lys 510	Thr	Leu	
Pro	Thr	Leu 515	Ser	Glu	Asp	Pro	Ser 520	Gly	Pro	Gly	Ile	Thr 525	Gly	Asn	Lys	
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												Thr				40
												ccg Pro				96
									-	_		gct Ala 45	-	_	-	144
												atc Ile				192
												act Thr				240

WO 00/29577

gag Glu	gaa Glu	a ato	c tco e Se:	c aa r Ası 8	n Hi	c caa s Glr	a gg	c cto y Le	c aca u Th: 90	r Pr	c cto o Le	g aaa u Lys	a ct s Le	u Al	c gcc a Ala 5	288
aag Lys	gaa Glu	a ggo	c aaa y Lys 100	s Ile	e gaq	g att	tte Phe	e Ard	g His	c att	t cto	g caq u Glr	g cg n Are	g Gl	a ttc u Phe	336
tca Ser	gga Gly	Pro	о Туг	c cad	g cco	ctt Leu	tco Ser 120	: Ar	a aaq g Lys	g tti s Phe	t act e Thi	gaç Glu 125	Tr	g tg o Cy	t tac s Tyr	384
ggt Gly	cct Pro 130	Val	g cgg . Arg	g gta g Val	tco Ser	ctg Leu 135	Туг	gac Asp	cto Leu	g tco Ser	tct Ser 140	. Val	gad Asp	c ago Se:	c tgg r Trp	432
gaa Glu 145	aag Lys	aac Asn	tcg Ser	gtg Val	Leu 150	Glu	ato	ato : Ile	gct Ala	ttt Phe 155	e His	tgc Cys	aac Lys	g ago	c ccg Pro 160	480
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ctg d Leu I 305	ctt Leu	tac Tyr	tac Tyr	Thr	cgg Arg 310	ggc Gly	ttt Phe	cag Gln	His	aca Thr 315	ggc Gly	atc Ile	tac Tyr	agt Ser	gtc Val 320	960

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INTERNATIONAL SEARCH REPORT

Intern nal Application No

PCT/US 99/26701 A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/12 C12N5/10 C07K14/705 C07K16/28 G01N33/53 C12Q1/68 A61P25/00 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 7 C12N C07K G01N C12Q A61P Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Category 9 Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X CATERINA M J ET AL: "THE CAPSAICIN 1-26 RECEPTOR: A HEAT-ACTIVATED ION CHANNEL IN THE PAIN PATHWAY" NATURE, GB, MACMILLAN JOURNALS LTD. LONDON, vol. 389, 23 October 1997 (1997-10-23), pages 816-824, XP002075020 ISSN: 0028-0836 cited in the application abstract figure 5 page 821, left-hand column, paragraph 2 P,X WO 99 09140 A (BRAKE ANTHONY ; JULIUS DAVID 1-26 J (US); UNIV CALIFORNIA (US); CATERINA) 25 February 1999 (1999-02-25) abstract 100% identity in 1717 BP overlap between SEQ ID NO 3 of W09909140 and SEQ ID NO 10 X Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents : "T" later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-*O* document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled in the art. document published prior to the international filing date but later than the priority date claimed *&* document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report

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10/05/2000

Lejeune, R

Authorized officer

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/26701

BxI	Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This Inte	emational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 24-26 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This Inte	mational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	n Protest Th additional search fees w re accompanied by th applicant's protest. No protest accompanied the payment if additional search fees.

INTERNATIONAL SEARCH REPORT

Intern Aal Application No
PCT/US 99/26701

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Category °	ation) DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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Ρ,Χ	WO 99 37675 A (BRAKE ANTHONY J ;JULIUS DAVID J (US); UNIV CALIFORNIA (US); CATERI) 29 July 1999 (1999-07-29) abstract 99.7% identity in 2533 BP overlap between SEQ ID NO 33 of W09937675 and SEQ ID NO 1 100% identity in 2380 BP overlap between SEQ ID NO 35 of W09937675 and SEQ ID NO 4 100% identity in 1717 BP overlap between SEQ ID NO 3 of W09937675 and SEQ ID NO 10	1-26
P,X	WO 99 37765 A (SMITHKLINE BEECHAM PLC) 29 July 1999 (1999-07-29) abstract 99.9% identity in 2351 BP overlap between SEQ ID NO 1 and 5 of WO9937765 and SEQ ID NO 4	1-26
Ρ,Χ	EP 0 943 683 A (SMITHKLINE BEECHAM PLC) 22 September 1999 (1999-09-22) abstract 99.6% identity in 2818 BP overlap between SEQ ID NO 1 of EP943683 and SEQ ID NO 1	1-26
Ρ,Χ	EP 0 953 638 A (SYNTHELABO) 3 November 1999 (1999-11-03) abstract 99.8% identity in 2786 BP overlap between SEQ ID NO 1 of EP953638 and SEQ ID NO 4	1-26
P,X	CATERINA ET AL: "A capsaicin-receptor homologue with a high threshold for noxious heat" NATURE,GB,MACMILLAN JOURNALS LTD. LONDON, vol. 398, no. 398, 1 April 1999 (1999-04-01), pages 436-441-441, XP002105951 ISSN: 0028-0836 the whole document 99.8% identity in 2401 BP overlap between the sequence of hVRL and SEQ ID NO 4 100% identity in 1752 BP overlap between the sequence of rVRL and SEQ ID NO 10	1-26
X	WO 98 39448 A (HUMAN GENOME SCI INC) 11 September 1998 (1998-09-11) abstract page 112 page 163 99.3% identity in 1947 BP overlap between SEQ ID NO 191 of WO9839448 and SEQ ID NO 4 98.1% identity in 1709 BP overlap between SEQ ID NO 307 of WO9839448 and SEQ ID NO 4	1-11

INTERNATIONAL SEARCH REPORT Information on patent family members

Intern (a) Application No PCT/US 99/26701

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Patent docume cited in search re		Publication date		Patent family member(s)	Publication dat
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	. A	29-07-1999	AU	2466799 A	29-07-1999 09-08-1999
·			AU WO	9115698 A 9909140 A	08-03-1999 25-02-1999
WO 9937765	A	29-07-1999	NONE		
EP 0943683	A	22-09-1999	JP	11279196 A	12-10-1999
EP 0953638	Α	03-11-1999	AU WO	2932799 A 9946377 A	27-09-1999 16-09-1999
WO 9839448	Α		NONE		

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(71) Applicant: MILLENNIUM PHARMACEUTICALS, INC. [US/US]; 75 Sidney Street, Cambridge, MA 02139 (US).

(72) Inventor: CURTIS, Rory, A., J.; 31 Constitution Drive, Southborough, MA 01772 (US).

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(54) Title: NOVEL MEMBERS OF THE CAPSAICIN/VANILLOID RECEPTOR FAMILY OF PROTEINS AND USES THEREOF

(57) Abstract

The invention provides isolated nucleic acids molecules, designated hVR-1, hVR-2, and rVR-2 nucleic acid molecules, which encode novel members of the Capsaicin/Vanilloid receptor family. The invention also provides antisense nucleic acid molecules, recombinant expression vectors containing hVR-1, hVR-2, and rVR-2 nucleic acid molecules, host cells into which the expression vectors have been introduced, and nonhuman transgenic animals in which an hVR-1, hVR-2, and rVR-2 gene has been introduced or disrupted. The invention still further provides isolated hVR-1, hVR-2, and rVR-2 proteins, fusion proteins, antigenic peptides and anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies. Diagnostic methods utilizing compositions of the invention are also provided.

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NOVEL MEMBERS OF THE CAPSAICIN/VANILLOID RECEPTOR FAMILY OF PROTEINS AND USES THEREOF

Background of the Invention

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Pain is initiated when the peripheral terminals of a subgroup of sensory neurons are activated by noxious chemical, mechanical or thermal stimuli. These neurons, called nociceptors, transmit information regarding tissue damage to pain-processing centres in the spinal chord and brain (Fields, H.L. Pain, McGraw-Hill, New York, 1987). Nociceptors are characterized in part, by their sensitivity to capsaicin, a vanilloidcontaining compound, and a natural product of capsicum peppers that is the active ingredient of many "hot" and spicy foods. In mammals, exposure of nociceptor terminals to capsaicin leads initially to excitation of the neuron and the consequent perception of pain and local release of inflammatory mediators. With prolonged exposure, nociceptor terminals become insensitive to capsaicin, as well as to other noxious stimuli (Szolcsanyi, J. in Capsaicin in the Study of Pain (ed. Wood, J.) 1-26 (Academic, London, 1993). This latter phenomenon of nociceptor desensitization underlies the seemingly paradoxical use of capsaicin as an analgesic agent in the treatment of painful disorders ranging from viral and diabetic neuropathies to rheumatoid arthritis (Campbell, E. in Capsaicin and the Studyof Pain (ed. Wood, J.) 255-272 (Academic, London, 1993); Szallasi, A. et al. (1996) Pain 68, 195-208). Some of this decreased sensitivity to noxious stimuli may result from reversible changes in the nociceptor, but the long-term loss of responsiveness can be explained by death of the nociceptor or destruction of its peripheral terminals following exposure to capsaicin (Jancso, G. et al. (1977) Nature 270, 741-743).

The cellular specificity of capsaicin action and its ability to evoke the sensation of burning pain have led to speculation that the target of capsaicin action plays an important physiological role in the detection of painful stimuli. Indeed, capsaicin may elicit the perception of pain by mimicking the actions of a physiological stimulus or an endogenous ligand produced during tissue injury (James, I.F., Kinkina, N.N. & Wood, J.N. in *Capsaicin in the Study of Pain* (ed. Wood, J.N.) 83-104 (Academic, London, 1993).

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Caterina M.J. et al. have recently determined the molecular basis underlying this phenomenon by characterizing a functional cDNA that encodes a vanilloid receptor (VR-1) in rat sensory ganglia (Caterina M. J. et al., (1997) Nature 389:816-824). VR-1 is a vanilloid-gated, nonselective cation channel that resembles members of the transient receptor potential (TRP) channel family, first identified as components of the Drosophila phototransduction pathway (Montell et al. (1989) Neuron 2:1313-1323).

Table of Contents

	A.	Sumr	nary of	the Inver	ntion	-3-			
10	B.	Brief	Descri	otion of th	he Drawings	-8-			
	C.	Detai	led Des	cription o	of the Invention	-10-			
		I.	Isola	Solated Nucleic Acid Molecules					
		II.	Isola	ted hVR-	1, hVR-2, and rVR-2 Proteins and				
		Anti- hVR-1, Anti-hVR-2, and Anti-rVR-2 Antibodies							
15		III.	Reco	mbinant i	Expression Vectors and Host Cells	-42-			
		IV.	Phan	al Compositions	-52-				
		V. Uses and Methods of the Invention							
			A.	Screen	ning Assays	-60-			
			B.	Detect	ion Assays	-67-			
20				1.	Chromosome Mapping	-67-			
				2.	Tissue Typing	-70-			
				3.	Use of Partial hVR-1, hVR-2, and rVR-2				
					Sequences in Forensic Biology	-71-			
			C.	Predic	tive Medicine	-72-			
25				1.	Diagnostic Assays	-73-			
				2.	Prognostic Assays	-75-			
				3	Monitoring of Effects During Clinical Trials	-81-			

Mathada of Treatment

D. Metho	ds of Treatment	- 83-
1.	Prophylactic Methods	-84-
2.	Therapeutic Methods	- 84-
3.	Pharmacogenomics	-85-
D. Examples		-88-

Summary of the Invention

5

10

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The present invention is based, at least in part, on the discovery of novel members of the Capsaicin/Vanilloid family of receptors. Described herein is the isolation of the human orthologue of rat VR-1 (rVR-1), referred to herein as hVR-1, as well as another previously unknown member of the VR family of receptors, referred herein as VR-2, and specifically as human VR-2 (hVR-2, including an alternate form which contains a deletion) and rat VR-2 (rVR-2) nucleic acid and protein molecules. The hVR-1, hVR-2, and rVR-2 molecules of the present invention are useful as targets for developing modulating agents to regulate a variety of cellular processes, *e.g.*, cellular processes involved in pain. Accordingly, in one aspect, this invention provides isolated nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins and fragments thereof, as well as nucleic acid fragments suitable as primers or hybridization probes for the detection of hVR-1, hVR-2, and rVR-2-encoding nucleic acids.

In one embodiment, an hVR-1, hVR-2, or rVR-2 nucleic acid molecule of the invention is at least 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identical to the nucleotide sequence (*e.g.*, to the entire length of the nucleotide sequence) shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof.

In another embodiment, the isolated nucleic acid molecule includes the nucleotide sequence shown SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof. In another embodiment, the nucleic acid molecule includes at least 10, 15, 20, or more contiguous nucleotides of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

-4-

In another embodiment, an hVR-1, hVR-2, and rVR-2 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11. In one embodiment, an hVR-1, hVR-2, and rVR-2 nucleic acid molecule includes a nucleotide sequence encoding a protein having an amino acid sequence at least 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more identical to the entire length of the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.

Another embodiment of the invention features nucleic acid molecules, preferably hVR-1, hVR-2, and rVR-2 nucleic acid molecules, which specifically detect hVR-1, hVR-2, and rVR-2 nucleic acid molecules relative to nucleic acid molecules encoding 10 non-hVR-1, non-hVR-2, and non-hVR-2 proteins. For example, in one embodiment, such a nucleic acid molecule is at least 100-150, 1150-200, 200-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, 550-600, 600-700, 700-800, 800-900, 900-1000, 1088, or more nucleotides in length and hybridizes under stringent conditions to a nucleic acid molecule comprising the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 15 6, 7, 9, 10, or 12. In preferred embodiments, the nucleic acid molecules are at least 15 (e.g., contiguous) nucleotides in length and hybridize under stringent conditions to nucleotides 1-17, 3696-3863, or 3901-3909 of SEQ ID NO:1. In other preferred embodiments, the nucleic acid molecules comprise nucleotides 1-17, 3696-3863, or 3901-3909 of SEQ ID NO:1. In yet other preferred embodiments, the nucleic acid 20 molecules consist of nucleotides 1-17, 3696-3863, or 3901-3909 of SEO ID NO:1. In preferred embodiments, the nucleic acid molecules are at least 15 (e.g., contiguous) nucleotides in length and hybridize under stringent conditions to nucleotides 1944-2003 of SEQ ID NO:4. In other preferred embodiments, the nucleic acid molecules comprise 25 nucleotides 1944-2003 of SEQ ID NO:4. In yet other preferred embodiments, the nucleic acid molecules consist of nucleotides 1944-2003 of SEQ ID NO:4.

In other embodiments, the nucleic acid molecule encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions and is encoded by the same locus as hVR-1, hVR-2 or rVR-2.

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Another embodiment of the invention provides a nucleic acid molecule that encodes a naturally occurring orthologue of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the nucleic acid molecule hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions.

Another embodiment of the invention provides an isolated nucleic acid molecule which is antisense to an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, e.g., the coding strand of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule.

Since the hVR2 (the alternate form) and rVR2 sequences represent fragments of the entire coding regions of these genes, another embodiment of the invention provides the complete gene sequences. A skilled artisan can readily isolate such molecule using the sequences disclosed herein.

Another aspect of the invention provides a vector comprising an hVR-1, an hVR-2, or a rVR-2 nucleic acid molecule. In certain embodiments, the vector is a recombinant expression vector. In another embodiment, the invention provides a host cell containing a vector of the invention. In yet another embodiment, the invention provides a host cell containing a nucleic acid molecule of the invention. The invention also provides a method for producing a protein, preferably an hVR-1, hVR-2, and rVR-2 protein, by culturing in a suitable medium, a host cell, e.g., a mammalian host cell such as a non-human mammalian cell. of the invention containing a recombinant expression vector, such that the protein is produced.

Another aspect of this invention features isolated or recombinant hVR-1, hVR-2, and rVR-2 proteins and polypeptides. In one embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain. In another embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain and at least one proline rich domain. In yet another embodiment, the isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain. In yet another embodiment, the protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain and has

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an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11. In another embodiment, the protein, preferably an hVR-1, hVR-2, or rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain and plays a role in the development and regulation of pain. In yet another embodiment, the protein, preferably an hVR-1, hVR-2, and rVR-2 protein, includes at least one transmembrane domain, at least one proline rich domain, and at least one ankyrin repeat domain and is encoded by a nucleic acid molecule having a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

In another embodiment, the invention features fragments of the protein having the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15, 30, 40, 50, 60, 70, 80, 90, or 100 amino acids (e.g., contiguous amino acids).

In another embodiment, the invention features an isolated protein, preferably an hVR-1, hVR-2, and rVR-2 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence at least about 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous to a nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof. This invention further features an isolated protein, preferably an hVR-1, hVR-2, or rVR-2 protein, which is encoded by a nucleic acid molecule consisting of a nucleotide sequence which hybridizes under stringent hybridization conditions to a nucleic acid molecule consisting of the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof.

The proteins of the present invention or portions thereof, e.g., biologically active portions thereof, can be operatively linked to a non-hVR-1, non-hVR-2, or non-rVR-2 polypeptide (e.g., heterologous amino acid sequences) to form fusion proteins. The invention further features antibodies, such as monoclonal or polyclonal antibodies, that specifically bind proteins of the invention, preferably hVR-1, hVR-2, and rVR-2 proteins. In addition, the hVR-1, hVR-2, and rVR-2 proteins or biologically active

- 7 -

portions thereof can be incorporated into pharmaceutical compositions, which optionally include pharmaceutically acceptable carriers.

In another aspect, the present invention provides a method for detecting the presence of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide in a biological sample by contacting the biological sample with an agent capable of detecting an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide such that the presence of an hVR-1, hVR-2, and rVR-2 nucleic acid molecule, protein or polypeptide is detected in the biological sample.

In another aspect, the present invention provides a method for detecting the presence of hVR-1, hVR-2, and rVR-2 activity in a biological sample by contacting the biological sample with an agent capable of detecting an indicator of hVR-1, hVR-2, and rVR-2 activity such that the presence of hVR-1, hVR-2, and rVR-2 activity is detected in the biological sample.

In another aspect, the invention provides a method for modulating hVR-1, hVR-2, and rVR-2 activity comprising contacting a cell capable of expressing hVR-1, hVR-2, and rVR-2 with an agent that modulates hVR-1, hVR-2, and rVR-2 activity such that hVR-1, hVR-2, and rVR-2 activity in the cell is modulated. In one embodiment, the agent inhibits hVR-1, hVR-2, and rVR-2 activity. In another embodiment, the agent stimulates hVR-1, hVR-2, and rVR-2 activity. In one embodiment, the agent is an antibody that specifically binds to an hVR-1, hVR-2, and rVR-2 protein. In another embodiment, the agent modulates expression of hVR-1, hVR-2, and rVR-2 by modulating transcription of an hVR-1, hVR-2, and rVR-2 gene or translation of an hVR-1, hVR-2, and rVR-2 mRNA. In yet another embodiment, the agent is a nucleic acid molecule having a nucleotide sequence that is antisense to the coding strand of an hVR-1, hVR-2, and rVR-2 mRNA or an hVR-1, hVR-2, and rVR-2 gene.

In one embodiment, the methods of the present invention are used to treat a subject having a disorder characterized by aberrant hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity by administering an agent which is an hVR-1, hVR-2, and rVR-2 modulator to the subject. In one embodiment, the hVR-1, hVR-2, and rVR-2 modulator is an hVR-1, hVR-2, and rVR-2 protein. In another embodiment the hVR-1, hVR-2, and rVR-2 modulator is an hVR-1, hVR-2, and rVR-2 nucleic acid molecule. In

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yet another embodiment, the hVR-1, hVR-2, and rVR-2 modulator is a peptide, peptidomimetic, or other small molecule. In a further embodiment, the disorder characterized by aberrant hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression is a pain disorder, e.g., hyperalgesia.

The present invention also provides a diagnostic assay for identifying the presence or absence of a genetic alteration characterized by at least one of (i) aberrant modification or mutation of a gene encoding an hVR-1, hVR-2, and rVR-2 protein; (ii) mis-regulation of the gene; and (iii) aberrant post-translational modification of an hVR-1, hVR-2, and rVR-2 protein, wherein a wild-type form of the gene encodes a protein with an hVR-1, hVR-2, and rVR-2 activity (as described herein).

In another aspect the invention provides a method for identifying a compound that binds to or modulates the activity of an hVR-1, hVR-2, and rVR-2 protein, by providing an indicator composition comprising an hVR-1, hVR-2, and rVR-2 protein having hVR-1, hVR-2, and rVR-2 activity, contacting the indicator composition with a test compound, and determining the effect of the test compound on hVR-1, hVR-2, and rVR-2 activity in the indicator composition to identify a compound that modulates the activity of an hVR-1, hVR-2, and rVR-2 protein.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

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Brief Description of the Drawings

Figure 1 depicts the full length cDNA sequence and predicted amino acid sequence of human VR-1 (hVR-1). The nucleotide sequence corresponds to nucleic acids 1 to 3909 of SEQ ID NO:1. The amino acid sequence corresponds to amino acids 1 to 839 of SEQ ID NO:2. The coding region without the 5' and 3' untranslated regions of the human VR-1 (hVR-1) gene is shown in SEQ ID NO:3.

Figure 2 depicts the full length cDNA sequence and predicted amino acid sequence of human VR-2 (hVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 2809 of SEQ ID NO:4. The amino acid sequence corresponds to amino acids 1 to 764 of SEQ ID NO:5. The coding region without the 5' and 3' untranslated regions of the human VR-2 (hVR-2) gene is shown in SEQ ID NO:6.

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Figure 3 depicts the partial cDNA sequence and partial predicted amino acid sequence of an alternate form of human VR-2 (hVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 1489 of SEQ ID NO:7. The amino acid sequence corresponds to amino acids 1 to 436 of SEQ ID NO:8. The coding region without the 5' and 3' untranslated regions of the alternate form of human VR-2 (hVR-2) gene is shown in SEQ ID NO:9.

Figure 4 depicts the partial cDNA sequence and partial predicted amino acid sequence of rat VR-2 (rVR-2). The nucleotide sequence corresponds to nucleic acids 1 to 1794 of SEQ ID NO:10. The amino acid sequence corresponds to amino acids 1 to 554 of SEQ ID NO:11. The coding region without the 5' and 3' untranslated regions of the rat VR-2 (rVR-2) gene is shown in SEQ ID NO:12.

Figure 5 depicts an alignment of the hVR-1 protein (SEQ ID NO:2) with the human VR-2 protein (SEQ ID NO:5) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

Figure 6 depicts an alignment of the hVR-1 nucleotide sequence (SEQ ID NO:1) with the human VR-2 nucleotide sequence (SEQ ID NO:4) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3.

Figure 7 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the rat VR-2 protein (SEQ ID NO:11) using the CLUSTAL W (1.74) multiple sequence alignment program.

Figure 8 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the rat VR-2 protein (SEQ ID NO:11) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

Figure 9 depicts an alignment of the hVR-1 nucleotide sequence (SEQ ID NO:1) with the rat VR-1 nucleotide sequence (Accession Number:AF029310) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3.

Figure 10 depicts an alignment of the hVR-1 protein (SEQ ID NO:2) with the rat

VR-1 protein (Accession Number:AF029310) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4.

Figure 11 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the human VR-2 protein (alternate form) (SEQ ID NO:8) using the CLUSTAL W (1.74) multiple sequence alignment program.

Figure 12 depicts a structural, hydrophobicity, and antigenicity analysis of the hVR-1 protein.

Figure 13 depicts the results of a search using the amino acid sequence of the hVR-1 protein against the HMM database.

Figure 14 depicts a structural, hydrophobicity, and antigenicity analysis of the hVR-2 protein.

10 Figure 15 depicts the results of a search using the amino acid sequence of the hVR-2 protein against the HMM database.

Figure 16 depicts the predicted full length amino acid sequence of the human VR-2 protein (alternate form) (SEQ ID NO:20).

Figure 17 depicts an alignment of the hVR-2 protein (SEQ ID NO:5) with the predicted full length human VR-2 protein (alternate form) (SEQ ID NO:20) using the CLUSTAL W (1.74) multiple sequence alignment program.

Detailed Description of the Invention

The present invention is based, at least in part, on the discovery of nucleic acid and amino acid molecules which are novel members of the Capsaicin/Vanilloid family of receptors. Described herein is the isolation of the human orthologue of rat VR-1 (rVR-1), referred to herein as hVR-1, as well as another previously unknown member of the VR family of receptors, referred herein as VR-2, and specifically as human VR-2 (hVR-2) and rat VR-2 (rVR-2) nucleic acid and protein molecules. The hVR-1, hVR-2, and rVR-2 molecules were identified based on their sequence similarity to the known rat vanilloid receptor (VR-1). VR-1 is a vanilloid gated, non-selective cation channel which resembles members of the transient receptor potential (TRP) ion channel family (described in Montell *et al.* (1989) *Neuron* 2:1313-1323) that mediate the influx of extracellular calcium in response to depletion of intracellular calcium stores. The rat VR-1 cDNA contains an open reading frame of 2514 nucleotides that encodes a protein of 838 amino acids. Hydrophilicity analysis has indicated that rat VR-1 contains six

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- 11 -

transmembrane domains (predicted to be mostly α-helices) with an additional short hydrophobic stretch between transmembrane regions 5 and 6. The amino terminal hydrophilic segment contains a relatively proline rich region followed by three ankyrin repeat domains. The rat VR-1 is expressed in small diameter neurons within sensory ganglia. The present hVR-1 sequence is the human orthologue of rVR-1. As described in further detail *infra*, the human VR-1 is expressed in nodose, trigeminal sensory neurons, as well as in some, but not all, small dorsal root ganglion (DRG) neurons and in a few medium sized DRG neurons.

The hVR-1, hVR-2, and rVR-2 molecules of the present invention play a role in pain signaling mechanisms. As used herein, the term "pain signaling mechanisms" includes the cellular mechanisms involved in the development and regulation of pain, e.g., pain elicited by noxious chemical, mechanical, or thermal stimuli, in a subject, e.g., a mammal such as a human. In mammals, the initial detection of noxious chemical, mechanical, or thermal stimuli, a process referred to as "nociception", occurs predominantly at the peripheral terminals of specialized, small diameter primary afferent neurons, called polymodal nociceptors. These afferent neurons transmit the information to the central nervous system, evoking a perception of pain or discomfort and initiating appropriate protective reflexes. Capsaicin/Vanilloid receptors, e.g., the hVR-1, hVR-2, and rVR-2 molecules of the present invention, present on these afferent neurons, are involved in detecting these noxious chemical, mechanical, or thermal stimuli and transducing this information into membrane depolarization events. Thus, the hVR-1, hVR-2, and rVR-2 molecules by participating in pain signaling mechanisms, can modulate pain elicitation and provide novel diagnostic targets and therapeutic agents to control pain.

The hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control pain in a variety of disorders, diseases, or conditions which are characterized by a deregulated, e.g., upregulated or downregulated, pain response. For example, the hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control the exaggerated pain response elicited during various forms of tissue injury, e.g., inflammation, infection, and ischemia, usually referred to as hyperalgesia (described in, for example, Fields, H.L. (1987) Pain, New York:McGraw-

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Hill). Moreover, the hVR-1, hVR-2, and rVR-2 molecules provide novel diagnostic targets and therapeutic agents to control pain associated with muscoloskeletal disorders, e.g., joint pain; tooth pain; headaches; pain associated with surgery, or neuropathic pain.

As the hVR-1 gene maps to a region of human chromosome 17 between WI-5436 (7.7cR) and WI-6584 (18.9cR) (Example 6), which has been associated with myasthenia gravis, Smith-Magenis syndrome, CORD5, Cone-rod dysrtophy, and breast cancer, the hVR-1 molecule may provide novel diagnostic targets and therapeutic agents to treat, diagnose, or prognose these disorders or other disorders linked to this chromosomal region. Similarly, as the hVR-2 gene maps to a region of human chromosome 17 between AFMA043ZB5 (23.3 cR) and D17S721 (29.3cR) (Example 6) which has been associated with myasthenia gravis, Smith-Magenis syndrome, CORD5, Cone-rod dysrtophy, choroidal dystrophy, central areolar, and retinal cone dystrophy, the hVR-2 molecule may provide novel diagnostic targets and therapeutic agents to treat, diagnose, or prognose these disorders or other disorders linked to this chromosomal region.

The term "family" when referring to the protein and nucleic acid molecules of the invention is intended to mean two or more proteins or nucleic acid molecules having a common structural domain or motif and having sufficient amino acid or nucleotide sequence homology as defined herein. Such family members can be naturally or non-naturally occurring and can be from either the same or different species. For example, a family can contain a first protein of human origin, as well as other, distinct proteins of human origin or alternatively, can contain homologues of non-human origin. Members of a family may also have common functional characteristics.

For example, the family of hVR-1, hVR-2, and rVR-2 proteins comprise at least one, and preferably six "transmembrane domains." As used herein, the term "transmembrane domain" includes an amino acid sequence of about 15 amino acid residues in length which spans the plasma membrane. More preferably, a transmembrane domain includes about at least 20, 25, 30, 35, 40, or 45 amino acid residues and spans the plasma membrane. Transmembrane domains are rich in hydrophobic residues, and typically have a helical structure. In a embodiment, at least 50%, 60%, 70%, 80%, 90%, 95% or more of the amino acid residues of a

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transmembrane domain are hydrophobic, *e.g.*, leucines, isoleucines, tyrosines, or tryptophans. Transmembrane domains are described in, for example, Zagotta W.N. et al, (1996) *Annual Rev. Neurosci.* 19: 235-63, the contents of which are incorporated herein by reference. Amino acid residues 434-455, 480-495, (509-531; based on homology to the rat VR-1) or 514-531, (543-569; based on homology to the rat VR-1) or 538-555, (577-596; based on homology to the rat VR-1) or 580-599, and (656-683; based on homology to the rat VR-1) or 658-682 of hVR-1 (SEQ ID NO:2) and amino acid residues 391-410, 431-448, 459-476, 486-508, 538-556, and 621-645 of hVR-2 (SEQ ID NO:5) comprise transmembrane domains.

In another embodiment, an hVR-1, hVR-2, and rVR-2 of the present invention is identified based on the presence of a "proline rich domain" in the protein or corresponding nucleic acid molecule. As used herein, the term "proline rich domain" includes an amino acid sequence of about 4-6 amino acid residues in length having the general sequence X-Pro-X-X-Pro-X (where X can be any amino acid). Proline rich domains are usually located in a helical structure and bind through hydrophobic interactions to SH3 domains. SH3 domains recognize proline rich domains in both forward and reverse orientations. Proline rich domains are described in, for example, Sattler M. et al. (1998) Leukemia 12:637-644, the contents of which are incorporated herein by reference.

In another embodiment, an hVR-1, hVR-2, and rVR-2 of the present invention is identified based on the presence of an "ankyrin repeat domain" in the protein or corresponding nucleic acid molecule. As used herein, the term "ankyrin repeat domain" includes a protein domain having an amino acid sequence of about 30-50 amino acid residues and having a bit score for the alignment of the sequence to the ankyrin repeat domain (HMM) of at least 6. Preferably, an ankyrin repeat domain includes at least about 30-45, more preferably about 30-40 amino acid residues, or about 30-35 amino acids and has a bit score for the alignment of the sequence to the ankyrin repeat domain (HMM) of at least 3-10, more preferably 10-30, more preferably 30-50, even more preferably 50-75, 75-100, 100-200 or greater. The ankyrin repeat domain HMM has been assigned the PFAM Accession PF00023 (http://genome.wustl.edu/Pfam/.html). Ankyrin repeats are involved in protein-protein interactions and are described in, for

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example, Ketchum K.A et al. (1996) FEBS Letters 378:19-26, the contents of which are incorporated herein by reference.

To identify the presence of an ankyrin repeat domain in an hVR-1, hVR-2, and rVR-2 protein and make the determination that a protein of interest has a particular profile, the amino acid sequence of the protein is searched against a database of HMMs 5 (e.g., the Pfam database, release 2.1) using the default parameters (http://www.sanger.ac.uk/Software/Pfam/HMM_search). A description of the Pfam database can be found in Sonhammer et al. (1997) Proteins 28(3)405-420 and a detailed description of HMMs can be found, for example, in Gribskov et al.(1990) Meth. Enzymol. 183:146-159; Gribskov et al. (1987) Proc. Natl. Acad. Sci. USA 84:4355-10 4358; Krogh et al. (1994) J. Mol. Biol. 235:1501-1531; and Stultz et al. (1993) Protein Sci. 2:305-314, the contents of which are incorporated herein by reference. A search was performed against the HMM database resulting in the identification of three ankyrin repeat domains in the amino acid sequence of SEQ ID NO:2 (at about residues 201-233, 248-283, and 333-361) and SEQ ID NO:5 (at about residues 162-194, 208-243, and 293-15 328). The results of the searches are set forth in Figures 13 and 15.

Isolated proteins of the present invention, preferably hVR-1, hVR-2, and rVR-2 proteins, have an amino acid sequence sufficiently identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11 or are encoded by a nucleotide sequence sufficiently identical to SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. As used herein, the term "sufficiently identical" refers to a first amino acid or nucleotide sequence which contains a sufficient or minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain) amino acid residues or nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences share common structural domains or motifs and/or a common functional activity. For example, amino acid or nucleotide sequences which share common structural domains have at least 30%, 40%, or 50% identity, preferably 60% identity, more preferably 70%-80%, and even more preferably 90-95% identity across the amino acid sequences of the domains and contain at least one and preferably two structural domains or motifs, are defined herein as sufficiently identical. Furthermore, amino acid or nucleotide sequences which share at least 30%, 40%,

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WO 00/29577 PCT/US99/26701 - 15 -

or 50%, preferably 60%, more preferably 70-80%, or 90-95% identity and share a common functional activity are defined herein as sufficiently identical.

As used interchangeably herein, an "hVR-1, hVR-2, and rVR-2 activity", "biological activity of hVR-1, hVR-2, and rVR-2" or "functional activity of hVR-1, hVR-2, and rVR-2", refers to an activity exerted by an hVR-1, hVR-2, and rVR-2 protein, polypeptide or nucleic acid molecule on an hVR-1, hVR-2, and rVR-2 responsive cell or on an hVR-1, hVR-2, and rVR-2 protein substrate, as determined in vivo, or in vitro, according to standard techniques. In one embodiment, an hVR-1, hVR-2, and rVR-2 activity is a direct activity, such as an association with an hVR-1, hVR-2, and rVR-2-target molecule. As used herein, a "target molecule" or "binding partner" is a molecule with which an hVR-1, hVR-2, and rVR-2 protein binds or interacts in nature, such that hVR-1, hVR-2, and rVR-2-mediated function is achieved. An hVR-1, hVR-2, and rVR-2 target molecule can be a non-hVR-1, non-hVR-2, and non-rVR-2 molecule or an hVR-1, hVR-2, and rVR-2 protein or polypeptide of the present invention. In an exemplary embodiment, an hVR-1, hVR-2, and rVR-2 target molecule is an hVR-1, hVR-2, and rVR-2 ligand, e.g., capsaicin. Alternatively, an hVR-1, hVR-2, and rVR-2 activity is an indirect activity, such as a cellular signaling activity mediated by interaction of the hVR-1, hVR-2, and rVR-2 protein with an hVR-1, hVR-2, and rVR-2 ligand.

Accordingly, another embodiment of the invention features isolated hVR-1, 20 hVR-2, and rVR-2 proteins and polypeptides having an hVR-1, hVR-2, and rVR-2 activity. Other proteins of the invention are hVR-1, hVR-2, and rVR-2 proteins having at least one, and preferably six, transmembrane domains and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Yet other proteins of the invention are hVR-1, hVR-2, and rVR-2 proteins having at least one transmembrane domain, at least one proline rich 25 domain and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Other proteins of the invention are hVR-1, hVR-2, and rVR-2 proteins having at least one transmembrane domain, at least one proline rich domain, at least one ankyrin repeat domain and, preferably, an hVR-1, hVR-2, and rVR-2 activity. Additional proteins of the invention have at least one transmembrane domain, at least one proline rich domain, at least one ankyrin repeat domain, and are, preferably, encoded by a nucleic acid molecule having a 30 nucleotide sequence which hybridizes under stringent hybridization conditions to a

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- 16 -

nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

The nucleotide sequence of the full length hVR-1 cDNA and the predicted amino acid sequence of the hVR-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively.

The nucleotide sequence of the full length hVR-2 cDNA and the predicted amino acid sequence of the hVR-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively.

The nucleotide sequence of the partial hVR-2 (alternate form) cDNA and the predicted amino acid sequence of the hVR-2 (alternate form) polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively.

The nucleotide sequence of the partial rVR-2 cDNA and the predicted amino acid sequence of the rVR-2 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively.

Various aspects of the invention are described in further detail in the following subsections:

I. Isolated Nucleic Acid Molecules

One aspect of the invention pertains to isolated nucleic acid molecules that encode hVR-1, hVR-2, and rVR-2 proteins or biologically active portions thereof, as well as nucleic acid fragments sufficient for use as hybridization probes to identify hVR-1, hVR-2, and rVR-2-encoding nucleic acid molecules (*e.g.*, hVR-1, hVR-2, and rVR-2 mRNA) and fragments for use as PCR primers for the amplification or mutation of hVR-1, hVR-2, and rVR-2 nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (*e.g.*, cDNA or genomic DNA) and RNA molecules (*e.g.*, mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

The term "isolated nucleic acid molecule" includes nucleic acid molecules which are separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. For example, with regards to genomic DNA, the term "isolated"

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- 17 -

includes nucleic acid molecules which are separated from the chromosome with which the genomic DNA is naturally associated. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated hVR-1, hVR-2, and rVR-2 nucleic acid molecule can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. Using all or portion of the nucleic acid sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, as a hybridization probe, hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be isolated using standard hybridization and cloning techniques (e.g., as described in Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

Moreover, a nucleic acid molecule encompassing all or a portion of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, can be isolated by the polymerase chain reaction (PCR) using synthetic oligonucleotide primers designed based upon the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to hVR-1, hVR-2, and rVR-2 nucleotide sequences can be prepared by standard synthetic techniques, *e.g.*, using an automated DNA synthesizer.

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In one embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:1. The sequence of SEQ ID NO:1 corresponds to the full length hVR-1 encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:4. The sequence of SEQ ID NO:4 corresponds to the full length hVR-2 encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:7. The sequence of SEQ ID NO:7 corresponds to a fragment of the hVR-2 (alternate form) encoding cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises the nucleotide sequence shown in SEQ ID NO:10. The sequence of SEQ ID NO:10 corresponds to a fragment of the rVR-2 cDNA.

In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule which is a complement of the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion of any of these nucleotide sequences. A nucleic acid molecule which is complementary to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, is one which is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, such that it can hybridize to the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 thereby forming a stable duplex.

In still another embodiment, an isolated nucleic acid molecule of the present invention comprises a nucleotide sequence which is at least about 60%, 65%, 70%, 75%, 80%, 83%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous to the entire length of the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion of any of these nucleotide sequences.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, for example, a fragment which can be used as a probe or primer or a fragment encoding a portion of an hVR-1, hVR-2, and rVR-2 protein, e.g., a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein. The nucleotide sequence determined from the cloning of the hVR-1, hVR-2, and rVR-2 gene allows for the generation of probes and primers

designed for use in identifying and/or cloning other hVR-1, hVR-2, and rVR-2 family members, as well as hVR-1, hVR-2, and rVR-2 homologues from other species. The probe/primer typically comprises a substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12 or 15, preferably about 20 or 25, more preferably about 30, 35, 40, 45, 50, 55, 60, 65, 75, or 100 consecutive nucleotides of a sense sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, of an anti-sense sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or of a naturally occurring allelic variant or mutant of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In an exemplary embodiment, a nucleic acid molecule of the present invention comprises a nucleotide sequence which is greater than 100-150, 150-200, 200-250, 250-300, 300-350, 350-400, 400-450, 450-500, 500-550, 550-600, 600-650, 650-700, 700-750, 750-800, 800-850, 850-900, 900-950, 950-1000,

Probes based on the hVR-1, hVR-2, and rVR-2 nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In preferred embodiments, the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress an hVR-1, hVR-2, and rVR-2 protein, such as by measuring a level of an hVR-1, hVR-2, and rVR-2-encoding nucleic acid in a sample of cells from a subject e.g., detecting hVR-1, hVR-2, and rVR-2 mRNA levels or determining whether a genomic hVR-1, hVR-2, and rVR-2 gene has been mutated or deleted.

1088, or more nucleotides in length and hybridizes under stringent hybridization

conditions to a nucleic acid molecule of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12.

A nucleic acid fragment encoding a "biologically active portion of an hVR-1, hVR-2, and rVR-2 protein" can be prepared by isolating a portion of the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, which encodes a polypeptide having an hVR-1, hVR-2, and rVR-2 biological activity (the biological activities of the hVR-1, hVR-2, and rVR-2 proteins are described herein), expressing the encoded portion of the hVR-1, hVR-2, and rVR-2 protein (e.g., by recombinant expression *in vitro*) and assessing the activity of the encoded portion of the hVR-1, hVR-2, and rVR-2 protein.

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The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, due to degeneracy of the genetic code and thus encode the same hVR-1, hVR-2, and rVR-2 proteins as those encoded by the nucleotide sequence shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11.

In addition to the hVR-1, hVR-2, and rVR-2 nucleotide sequences shown in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the hVR-1, hVR-2, and rVR-2 proteins may exist within a population (*e.g.*, the human population). Such genetic polymorphism in the hVR-1, hVR-2, and rVR-2 genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules which include an open reading frame encoding an hVR-1, hVR-2, and rVR-2 protein, preferably a mammalian hVR-1, hVR-2, and rVR-2 protein, and can further include noncoding regulatory sequences, and introns.

Allelic variants of hVR-1, hVR-2, and rVR-2 include both functional and non-functional hVR-1, hVR-2, and rVR-2 proteins. Functional allelic variants are naturally occurring amino acid sequence variants of the hVR-1, hVR-2, and rVR-2 protein that maintain the ability to bind an hVR-1, hVR-2, and rVR-2 ligand and/or modulate a pain signaling mechanism. Functional allelic variants will typically contain only conservative substitution of one or more amino acids of SEQ ID NO:2, 5, 8, or 11, or substitution, deletion or insertion of non-critical residues in non-critical regions of the protein.

Non-functional allelic variants are naturally occurring amino acid sequence variants of the hVR-1, hVR-2, and rVR-2 protein that do not have the ability to either bind an hVR-1, hVR-2, and rVR-2 ligand and/or modulate a pain signaling mechanism. Non-functional allelic variants will typically contain a non-conservative substitution, a deletion, or insertion or premature truncation of the amino acid sequence of SEQ ID

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- 21 -

NO:2, 5, 8, or 11, or a substitution, insertion or deletion in critical residues or critical regions.

The present invention further provides non-human orthologues of the hVR-2 and rVR-2 protein. Orthologues of the hVR-2 and rVR-2 protein are proteins that are isolated from non-human and non-rat organisms and possess the same hVR-2 and rVR-2 ligand binding and/or modulation of pain signaling mechanism capabilities of the hVR-2 and rVR-2 proteins. Orthologues of the hVR-2 and rVR-2 proteins can readily be identified as comprising an amino acid sequence that is substantially homologous to SEQ ID NO: 4, 6, 8 or 10.

Moreover, nucleic acid molecules encoding other hVR-1, hVR-2, and rVR-2 family members and, thus, which have a nucleotide sequence which differs from the hVR-1, hVR-2, and rVR-2 sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, are intended to be within the scope of the invention. For example, another hVR-1, hVR-2, and rVR-2 cDNA can be identified based on the nucleotide sequence of hVR-1, hVR-2, and rVR-2. Moreover, nucleic acid molecules encoding VR-2 proteins from different species, and which, thus, have a nucleotide sequence which differs from the hVR-2 and rVR-2 sequences of SEQ ID NO:4, 6, 8, or 10 are intended to be within the scope of the invention. For example, a mouse hVR-2 cDNA can be identified based on the nucleotide sequence of the human VR-2 (hVR-2) or the rat VR-2 (rVR-2).

Nucleic acid molecules corresponding to natural allelic variants and homologues of the hVR-1, hVR-2, and rVR-2 cDNAs of the invention can be isolated based on their homology to the hVR-1, hVR-2, and rVR-2 nucleic acids disclosed herein using the cDNAs disclosed herein, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. Nucleic acid molecules corresponding to natural allelic variants and homologues of the hVR-1, hVR-2, and rVR-2 cDNAs of the invention can further be isolated by mapping to the same chromosome or locus as the hVR-1, hVR-2, and rVR-2 gene.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 15, 20, 25, 30 or more nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. In other embodiment, the nucleic acid is at least

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- 22 -

30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, or 950 nucleotides in length. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% identical to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 70%, more preferably at least about 80%, even more preferably at least about 85% or 90% identical to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, nonlimiting example of stringent hybridization conditions are hybridization in 6X sodium 10 chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50°C, preferably at 55°C, more preferably at 60°C, and even more prefcrably at 65°C. Preferably, an isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 corresponds to a naturally-occurring nucleic acid molecule. As used herein, a 15 "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein).

In addition to naturally-occurring allelic variants of the hVR-1, hVR-2, and rVR-2 sequences that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, thereby leading to changes in the amino acid sequence of the encoded hVR-1, hVR-2, and rVR-2 proteins, without altering the functional ability of the hVR-1, hVR-2, and rVR-2 proteins. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of hVR-1, hVR-2, and rVR-2 (e.g., the sequence of SEQ ID NO:2, 5, 8, or 11) without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the hVR-1, hVR-2, and rVR-2 proteins of the present invention, are predicted to be particularly unamenable to alteration. Furthermore,

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additional amino acid residues that are conserved between the hVR-1, hVR-2, and rVR-2 proteins of the present invention and other members of the Capsaicin/Vanilloid receptor family are not likely to be amenable to alteration.

Accordingly, another aspect of the invention pertains to nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins that contain changes in amino acid residues that are not essential for activity. Such hVR-1, hVR-2, and rVR-2 proteins differ in amino acid sequence from SEQ ID NO:2, 5, 8, or 11, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to SEQ ID NO:2, 5, 8, or 11.

An isolated nucleic acid molecule encoding an hVR-1, hVR-2, and rVR-2 protein homologous to the protein of SEQ ID NO:2, 5, 8, or 11 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, such that one or more amino acid substitutions. additions or deletions are introduced into the encoded protein. Mutations can be introduced into SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in an hVR-1, hVR-2, and rVR-2 protein is preferably replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of an hVR-1, hVR-

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WO 00/29577 PCT/US99/26701
- 24 -

2, and rVR-2 coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for hVR-1, hVR-2, and rVR-2 biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or

In a embodiment, a mutant hVR-1, hVR-2, and rVR-2 protein can be assayed for the ability to (1) interact with a non-hVR-1, non-hVR-2, or non- rVR-2 protein molecule, e.g., a vanilloid compound such as capsaicin; (2) modulate intracellular calcium concentration; (3) activate an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; or (4) modulate a pain signaling mechanism.

In addition to the nucleic acid molecules encoding hVR-1, hVR-2, and rVR-2 proteins described above, another aspect of the invention pertains to isolated nucleic acid molecules which are antisense thereto. An "antisense" nucleic acid comprises a nucleotide sequence which is complementary to a "sense" nucleic acid encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire hVR-1, hVR-2, and rVR-2 coding strand, or to only a portion thereof. In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding hVR-1, hVR-2, and rVR-2. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (e.g., the coding region of hVR-1, hVR-2, and rVR-2). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding hVR-1, hVR-2, and rVR-2. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding hVR-1, hVR-2, and rVR-2 disclosed herein, antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of hVR-1, hVR-2, and rVR-2 mRNA, but more preferably is an oligonucleotide which is antisense to only a portion of the coding

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or noncoding region of hVR-1, hVR-2, and rVR-2 mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of hVR-1, hVR-2, and rVR-2 mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation 5 reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine 10 substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, 15 N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, 20 queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5- oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (i.e., RNA 25 transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an hVR-1, hVR-2, and rVR-2 protein to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The

WO 00/29577 PCT/US99/26701 - 26 -

hybridization can be by conventional nucleotide complementarity to form a stable duplex. or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention include direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, *e.g.*, by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an -anomeric nucleic acid molecule. An α-anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids*. *Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue *et al.* (1987) *FEBS Lett.* 215:327-330).

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) Nature 334:585-591)) can be used to catalytically cleave hVR-1, hVR-2, and rVR-2 mRNA transcripts to thereby inhibit translation of hVR-1, hVR-2, and rVR-2 mRNA. A ribozyme having specificity for an hVR-1, hVR-2, and rVR-2-encoding nucleic acid can be designed based upon the nucleotide sequence of an hVR-1, hVR-2, and rVR-2 cDNA disclosed herein (i.e., SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12). For example, a derivative of a Tetrahymena L-19 IVS

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WO 00/29577 PCT/US99/26701 - 27 -

RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an hVR-1, hVR-2, and rVR-2-encoding mRNA. See, e.g., Cech et al. U.S. Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, hVR-1, hVR-2, and rVR-2 mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, for example, Bartel, D. and Szostak, J.W. (1993) *Science* 261:1411-1418.

Alternatively, hVR-1, hVR-2, and rVR-2 gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the hVR-1, hVR-2, and rVR-2 (e.g., the hVR-1, hVR-2, and rVR-2 promoter and/or enhancers) to form triple helical structures that prevent transcription of the hVR-1, hVR-2, and rVR-2 gene in target cells. See generally, Helene, C. (1991) *Anticancer Drug Des.* 6(6):569-84; Helene, C. et al. (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher, L.J. (1992) *Bioassays* 14(12):807-15.

In yet another embodiment, the hVR-1, hVR-2, and rVR-2 nucleic acid molecules of the present invention can be modified at the base moiety, sugar moiety or phosphate backbone to improve, e.g., the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acid molecules can be modified to generate peptide nucleic acids (see Hyrup B. et al. (1996) Bioorganic & Medicinal Chemistry 4 (1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, e.g., DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup B. et al. (1996) supra; Perry-O'Keefe et al. Proc. Natl. Acad. Sci. 93: 14670-675.

PNAs of hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, for example, inducing transcription or translation arrest or inhibiting replication. PNAs of hVR-1,

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hVR-2, and rVR-2 nucleic acid molecules can also be used in the analysis of single base pair mutations in a gene, (e.g., by PNA-directed PCR clamping); as 'artificial restriction enzymes' when used in combination with other enzymes, (e.g., S1 nucleases (Hyrup B. (1996) supra)); or as probes or primers for DNA sequencing or hybridization (Hyrup B. et al. (1996) supra; Perry-O'Keefe supra).

In another embodiment, PNAs of hVR-1, hVR-2, and rVR-2 can be modified, (e.g., to enhance their stability or cellular uptake), by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of hVR-1, hVR-2, and rVR-2 nucleic acid molecules can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, (e.g., RNAse H and DNA polymerases), to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup B. (1996) supra). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup B. (1996) supra and Finn P.J. et al. (1996) Nucleic Acids Res. 24 (17): 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs, e.g., 5'-(4methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used as a between the PNA and the 5' end of DNA (Mag, M. et al. (1989) Nucleic Acid Res. 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn P.J. et al. (1996) supra). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser, K.H. et al. (1975) Bioorganic Med. Chem. Lett. 5: 1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors in vivo), or agents facilitating transport across the cell membrane (see, e.g., Letsinger et al. (1989) Proc. Natl. Acad. Sci. USA 86:6553-6556; Lemaitre et al. (1987) Proc. Natl. Acad. Sci. USA 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, e.g., PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization-

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triggered cleavage agents (See, e.g., Krol et al. (1988) Bio-Techniques 6:958-976) or intercalating agents. (See, e.g., Zon (1988) Pharm. Res. 5:539-549). To this end, the oligonucleotide may be conjugated to another molecule, (e.g., a peptide, hybridization triggered cross-linking agent, transport agent, or hybridization-triggered cleavage agent).

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II. Isolated hVR-1, hVR-2, and rVR-2 Proteins and Anti-hVR-1, Anti-hVR-2, and Anti-rVR-2 Antibodies

One aspect of the invention pertains to isolated hVR-1, hVR-2, and rVR-2 proteins, and biologically active portions thereof, as well as polypeptide fragments suitable for use as immunogens to raise anti-hVR-2, anti-hVR-2, and anti-rVR-2 antibodies. In one embodiment, native hVR-1, hVR-2, and rVR-2 proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, hVR-1, hVR-2, and rVR-2 proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, an hVR-1, hVR-2, and rVR-2 protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the hVR-1, hVR-2, and rVR-2 protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of hVR-1, hVR-2, and rVR-2 protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of hVR-1, hVR-2, and rVR-2 protein having less than about 30% (by dry weight) of non-hVR-1, hVR-2, and rVR-2 protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-hVR-1, hVR-2, and rVR-2 protein, still more preferably less than about 5% non-hVR-1, hVR-2, and rVR-2 protein, and most preferably less than about 5% non-hVR-1, non-hVR-2, and non-rVR-2 protein. When the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture

WO 00/29577 PCT/US99/26701 - 30 -

medium, *i.e.*, culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of hVR-1, hVR-2, and rVR-2 protein in which the protein is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of hVR-1, hVR-2, and rVR-2 protein having less than about 30% (by dry weight) of chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, more preferably less than about 20% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, still more preferably less than about 10% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals, and most preferably less than about 5% chemical precursors or non-hVR-1, hVR-2, and rVR-2 chemicals.

15 As used herein, a "biologically active portion" of an hVR-1, hVR-2, and rVR-2 protein includes a fragment of an hVR-1, hVR-2, and rVR-2 protein which participates in an interaction between an hVR-1, hVR-2, and rVR-2 molecule and a non-hVR-1, non-hVR-2, and non-rVR-2 molecule, respectively. Biologically active portions of an hVR-1, hVR-2, and rVR-2 protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the hVR-1, hVR-20 2, and rVR-2 protein, e.g., the amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11, which include less amino acids than the full length hVR-1, hVR-2, and rVR-2 proteins, and exhibit at least one activity of an hVR-1, hVR-2, and rVR-2 protein. Typically, biologically active portions comprise a domain or motif with at least one activity of the hVR-1, hVR-2, and rVR-2 protein, e.g., binding of an hVR-1, hVR-2, and rVR-2 ligand 25 such as a vanilloid compound, e.g., Capsaicin. A biologically active portion of an hVR-1, hVR-2, and rVR-2 protein can be a polypeptide which is, for example, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200 or more amino acids in length. Biologically active portions of an hVR-1, hVR-2, and rVR-2 protein can be used as targets for developing agents which modulate an hVR-1, hVR-2, and rVR-2 mediated activity, e.g., a pain signaling 30 mechanism.

- 31 -

In one embodiment, a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein comprises at least one transmembrane domain, and/or at least one proline rich domain, and/or at least one ankyrin repeat domain. It is to be understood that a biologically active portion of an hVR-1, hVR-2, and rVR-2 protein of the present invention may contain at least one of the above-identified structural domains. A more biologically active portion of an hVR-1, hVR-2, and rVR-2 protein may contain at least two of the above-identified structural domains. Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native hVR-1, hVR-2, and rVR-2 protein.

In a embodiment, the hVR-1, hVR-2, and rVR-2 protein has an amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11. In other embodiments, the hVR-1, hVR-2, and rVR-2 protein is substantially homologous to SEQ ID NO:2, 5, 8, or 11, and retains the functional activity of the protein of SEQ ID NO:2, 5, 8, or 11, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail in subsection I above. Accordingly, in another embodiment, the hVR-1, hVR-2, and rVR-2 protein is a protein which comprises an amino acid sequence at least about 60%, 65%, 70%, 75%, 80%, 85%, 87%, 90%, 95%, 98% or more homologous to SEQ ID NO:2, 5, 8, or 11.

To determine the percent identity of two amino acid sequences or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, or 90% of the length of the reference sequence (e.g., when aligning a second sequence to the hVR-1, hVR-2, and rVR-2 amino acid sequence of SEQ ID NO:2, 5, 8, or 11, having 177 amino acid residues, at least 80, preferably at least 100, more preferably at least 120, even more preferably at least 140, and even more preferably at least 150, 160 or 170 amino acid residues are aligned). The amino acid residues or nucleotides at

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- 32 -

corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In a embodiment, the 10 percent identity between two amino acid sequences is determined using the Needleman and Wunsch (J. Mol. Biol. (48):444-453 (1970)) algorithm which has been incorporated into the GAP program in the GCG software package (available at http://www.gcg.com), using either a Blossum 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another embodiment, the 15 percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (available at http://www.gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. In another embodiment, the percent identity between two amino acid or nucleotide sequences is determined using the algorithm of E. Meyers and W. 20 Miller (CABIOS, 4:11-17 (1989)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The nucleic acid and protein sequences of the present invention can further be

25 used as a "query sequence" to perform a search against public databases to, for example, identify other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (1990) J.

Mol. Biol. 215:403-10. BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences

30 homologous to hVR-1, hVR-2, and rVR-2 nucleic acid molecules of the invention.

BLAST protein searches can be performed with the XBLAST program, score = 50,

- 33 -

wordlength = 3 to obtain amino acid sequences homologous to hVR-1, hVR-2, and rVR-2 protein molecules of the invention. To obtain gapped alignments for comparison purposes. Gapped BLAST can be utilized as described in Altschul *et al.*, (1997) *Nucleic Acids Res.* 25(17):3389-3402. When utilizing BLAST and Gapped BLAST programs, the default parameters of the respective programs (*e.g.*, XBLAST and NBLAST) can be used. See http://www.ncbi.nlm.nih.gov.

The invention also provides hVR-1, hVR-2, and rVR-2 chimeric or fusion proteins. As used herein, an hVR-1, hVR-2, and rVR-2 "chimeric protein" or "fusion protein" comprises an hVR-1, hVR-2, and rVR-2 polypeptide operatively linked to a non-hVR-1, hVR-2, and rVR-2 polypeptide. An "hVR-1, hVR-2, and rVR-2 10 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to hVR-1, hVR-2, and rVR-2, whereas a "non-hVR-1, non-hVR-2, and non-rVR-2 polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein which is not substantially homologous to the hVR-1, hVR-2, and rVR-2 protein, e.g., a protein which is different from the hVR-1, hVR-2, and rVR-2 protein and which 15 is derived from the same or a different organism. Within an hVR-1, hVR-2, and rVR-2 fusion protein the hVR-1, hVR-2, and rVR-2 polypeptide can correspond to all or a portion of an hVR-1, hVR-2, and rVR-2 protein. In a embodiment, an hVR-1, hVR-2, and rVR-2 fusion protein comprises at least one biologically active portion of an hVR-1, hVR-2, and rVR-2 protein. In another embodiment, an hVR-1, hVR-2, and rVR-2 fusion protein comprises at least two biologically active portions of an hVR-1, hVR-2, and rVR-2 protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the hVR-1, hVR-2, and rVR-2 polypeptide and the non-hVR-1, nonhVR-2, and non-rVR-2 polypeptide are fused in-frame to each other. The non-hVR-1, hVR-2, and rVR-2 polypeptide can be fused to the N-terminus or C-terminus of the 25 hVR-1, hVR-2, and rVR-2 polypeptide.

For example, in one embodiment, the fusion protein is a GST-hVR-1, GST-hVR-2, and GST-rVR-2 fusion protein in which the hVR-1, hVR-2, and rVR-2 sequences are fused to the C-terminus of the GST sequences. Such fusion proteins can facilitate the purification of recombinant hVR-1, hVR-2, and rVR-2.

- 34 -

In another embodiment, the fusion protein is an hVR-1, hVR-2, and rVR-2 protein containing a heterologous signal sequence at its N-terminus. In certain host cells (e.g., mammalian host cells), expression and/or secretion of hVR-1, hVR-2, and rVR-2 can be increased through use of a heterologous signal sequence.

The hVR-1, hVR-2, and rVR-2 fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject *in vivo*. The hVR-1, hVR-2, and rVR-2 fusion proteins can be used to affect the bioavailability of an hVR-1, hVR-2, and rVR-2 substrate. Use of hVR-1, hVR-2, and rVR-2 fusion proteins may be useful therapeutically for the treatment of disorders caused by, for example, (i) aberrant modification or mutation of a gene encoding an hVR-1, hVR-2, and rVR-2 protein; (ii) mis-regulation of the hVR-1, hVR-2, and rVR-2 gene; and (iii) aberrant post-translational modification of an hVR-1, hVR-2, and rVR-2 protein.

Moreover, the hVR-1, hVR-2, and rVR-2-fusion proteins of the invention can be used as immunogens to produce anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies in a subject, to purify hVR-1, hVR-2, and rVR-2 ligands and in screening assays to identify molecules which inhibit the interaction of hVR-1, hVR-2, and rVR-2 with an hVR-1, hVR-2, and rVR-2 substrate.

Preferably, an hVR-1, hVR-2, and rVR-2 chimeric or fusion protein of the invention is produced by standard recombinant DNA techniques. For example, DNA 20 fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, 25 the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Current Protocols in Molecular Biology, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression 30 vectors are commercially available that already encode a fusion moiety (e.g., a GST

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polypeptide). An hVR-1, hVR-2, and rVR-2-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the hVR-1, hVR-2, and rVR-2 protein.

The present invention also pertains to variants of the hVR-1, hVR-2, and rVR-2 proteins which function as either hVR-1, hVR-2, and rVR-2 agonists (mimetics) or as hVR-1, hVR-2, and rVR-2 antagonists. Variants of the hVR-1, hVR-2, and rVR-2 proteins can be generated by mutagenesis, *e.g.*, discrete point mutation or truncation of an hVR-1, hVR-2, and rVR-2 protein. An agonist of the hVR-1, hVR-2, and rVR-2 proteins can retain substantially the same, or a subset, of the biological activities of the naturally occurring form of an hVR-1, hVR-2, and rVR-2 protein. An antagonist of an hVR-1, hVR-2, and rVR-2 protein can inhibit one or more of the activities of the naturally occurring form of the hVR-1, hVR-2, and rVR-2 protein by, for example, competitively modulating an hVR-1, hVR-2, and rVR-2-mediated activity of an hVR-1, hVR-2, and rVR-2 protein. Thus, specific biological effects can be elicited by treatment with a variant of limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the protein has fewer side effects in a subject relative to treatment with the naturally occurring form of the hVR-1, hVR-2, and rVR-2 protein.

In one embodiment, variants of an hVR-1, hVR-2, and rVR-2 protein which 20 function as either hVR-1, hVR-2, and rVR-2 agonists (mimetics) or as hVR-1, hVR-2, and rVR-2 antagonists can be identified by screening combinatorial libraries of mutants, e.g., truncation mutants, of an hVR-1, hVR-2, and rVR-2 protein for hVR-1, hVR-2, and rVR-2 protein agonist or antagonist activity. In one embodiment, a variegated library of hVR-1, hVR-2, and rVR-2 variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of 25 hVR-1, hVR-2, and rVR-2 variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential hVR-1, hVR-2, and rVR-2 sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g., for phage display) containing the set of hVR-1, hVR-2, and rVR-2 sequences therein. There are a 30 variety of methods which can be used to produce libraries of potential hVR-1, hVR-2,

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- 36 -

and rVR-2 variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential hVR-1, hVR-2, and rVR-2 sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, e.g., Narang, S.A. (1983) *Tetrahedron* 39:3; Itakura et al. (1984) *Annu. Rev. Biochem.* 53:323; Itakura et al. (1984) *Science* 198:1056; Ike et al. (1983) *Nucleic Acid Res.* 11:477.

In addition, libraries of fragments of an hVR-1, hVR-2, and rVR-2 protein coding sequence can be used to generate a variegated population of hVR-1, hVR-2, and rVR-2 fragments for screening and subsequent selection of variants of an hVR-1, hVR-2, and rVR-2 protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of an hVR-1, hVR-2, and rVR-2 coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA which can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal, C-terminal and internal fragments of various sizes of the hVR-1, hVR-2, and rVR-2 protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of hVR-1, hVR-2, and rVR-2 proteins. The most widely used techniques, which are amenable to high through-put analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recrusive ensemble mutagenesis

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- 37 -

(REM), a new technique which enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify hVR-1, hVR-2, and rVR-2 variants (Arkin and Yourvan (1992) *Proc. Natl. Acad. Sci. USA* 89:7811-7815; Delgrave *et al.* (1993) *Protein Engineering* 6(3):327-331).

In one embodiment, cell based assays can be exploited to analyze a variegated hVR-1, hVR-2, and rVR-2 library. For example, a library of expression vectors can be transfected into a cell line, e.g., a neuronal cell line, which ordinarily responds to a particular ligand in an hVR-1, hVR-2, and rVR-2-dependent manner. The transfected cells are then contacted with the ligand and the effect of expression of the mutant on signaling by the ligand can be detected, e.g., by measuring intracellular calcium concentration, neuronal membrane depolarization, or the activity of an hVR-1, hVR-2, and rVR-2-regulated transcription factor. Plasmid DNA can then be recovered from the cells which score for inhibition, or alternatively, potentiation of signaling by the ligand, and the individual clones further characterized.

An isolated hVR-1, hVR-2, and rVR-2 protein, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind hVR-1, hVR-2, and rVR-2 using standard techniques for polyclonal and monoclonal antibody preparation. A full-length hVR-1, hVR-2, and rVR-2 protein can be used or, alternatively, the invention provides antigenic peptide fragments of hVR-1, hVR-2, and rVR-2 for use as

20 immunogens. The antigenic peptide of hVR-1, hVR-2, and rVR-2 comprises at least 8 amino acid residues of the amino acid sequence shown in SEQ ID NO:2, 5, 8, or 11 and encompasses an epitope of hVR-1, hVR-2, and rVR-2 such that an antibody raised against the peptide forms a specific immune complex with hVR-1, hVR-2, and rVR-2. Preferably, the antigenic peptide comprises at least 10 amino acid residues, more preferably at least 15 amino acid residues, even more preferably at least 20 amino acid residues, and most preferably at least 30 amino acid residues.

Epitopes encompassed by the antigenic peptide are regions of hVR-1, hVR-2, and rVR-2 that are located on the surface of the protein, e.g., hydrophilic regions, as well as regions with high antigenicity (see, for example, Figures 12 and 14).

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- 38 -

An hVR-1, hVR-2, and rVR-2 immunogen typically is used to prepare antibodies by immunizing a suitable subject, (e.g., rabbit, goat, mouse or other mammal) with the immunogen. An appropriate immunogenic preparation can contain, for example, recombinantly expressed hVR-1, hVR-2, and rVR-2 protein or a chemically synthesized hVR-1, hVR-2, and rVR-2 polypeptide. The preparation can further include an adjuvant, such as Freund's complete or incomplete adjuvant, or similar immunostimulatory agent. Immunization of a suitable subject with an immunogenic hVR-1, hVR-2, and rVR-2 preparation induces a polyclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody response.

Accordingly, another aspect of the invention pertains to anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site which specifically binds (immunoreacts with) an antigen, such as hVR-1, hVR-2, and rVR-2. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin. The invention provides polyclonal and monoclonal antibodies that bind hVR-1, hVR-2, and rVR-2. The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen binding site capable of immunoreacting with a particular epitope of hVR-1, hVR-2, and rVR-2. A monoclonal antibody composition thus typically displays a single binding affinity for a particular hVR-1, hVR-2, and rVR-2 protein with which it immunoreacts.

Polyclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies can be prepared as described above by immunizing a suitable subject with an hVR-1, hVR-2, and rVR-2 immunogen. The anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized hVR-1, hVR-2, and rVR-2. If desired, the antibody molecules directed against hVR-1, hVR-2, and rVR-2 can be isolated from the mammal (e.g., from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction. At an

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- 39 -

appropriate time after immunization, e.g., when the anti-hVR-1, anti-hVR-2, and antirVR-2 antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) Nature 256:495-497) (see also, Brown et al. (1981) J. Immunol. 127:539-46; Brown et al. (1980) J. Biol. Chem .255:4980-83; Yeh et al. (1976) Proc. Natl. Acad. Sci. USA 76:2927-31; and Yeh et al. (1982) Int. J. Cancer 29:269-75), the more recent human B cell hybridoma technique (Kozbor et al. (1983) Immunol Today 4:72), the EBVhybridoma technique (Cole et al. (1985), Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing 10 monoclonal antibody hybridomas is well known (see generally R. H. Kenneth, in Monoclonal Antibodies: A New Dimension In Biological Analyses, Plenum Publishing Corp., New York, New York (1980); E. A. Lerner (1981) Yale J. Biol. Med., 54:387402; M. L. Gefter et al. (1977) Somatic Cell Genet. 3:23136). Briefly, an immortal cell line (typically a myeloma) is fused to lymphocytes (typically splenocytes) from a mammal 15 immunized with an hVR-1, hVR-2, and rVR-2 immunogen as described above, and the culture supernatants of the resulting hybridoma cells are screened to identify a hybridoma producing a monoclonal antibody that binds hVR-1, hVR-2, and rVR-2.

Any of the many well known protocols used for fusing lymphocytes and immortalized cell lines can be applied for the purpose of generating an anti-hVR-1, anti-20 hVR-2, and anti-rVR-2 monoclonal antibodies (see, e.g., G. Galfre et al. (1977) Nature 266:55052; Gefter et al. Somatic Cell Genet., cited supra; Lerner, Yale J. Biol. Med., cited supra; Kenneth, Monoclonal Antibodies, cited supra). Moreover, the ordinarily skilled worker will appreciate that there are many variations of such methods which also would be useful. Typically, the immortal cell line (e.g., a myeloma cell line) is derived 25 from the same mammalian species as the lymphocytes. For example, murine hybridomas can be made by fusing lymphocytes from a mouse immunized with an immunogenic preparation of the present invention with an immortalized mouse cell line. immortal cell lines are mouse myeloma cell lines that are sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). Any of a 30 number of myeloma cell lines can be used as a fusion partner according to standard

- 40 -

techniques, *e.g.*, the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines. These myeloma lines are available from ATCC. Typically, HAT-sensitive mouse myeloma cells are fused to mouse splenocytes using polyethylene glycol ("PEG"). Hybridoma cells resulting from the fusion are then selected using HAT medium, which kills unfused and unproductively fused myeloma cells (unfused splenocytes die after several days because they are not transformed). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind hVR-1, hVR-2, and rVR-2, *e.g.*, using a standard ELISA assay.

10 Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with hVR-1, hVR-2, and rVR-2 to thereby isolate immunoglobulin library members that bind hVR-1, hVR-2, and rVR-2. Kits for generating and screening phage display libraries are commercially available (e.g., the 15 Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; and the Stratagene SurfZAPTM Phage Display Kit, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, Ladner et al. U.S. Patent No. 5,223,409; Kang et al. PCT International Publication No. WO 92/18619; Dower et al. 20 PCT International Publication No. WO 91/17271; Winter et al. PCT International Publication WO 92/20791; Markland et al. PCT International Publication No. WO 92/15679; Breitling et al. PCT International Publication WO 93/01288; McCafferty et al. PCT International Publication No. WO 92/01047; Garrard et al. PCT International Publication No. WO 92/09690; Ladner et al. PCT International Publication No. WO 25 90/02809; Fuchs et al. (1991) Bio/Technology 9:1370-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; Griffiths et al. (1993) EMBO J 12:725-734; Hawkins et al. (1992) J. Mol. Biol. 226:889-896; Clarkson et al. (1991) Nature 352:624-628; Gram et al. (1992) Proc. Natl. Acad. Sci. USA 89:3576-3580; Garrad et al. (1991) Bio/Technology 9:1373-1377; Hoogenboom et al. 30

- 41 -

(1991) Nuc. Acid Res. 19:4133-4137; Barbas et al. (1991) Proc. Natl. Acad. Sci. USA 88:7978-7982; and McCafferty et al. Nature (1990) 348:552-554.

Additionally, recombinant anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, 5 are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson et al. International Application No. PCT/US86/02269; Akira, et al. European Patent Application 184,187; Taniguchi, M., European Patent Application 171,496; Morrison et al. European Patent Application 10 173,494; Neuberger et al. PCT International Publication No. WO 86/01533; Cabilly et al. U.S. Patent No. 4,816,567; Cabilly et al. European Patent Application 125,023; Better et al. (1988) Science 240:1041-1043; Liu et al. (1987) Proc. Natl. Acad. Sci. USA 84:3439-3443; Liu et al. (1987) J. Immunol. 139:3521-3526; Sun et al. (1987) Proc. Natl. Acad. Sci. USA 84:214-218; Nishimura et al. (1987) Canc. Res. 47:999-1005; Wood et al. (1985) Nature 314:446-449; and Shaw et al. (1988) J. Natl. Cancer Inst. 80:1553-1559); Morrison, S. L. (1985) Science 229:1202-1207; Oi et al. (1986) BioTechniques 4:214; Winter U.S. Patent 5,225,539; Jones et al. (1986) Nature 321:552-525; Verhoeyan et al. (1988) Science 239:1534; and Beidler et al. (1988) J. 20 Immunol. 141:4053-4060.

An anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody (*e.g.*, monoclonal antibody) can be used to isolate hVR-1, hVR-2, and rVR-2 by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can facilitate the purification of natural hVR-1, hVR-2, and rVR-2 from cells and of recombinantly produced hVR-1, hVR-2, and rVR-2 expressed in host cells. Moreover, an anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody can be used to detect hVR-1, hVR-2, and rVR-2 protein (*e.g.*, in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the hVR-1, hVR-2, and rVR-2 protein. Anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can

- 42 -

be facilitated by coupling (*i.e.*, physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

III. Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression 15 vectors, containing a nucleic acid encoding an hVR-1, hVR-2, and rVR-2 protein (or a portion thereof). As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, 20 wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated 25 along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most 30 commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective

- 43 -

retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, which is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner which allows for expression 10 of the nucleotide sequence (e.g., in an in vitro transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; Gene Expression Technology: Methods in Enzymology 185, 15 Academic Press, San Diego, CA (1990). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cells and those which direct expression of the nucleotide sequence only in certain host cells (e.g., tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, and the like. The 20 expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (e.g., hVR-1, hVR-2, and rVR-2 proteins, mutant forms of hVR-1, hVR-2, and rVR-2 proteins, fusion proteins, and the like).

The recombinant expression vectors of the invention can be designed for expression of hVR-1, hVR-2, and rVR-2 proteins in prokaryotic or eukaryotic cells. For example, hVR-1, hVR-2, and rVR-2 proteins can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). Alternatively, the

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recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith, D.B. and Johnson, K.S. (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Purified fusion proteins can be utilized in hVR-1, hVR-2, and rVR-2 activity assays, (e.g., direct assays or competitive assays described in detail below), or to, for example, generate antibodies specific for hVR-1, hVR-2, and rVR-2 proteins. In a embodiment, an hVR-1, hVR-2, and rVR-2 fusion protein expressed in a retroviral expression vector of the present invention can be utilized to infect bone marrow cells which are subsequently transplanted into irradiated recipients. The pathology of the subject recipient is then examined after sufficient time has passed (e.g., six (6) weeks).

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amann *et al.*, (1988) *Gene* 69:301-315) and pET 11d (Studier *et al.*, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 60-89). Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 gn10-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral

- 45 -

polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident prophage harboring a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter.

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein (Gottesman, S., *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, California (1990) 119-128). Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada *et al.*, (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the hVR-1, hVR-2, and rVR-2 expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerivisae* include pYepSec1 (Baldari, *et al.*, (1987) *Embo J.* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.*, (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, CA), and picZ (InVitrogen Corp, San Diego, CA).

Alternatively, hVR-1, hVR-2, and rVR-2 proteins can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith et al. (1983) Mol. Cell Biol. 3:2156-2165) and the pVL series (Lucklow and Summers (1989) Virology 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, B. (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J.* 6:187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see chapters 16 and 17 of Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring*

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- 46 -

Harbor Laboratory. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-5 specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert et al. (1987) Genes Dev. 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) Adv. Immunol. 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) EMBO J. 8:729-733) and immunoglobulins (Banerji et al. (1983) Cell 10 33:729-740; Queen and Baltimore (1983) Cell 33:741-748), neuron-specific promoters (e.g., the neurofilament promoter; Byrne and Ruddle (1989) Proc. Natl. Acad. Sci. USA 86:5473-5477), pancreas-specific promoters (Edlund et al. (1985) Science 230:912-916), and mammary gland-specific promoters (e.g., milk whey promoter; U.S. Patent No. 4,873,316 and European Application Publication No. 264,166). Developmentally-15 regulated promoters are also encompassed, for example the murine hox promoters (Kessel and Gruss (1990) Science 249:374-379) and the α -fetoprotein promoter (Campes and Tilghman (1989) Genes Dev. 3:537-546).

The expression characteristics of an endogenous hVR-1, hVR-2, and rVR-2 gene within a cell line or microorganism may be modified by inserting a heterologous DNA regulatory element into the genome of a stable cell line or cloned microorganism such that the inserted regulatory element is operatively linked with the endogenous hVR-1, hVR-2, and rVR-2 gene. For example, an endogenous hVR-1, hVR-2, and rVR-2 gene which is normally "trancriptionally silent", i.e., a hVR-1, hVR-2, and rVR-2 gene which is normally not expressed, or is expressed only at very low levels in a cell line or microorganism, may be activated by inserting a regulatory element which is capable of promoting the expression of a normally expressed gene product in that cell line or microorganism. Alternatively, a transcriptionally silent, endogenous hVR-1, hVR-2, and rVR-2 gene, may be activated by insertion of a promiscuous regulatory element that works across cell types.

PCT/US99/26701 WO 00/29577

- 47 -

A heterologous regulatory element may be inserted into a stable cell line or cloned microorganism, such that it is operatively linked with an endogenous hVR-1, hVR-2, and rVR-2 gene, using techniques, such as targeted homologous recombination, which are well known to those of skill in the art, and described e.g., in Chappel, U.S.

Patent No.: 5,272,071; PCT publication No. WO 91/06667, published May 16, 1991.

The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That is, the DNA molecule is operatively linked to a regulatory sequence in a manner which allows for expression (by transcription of the DNA molecule) of an RNA molecule which is antisense to hVR-1, hVR-2, and rVR-2 mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen which direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory sequences can be chosen which direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene expression using antisense genes see Weintraub, H. et al., Antisense RNA as a molecular tool for genetic analysis, Reviews - Trends in Genetics, Vol. 1(1) 1986.

Another aspect of the invention pertains to host cells into which an hVR-1, hVR-2, and rVR-2 nucleic acid molecule of the invention is introduced, e.g., an hVR-1, hVR-2, and rVR-2 nucleic acid molecule within a recombinant expression vector or an hVR-1, hVR-2, and rVR-2 nucleic acid molecule containing sequences which allow it to 25 homologously recombine into a specific site of the host cell's genome. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, 30

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- 48 -

in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, an hVR-1, hVR-2, and rVR-2 protein can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, et al. (Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding an hVR-1, hVR-2, and rVR-2 protein or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) an hVR-1, hVR-2, and rVR-2 protein.

Accordingly, the invention further provides methods for producing an hVR-1, hVR-2, and rVR-2 protein using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression

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WO 00/29577 PCT/US99/26701
- 49 -

vector encoding an hVR-1, hVR-2, and rVR-2 protein has been introduced) in a suitable medium such that an hVR-1, hVR-2, and rVR-2 protein is produced. In another embodiment, the method further comprises isolating an hVR-1, hVR-2, and rVR-2 protein from the medium or the host cell.

The host cells of the invention can also be used to produce non-human transgenic animals. For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which hVR-1, hVR-2, and rVR-2-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous hVR-1, hVR-2, and rVR-2 sequences have been introduced into their genome or homologous recombinant animals in which endogenous hVR-1, hVR-2, and rVR-2 sequences have been altered. Such animals are useful for studying the function and/or activity of an hVR-1, hVR-2, and rVR-2 and for identifying and/or evaluating modulators of hVR-1, hVR-2, and rVR-2 activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more preferably a mouse, in which an endogenous hVR-1, hVR-2, and rVR-2 gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing an hVR-1, hVR-2, and rVR-2-encoding nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The hVR-1, hVR-2, and rVR-2 cDNA sequence of SEQ ID NO:1, 3, 5, 7 or 9 can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of a hVR-2 gene, such as a

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mouse or rat hVR-2, e.g., the rVR-2 gene, can be used as a transgene. Alternatively, an hVR-1, hVR-2, and rVR-2 gene homologue, such as another member of the Capsaicin/Vanilloid family, can be isolated based on hybridization to the hVR-1, hVR-2, and rVR-2 cDNA sequences of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, (described further in subsection I above) and used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to an hVR-1, hVR-2, and rVR-2 transgene to direct expression of an hVR-1, hVR-2, and rVR-2 protein to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., Manipulating the Mouse Embryo, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of an hVR-1, hVR-2, and rVR-2 transgene in its genome and/or expression of hVR-1, hVR-2, and rVR-2 mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene encoding an hVR-1, hVR-2, and rVR-2 protein can further be bred to other transgenic animals carrying other transgenes.

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of an hVR-1, hVR-2, and rVR-2 gene into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, functionally disrupt, the hVR-1, hVR-2, and rVR-2 gene. The VR-1 or VR-2 gene can be a human gene (*e.g.*, the cDNA of SEQ ID NO:1, 3, 5, 4, 6, 7, or 9), but more preferably, is a non-human homologue of a hVR-1 and hVR-2 gene (*e.g.*, the cDNA of SEQ ID NO:10 or 12, or a cDNA isolated by stringent hybridization with the nucleotide sequence of SEQ ID NO: 1, 3, 5, 4, 6, 7, or 9). For example, a mouse VR-2 gene can be used to construct a homologous recombination nucleic acid molecule, *e.g.*, a vector, suitable for altering an endogenous VR-2 gene in the mouse genome. In a embodiment, the homologous recombination nucleic acid molecule is designed such that, upon homologous recombination, the

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endogenous hVR-1, hVR-2, and rVR-2 gene is functionally disrupted (i.e., no longer encodes a functional protein; also referred to as a "knock out" vector). Alternatively, the homologous recombination nucleic acid molecule can be designed such that, upon homologous recombination, the endogenous hVR-1, hVR-2, and rVR-2 gene is mutated or otherwise altered but still encodes functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous hVR-1, hVR-2, and rVR-2 protein). In the homologous recombination nucleic acid molecule, the altered portion of the hVR-1, hVR-2, and rVR-2 gene is flanked at its 5' and 3' ends by additional nucleic acid sequence of the hVR-1, hVR-2, and rVR-2 gene to allow for homologous recombination to occur between the exogenous hVR-1, hVR-2, and rVR-2 10 gene carried by the homologous recombination nucleic acid molecule and an endogenous hVR-1, hVR-2, and rVR-2 gene in a cell, e.g., an embryonic stem cell. The additional flanking hVR-1, hVR-2, and rVR-2 nucleic acid sequence is of sufficient length for successful homologous recombination with the endogenous gene. Typically, 15 several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the homologous recombination nucleic acid molecule (see, e.g., Thomas, K.R. and Capecchi, M. R. (1987) Cell 51:503 for a description of homologous recombination vectors). The homologous recombination nucleic acid molecule is introduced into a cell, e.g., an embryonic stem cell line (e.g., by electroporation) and cells in which the 20 introduced hVR-1, hVR-2, and rVR-2 gene has homologously recombined with the endogenous hVR-1, hVR-2, and rVR-2 gene are selected (see e.g., Li, E. et al. (1992) Cell 69:915). The selected cells can then injected into a blastocyst of an animal (e.g., a mouse) to form aggregation chimeras (see e.g., Bradley, A. in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E.J. Robertson, ed. (IRL, Oxford, 1987) 25 pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination nucleic acid molecules, e.g., vectors, or homologous recombinant animals are described further in Bradley, A. (1991) Current Opinion in Biotechnology 2:823-829 and in PCT

WO 00/29577

- 52 -

PCT/US99/26701

International Publication Nos.: WO 90/11354 by Le Mouellec *et al.*; WO 91/01140 by Smithies *et al.*; WO 92/0968 by Zijlstra *et al.*; and WO 93/04169 by Berns *et al.*

In another embodiment, transgenic non-humans animals can be produced which contain selected systems which allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, *e.g.*, Lakso *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.* (1991) *Science* 251:1351-1355. If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. (1997) Nature 385:810-813 and PCT International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G₀ phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The recontructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

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IV. Pharmaceutical Compositions

The hVR-1, hVR-2, and rVR-2 nucleic acid molecules, fragments of hVR-1, hVR-2, and rVR-2 proteins, and anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies (also referred to herein as "active compounds") of the invention can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically

- 53 -

acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, e.g., intravenous, intradermal, subcutaneous, oral (e.g., inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor ELTM (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for

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example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyetheylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a fragment of an hVR-1, hVR-2, and rVR-2 protein or an anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or

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lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, e.g., a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (e.g., with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will

protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems.

Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid.

Methods for preparation of such formulations will be apparent to those skilled in the art.

The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

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- 56 -

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (*i.e.*, the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

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- 57 -

As defined herein, a therapeutically effective amount of protein or polypeptide (i.e., an effective dosage) ranges from about 0.001 to 30 mg/kg body weight, preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight. The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments. In a preferred example, a subject is treated with antibody, protein, or polypeptide in the range of between about 0.1 to 20 mg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of antibody, protein, or polypeptide used for treatment may increase or decrease over the course of a particular treatment. Changes in dosage may result and become apparent from the results of diagnostic assays as described herein.

The present invention encompasses agents which modulate expression or activity. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (i.e., including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds.

It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon

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- 58 -

the identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention.

Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein.

When one or more of these small molecules is to be administered to an animal (e.g., a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, gender, and diet of the subject, the time of administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Patent 5,328,470) or by stereotactic injection (see *e.g.*, Chen *et al.* (1994) *Proc. Natl. Acad. Sci. USA* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, *e.g.*, retroviral vectors, the pharmaceutical preparation can include one or more cells which produce the gene delivery system.

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- 59 -

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

V. Uses and Methods of the Invention

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The nucleic acid molecules, proteins, protein homologues, and antibodies described herein can be used in one or more of the following methods: a) screening assays; b) predictive medicine (*e.g.*, diagnostic assays, prognostic assays, monitoring clinical trials, and pharmacogenetics); and c) methods of treatment (*e.g.*, therapeutic and prophylactic). As described herein, an hVR-1, hVR-2, and rVR-2 protein of the invention has one or more of the following activities: (1) it interacts with a non-hVR-1, non-hVR-2, and non-rVR-2 protein molecule, *e.g.*, a vanilloid compound such as capsaicin; (2) it modulates intracellular calcium concentration; (3) it activates an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; and (4) it modulates a pain signaling mechanism, and, thus, can be used to, for example, (1) modulate the interaction with a non-hVR-1, non-hVR-2, and non-rVR-2 protein molecule; (2) modulate intracellular calcium concentration; (3) activate an hVR-1, hVR-2, and rVR-2-dependent signal transduction pathway; and (4) modulate a pain signaling mechanism.

The isolated nucleic acid molecules of the invention can be used, for example, to express hVR-1, hVR-2, and rVR-2 protein (e.g., via a recombinant expression vector in 20 a host cell in gene therapy applications), to detect hVR-1, hVR-2, and rVR-2 mRNA (e.g., in a biological sample) or a genetic alteration in an hVR-1, hVR-2, and rVR-2 gene, and to modulate hVR-1, hVR-2, and rVR-2 activity, as described further below. The hVR-1, hVR-2, and rVR-2 proteins can be used to screen for naturally occurring hVR-1, hVR-2, and rVR-2 substrates, to screen for drugs or compounds which modulate 25 hVR-1, hVR-2, and rVR-2 activity, as well as to treat disorders characterized by insufficient or excessive production of hVR-1, hVR-2, and rVR-2 protein or production of hVR-1, hVR-2, and rVR-2 protein forms which have decreased or aberrant activity compared to hVR-1, hVR-2, and rVR-2 wild type protein (e.g., pain disorders). Moreover, the anti-hVR-1, anti-hVR-2, and anti-rVR-2 antibodies of the invention can be used to detect and isolate hVR-1, hVR-2, and rVR-2 proteins, regulate the 30

- 60 -

bioavailability of hVR-1, hVR-2, and rVR-2 proteins, and modulate hVR-1, hVR-2, and rVR-2 activity.

A. Screening Assays:

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The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, peptides, peptidomimetics, small molecules or other drugs) which bind to hVR-1, hVR-2, and rVR-2 proteins, have a stimulatory or inhibitory effect on, for example, hVR-1, hVR-2, and rVR-2 expression or hVR-1, hVR-2, and rVR-2 activity, or have a stimulatory or inhibitory effect on, for example, the expression or activity of hVR-1, hVR-2, and rVR-2 substrate.

In one embodiment, the invention provides assays for screening candidate or test compounds which are substrates of an hVR-1, hVR-2, and rVR-2 protein or polypeptide or biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of an hVR-1, hVR-2, and rVR-2 protein or polypeptide or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, K.S. (1997) *Anticancer Drug Des.* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:6909; Erb et al. (1994) Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al. (1994). J. Med. Chem. 37:2678; Cho et al. (1993) Science 261:1303; Carrell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al. (1994) Angew. Chem. Int. Ed. Engl. 33:2061; and in Gallop et al. (1994) J. Med. Chem. 37:1233.

- 61 -

Libraries of compounds may be presented in solution (e.g., Houghten (1992) Biotechniques 13:412-421), or on beads (Lam (1991) Nature 354:82-84), chips (Fodor (1993) Nature 364:555-556), bacteria (Ladner USP 5,223,409), spores (Ladner USP '409), plasmids (Cull et al. (1992) Proc Natl Acad Sci USA 89:1865-1869) or on phage (Scott and Smith (1990) Science 249:386-390); (Devlin (1990) Science 249:404-406); (Cwirla et al. (1990) Proc. Natl. Acad. Sci. 87:6378-6382); (Felici (1991) J. Mol. Biol. 222:301-310); (Ladner supra.).

In one embodiment, an assay is a cell-based assay in which a cell, e.g., a neuronal cell, which expresses an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity is determined. Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity can be accomplished by monitoring, for example, intracellular calcium concentration or membrane depolarization by, e.g., patch-clamp recordings in whole-cell, inside-out, and outside-out configurations (as described in, for example, Tominaga M. et al. (1998) Neuron 21:531-543). Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 activity can further be accomplished by monitoring the activity of an hVR-1, hVR-2, and rVR-2-regulated transcription factor. The cell, for example, can be of mammalian origin, e.g., a neuronal cell.

The ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 binding to a substrate or to bind to hVR-1, hVR-2, and rVR-2 can also be determined.

Determining the ability of the test compound to modulate hVR-1, hVR-2, and rVR-2 binding to a substrate can be accomplished, for example, by coupling the hVR-1, hVR-2, and rVR-2 substrate with a radioisotope or enzymatic label such that binding of the hVR-1, hVR-2, and rVR-2 substrate to hVR-1, hVR-2, and rVR-2 can be determined by detecting the labeled hVR-1, hVR-2, and rVR-2 substrate in a complex. Determining the ability of the test compound to bind hVR-1, hVR-2, and rVR-2 can be accomplished, for example, by coupling the compound with a radioisotope or enzymatic label such that binding of the compound to hVR-1, hVR-2, and rVR-2 can be determined by detecting the labeled hVR-1, hVR-2, and rVR-2 compound in a complex. For example, compounds (e.g., hVR-1, hVR-2, and rVR-2 substrates) can be labeled with ¹²⁵I, ³⁵S.

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14C, or 3H, either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

It is also within the scope of this invention to determine the ability of a compound (e.g., an hVR-1, hVR-2, and rVR-2 substrate) to interact with hVR-1, hVR-2, and rVR-2 without the labeling of any of the interactants. For example, a microphysiometer can be used to detect the interaction of a compound with hVR-1, hVR-2, and rVR-2 without the labeling of either the compound or the hVR-1, hVR-2, and rVR-2. McConnell, H. M. et al. (1992) Science 257:1906-1912. As used herein, a "microphysiometer" (e.g., Cytosensor) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between a compound and hVR-1, hVR-2, and rVR-2.

In yet another embodiment, an assay of the present invention is a cell-free assay in which an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is determined. biologically active portions of the hVR-1, hVR-2, and rVR-2 proteins to be used in assays of the present invention include fragments which participate in interactions with non-hVR-1, non-hVR-2, and non-rVR-2 molecules, e.g., fragments with high surface probability scores. Binding of the test compound to the hVR-1, hVR-2, and rVR-2 protein can be determined either directly or indirectly as described above. In a embodiment, the assay includes contacting the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof with a known compound which binds hVR-1, hVR-2, and rVR-2 to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with an hVR-1, hVR-2, and rVR-2 protein, wherein determining the ability of the test compound to interact with an hVR-1, hVR-2, and rVR-2 protein comprises determining the ability of the test 30

compound to preferentially bind to hVR-1, hVR-2, and rVR-2 or biologically active portion thereof as compared to the known compound.

In another embodiment, the assay is a cell-free assay in which an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to modulate (e.g., stimulate or inhibit) the activity of the hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof is determined. Determining the ability of the test compound to modulate the activity of an hVR-1, hVR-2, and rVR-2 protein can be accomplished, for example, by determining the ability of the hVR-1, hVR-2, and rVR-2 protein to bind to an hVR-1, hVR-2, and rVR-2 target molecule, e.g., a vanilloid compound such as capsaicin, by one of the methods described above for determining direct binding. Determining the ability of the hVR-1, hVR-2, and rVR-2 protein to bind to an hVR-1, hVR-2, and rVR-2 target molecule can also be accomplished using a technology such as real-time Biomolecular Interaction Analysis (BIA). Sjolander, S. and Urbaniczky, C. (1991) Anal. Chem. 63:2338-2345 and Szabo et al. (1995) Curr. Opin. Struct. Biol. 5:699-705. As used herein, "BIA" is a technology for studying biospecific interactions in real time, without labeling any of the interactants (e.g., BIAcore). Changes in the optical phenomenon of surface plasmon resonance (SPR) can be used as an indication of real-time reactions between biological molecules.

In an alternative embodiment, determining the ability of the test compound to modulate the activity of an hVR-1, hVR-2, and rVR-2 protein can be accomplished by determining the ability of the hVR-1, hVR-2, and rVR-2 protein to further modulate the activity of a downstream effector of an hVR-1, hVR-2, and rVR-2 target molecule. For example, the activity of the effector molecule on an appropriate target can be determined or the binding of the effector to an appropriate target can be determined as previously described.

In yet another embodiment, the cell-free assay involves contacting an hVR-1, hVR-2, and rVR-2 protein or biologically active portion thereof with a known compound which binds the hVR-1, hVR-2, and rVR-2 protein to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the hVR-1, hVR-2, and rVR-2 protein, wherein

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determining the ability of the test compound to interact with the hVR-1, hVR-2, and rVR-2 protein comprises determining the ability of the hVR-1, hVR-2, and rVR-2 protein to preferentially bind to or modulate the activity of an hVR-1, hVR-2, and rVR-2 target molecule.

The cell-free assays of the present invention are amenable to use of both soluble and/or membrane-bound forms of isolated proteins (*e.g.*, hVR-1, hVR-2, and rVR-2 proteins or biologically active portions thereof). In the case of cell-free assays in which a membrane-bound form of an isolated protein is used it may be desirable to utilize a solubilizing agent such that the membrane-bound form of the isolated protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, 3-[(3-cholamidopropyl)dimethylamminio]-1-propane sulfonate (CHAPS), 3-[(3-cholamidopropyl)dimethylamminio]-2-hydroxy-1-propane sulfonate (CHAPSO), or N-dodecyl=N,N-dimethyl-3-ammonio-1-propane sulfonate.

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either hVR-1, hVR-2, and rVR-2 or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to an hVR-1, hVR-2, and rVR-2 protein, or interaction of an hVR-1, hVR-2, and rVR-2 protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtitre plates, test tubes, and microcentrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase/hVR-1, hVR-2, and rVR-2 fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or hVR-1, hVR-2, and rVR-2 protein, and the mixture incubated under

conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtitre plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of hVR-1, hVR-2, and rVR-2 binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either an hVR-1, hVR-2, and rVR-2 protein or an hVR-1, hVR-2, and rVR-2 target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated hVR-1, hVR-2, and rVR-2 protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with hVR-1, hVR-2, and rVR-2 protein or target molecules but which do not interfere with binding of the hVR-1, hVR-2, and rVR-2 protein to its target molecule can be derivatized to the wells of the plate, and unbound target or hVR-1, hVR-2, and rVR-2 protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the hVR-1, hVR-2, and rVR-2 protein or target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the hVR-1, hVR-2, and rVR-2 protein or target molecule.

In another embodiment, modulators of hVR-1, hVR-2, and rVR-2 expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the cell is determined. The level of expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the presence of the candidate compound is compared to the level of expression of hVR-1, hVR-2, and rVR-2 mRNA or protein in the absence of the candidate compound. The candidate compound can then be identified as a modulator of hVR-1, hVR-2, and rVR-2 expression based on this comparison. For example, when expression of hVR-1, hVR-2, and rVR-2 mRNA or protein is greater (statistically significantly greater) in the presence

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of the candidate compound than in its absence, the candidate compound is identified as a stimulator of hVR-1, hVR-2, and rVR-2 mRNA or protein expression. Alternatively, when expression of hVR-1, hVR-2, and rVR-2 mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of hVR-1, hVR-2, and rVR-2 mRNA or protein expression. The level of hVR-1, hVR-2, and rVR-2 mRNA or protein expression in the cells can be determined by methods described herein for detecting hVR-1, hVR-2, and rVR-2 mRNA or protein.

In yet another aspect of the invention, the hVR-1, hVR-2, and rVR-2 proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al. (1993) Cell 72:223-232; Madura et al. (1993) J. Biol. Chem. 268:12046-12054; Bartel et al. (1993) Biotechniques 14:920-924; Iwabuchi et al. (1993) Oncogene 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with hVR-1, hVR-2, and rVR-2 ("hVR-1-binding proteins", "hVR-2-binding proteins", and "rVR-2-binding proteins" or "hVR-1-bp", 15 "hVR-2-bp", and "rVR-2-bp") and are involved in hVR-1, hVR-2, and rVR-2 activity. Such hVR-1, hVR-2, and rVR-2-binding proteins are also likely to be involved in the propagation of signals by the hVR-1, hVR-2, and rVR-2 proteins or hVR-1, hVR-2, and rVR-2 targets as, for example, downstream elements of an hVR-1, hVR-2, and rVR-2mediated signaling pathway, e.g., a pain signaling pathway. Alternatively, such hVR-1, 20 hVR-2, and rVR-2-binding proteins are likely to be hVR-1, hVR-2, and rVR-2 inhibitors.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for an hVR-1, hVR-2, and rVR-2 protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, in vivo, forming an hVR-1, hVR-2, and rVR-2-dependent complex, the DNA-binding and

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activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the hVR-1, hVR-2, and rVR-2 protein.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified as described herein (e.g., an hVR-1, hVR-2, and rVR-2 modulating agent, an antisense hVR-1, hVR-2, and rVR-2 nucleic acid molecule, an hVR-1, hVR-2, and rVR-2-specific antibody, or an hVR-1, hVR-2, and rVR-2-binding partner) can be used in an animal model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

Detection Assays В.

Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; (ii) identify an individual from a minute biological sample (tissue typing); and (iii) aid in forensic identification of a biological sample. These applications 25 are described in the subsections below.

1. Chromosome Mapping

Once the sequence (or a portion of the sequence) of a gene has been isolated, this sequence can be used to map the location of the gene on a chromosome. This process is 30 called chromosome mapping. Accordingly, portions or fragments of the hVR-1, hVR-2,

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and rVR-2 nucleotide sequences, described herein, can be used to map the location of the hVR-1, hVR-2, and rVR-2 genes on a chromosome. The mapping of the hVR-1, hVR-2, and rVR-2 sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease.

Briefly, hVR-1, hVR-2, and rVR-2 genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the hVR-1, hVR-2, and rVR-2 nucleotide sequences. Computer analysis of the hVR-1, hVR-2, and rVR-2 sequences can be used to predict primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the hVR-1, hVR-2, and rVR-2 sequences will yield an amplified fragment.

Somatic cell hybrids are prepared by fusing somatic cells from different mammals (e.g., human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but human cells can, the one human chromosome that contains the gene encoding the needed enzyme, will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of individual genes to specific human chromosomes. (D'Eustachio P. et al. (1983) Science 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycler. Using the hVR-1, hVR-2, and rVR-2 nucleotide sequences to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes. Other mapping strategies which can similarly be used to map an hVR-1, hVR-2, and rVR-2 sequence to its chromosome include *in situ* hybridization (described in Fan, Y. *et al.* (1990) *Proc. Natl. Acad. Sci.*

USA, 87:6223-27), pre-screening with labeled flow-sorted chromosomes, and pre-selection by hybridization to chromosome specific cDNA libraries.

Fluorescence *in situ* hybridization (FISH) of a DNA sequence to a metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical such as colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark bands develops on each chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a DNA sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000 bases, and more preferably 2,000 bases will suffice to get good results at a reasonable amount of time. For a review of this technique, see Verma *et al.*, Human Chromosomes: A Manual of Basic Techniques (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used for marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. (Such data are found, for example, in V. McKusick, Mendelian Inheritance in Man, available on-line through Johns Hopkins University Welch Medical Library). The relationship between a gene and a disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example, Egeland, J. et al. (1987) Nature, 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the hVR-1, hVR-2, and rVR-2 gene, can be determined. If a mutation is observed in some or all of the affected individuals but not

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in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

2. Tissue Typing

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The hVR-1, hVR-2, and rVR-2 sequences of the present invention can also be used to identify individuals from minute biological samples. The United States military, for example, is considering the use of restriction fragment length polymorphism (RFLP) for identification of its personnel. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for identification. This method does not suffer from the current limitations of "Dog Tags" which can be lost, switched, or stolen, making positive identification difficult. The sequences of the present invention are useful as additional DNA markers for RFLP (described in U.S. Patent 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an alternative technique which determines the actual base-by-base DNA sequence of selected portions of an individual's genome. Thus, the hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The hVR-1, hVR-2, and rVR-2 nucleotide sequences of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding

- 71 -

regions. It is estimated that allelic variation between individual humans occurs with a frequency of about once per each 500 bases. Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals.

If a panel of reagents from hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein is used to generate a unique identification database for an individual, those same reagents can later be used to identify tissue from that individual. Using the unique identification database, positive identification of the individual, living or dead, can be made from extremely small tissue samples.

3. Use of Partial hVR-1, hVR-2, and rVR-2 Sequences in Forensic Biology

DNA-based identification techniques can also be used in forensic biology. Forensic biology is a scientific field employing genetic typing of biological evidence found at a crime scene as a means for positively identifying, for example, a perpetrator of a crime. To make such an identification, PCR technology can be used to amplify DNA sequences taken from very small biological samples such as tissues, *e.g.*, hair or skin, or body fluids, *e.g.*, blood, saliva, or semen found at a crime scene. The amplified sequence can then be compared to a standard, thereby allowing identification of the origin of the biological sample.

The sequences of the present invention can be used to provide polynucleotide reagents, e.g., PCR primers, targeted to specific loci in the human genome, which can enhance the reliability of DNA-based forensic identifications by, for example, providing another "identification marker" (i.e. another DNA sequence that is unique to a particular individual). As mentioned above, actual base sequence information can be used for identification as an accurate alternative to patterns formed by restriction enzyme generated fragments. Examples of polynucleotide reagents include the hVR-1, hVR-2, and rVR-2 nucleotide sequences or portions thereof, e.g., fragments derived from SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 11 having a length of at least 20 bases, preferably at least 30 bases.

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The hVR-1, hVR-2, and rVR-2 nucleotide sequences described herein can further be used to provide polynucleotide reagents, e.g., labeled or labelable probes which can be used in, for example, an *in situ* hybridization technique, to identify a specific tissue, e.g., brain tissue. This can be very useful in cases where a forensic pathologist is presented with a tissue of unknown origin. Panels of such hVR-1, hVR-2, and rVR-2 probes can be used to identify tissue by species and/or by organ type.

In a similar fashion, these reagents, e.g., hVR-1, hVR-2, and rVR-2 primers or probes can be used to screen tissue culture for contamination (i.e. screen for the presence of a mixture of different types of cells in a culture).

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C. Predictive Medicine:

The present invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for 15 determining hVR-1, hVR-2, and rVR-2 protein and/or nucleic acid expression as well as hVR-1, hVR-2, and rVR-2 activity, in the context of a biological sample (e.g., blood, serum, cells, tissue) to thereby determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. The invention also provides for prognostic 20 (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with hVR-1, hVR-2, and rVR-2 protein, nucleic acid expression or activity. For example, mutations in an hVR-1, hVR-2, and rVR-2 gene can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to thereby phophylactically treat an individual prior to the onset of a disorder characterized 25 by or associated with hVR-1, hVR-2, and rVR-2 protein, nucleic acid expression or activity.

Another aspect of the invention pertains to monitoring the influence of agents (e.g., drugs, compounds) on the expression or activity of hVR-1, hVR-2, and rVR-2 in clinical trials.

These and other agents are described in further detail in the following sections.

1. Diagnostic Assays

An exemplary method for detecting the presence or absence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting hVR-1, hVR-2, and rVR-2 protein or nucleic acid (*e.g.*, mRNA, genomic DNA) that encodes hVR-1, hVR-2, and rVR-2 protein such that the presence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid is detected in the biological sample. A agent for detecting hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA. The nucleic acid probe can be, for example, a full-length hVR-1, hVR-2, and rVR-2 nucleic acid, such as the nucleic acid of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to hVR-1, hVR-2, and rVR-2 mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

An agent for detecting hVR-1, hVR-2, and rVR-2 protein is an antibody capable of binding to hVR-1, hVR-2, and rVR-2 protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof $(e.g., Fab \text{ or } F(ab')_{2})$ can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect hVR-1, hVR-2, and rVR-2 mRNA, protein, or genomic DNA in a biological sample in vitro as well as in vivo. For example, in vitro techniques for detection of hVR-1, hVR-2, and rVR-2 mRNA include Northern hybridizations and in situ hybridizations. In vitro

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- 74 -

techniques for detection of hVR-1, hVR-2, and rVR-2 protein include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of hVR-1, hVR-2, and rVR-2 genomic DNA include Southern hybridizations. Furthermore, *in vivo* techniques for detection of hVR-1, hVR-2, and rVR-2 protein include introducing into a subject a labeled anti-hVR-1, hVR-2, and rVR-2 antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A biological sample is a serum sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA, such that the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA in the control sample with the presence of hVR-1, hVR-2, and rVR-2 protein, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting the presence of hVR-1, hVR-2, and rVR-2 in a biological sample. For example, the kit can comprise a labeled compound or agent capable of detecting hVR-1, hVR-2, and rVR-2 protein or mRNA in a biological sample; means for determining the amount of hVR-1, hVR-2, and rVR-2 in the sample; and means for comparing the amount of hVR-1, hVR-2, and rVR-2 in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect hVR-1, hVR-2, and rVR-2 protein or nucleic acid.

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2. Prognostic Assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. As used herein, the term "aberrant" includes an hVR-1, hVR-2, and rVR-2 expression or activity which deviates from the wild type hVR-1, hVR-2, and rVR-2 expression or activity. Aberrant expression or activity includes increased or decreased expression or activity, as well as expression or activity which does not follow the wild type developmental pattern of expression or the subcellular pattern of expression. For example, aberrant hVR-1, hVR-2, and rVR-2 expression or activity is intended to include the cases in which a mutation in the hVR-1, hVR-2, and rVR-2 gene causes the hVR-1, hVR-2, and rVR-2 gene to be underexpressed or over-expressed and situations in which such mutations result in a non-functional hVR-1, hVR-2, and rVR-2 protein or a protein which does not function in a wild-type fashion, e.g., a protein which does not interact with an hVR-1, hVR-2, and rVR-2 ligand or one which interacts with a non-hVR-1, non-hVR-2, and non-rVR-2 ligand.

The assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing a disorder associated with a misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a pain disorder. Alternatively, the prognostic assays can be utilized to identify a subject having or at risk for developing a disorder associated with a misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a pain disorder. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity in which a test sample is obtained from a subject and hVR-1, hVR-2, and rVR-2 protein or nucleic acid (e.g., mRNA or genomic DNA) is detected, wherein the presence of hVR-1, hVR-2, and rVR-2 protein or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest. For example, a test sample can be a biological fluid (e.g., serum), cell sample, or tissue.

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- 76 -

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. For example, such methods can be used to determine whether a subject can be effectively treated with an agent for a pain disorder. Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity in which a test sample is obtained and hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity is detected (e.g., wherein the abundance of hVR-1, hVR-2, and rVR-2 protein or nucleic acid expression or activity is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity).

The methods of the invention can also be used to detect genetic alterations in an hVR-1, hVR-2, and rVR-2 gene, thereby determining if a subject with the altered gene is 15 at risk for a disorder characterized by misregulation in hVR-1, hVR-2, and rVR-2 protein activity or nucleic acid expression, such as a neurodegenerative disorder. In embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic alteration characterized by at least one of an alteration affecting the integrity of a gene encoding an hVR-1, hVR-2, and rVR-2-protein, or the 20 mis-expression of the hVR-1, hVR-2, and rVR-2 gene. For example, such genetic alterations can be detected by ascertaining the existence of at least one of 1) a deletion of one or more nucleotides from an hVR-1, hVR-2, and rVR-2 gene; 2) an addition of one or more nucleotides to an hVR-1, hVR-2, and rVR-2 gene; 3) a substitution of one or more nucleotides of an hVR-1, hVR-2, and rVR-2 gene, 4) a chromosomal 25 rearrangement of an hVR-1, hVR-2, and rVR-2 gene; 5) an alteration in the level of a messenger RNA transcript of an hVR-1, hVR-2, and rVR-2 gene, 6) aberrant modification of an hVR-1, hVR-2, and rVR-2 gene, such as of the methylation pattern of the genomic DNA, 7) the presence of a non-wild type splicing pattern of a messenger RNA transcript of an hVR-1, hVR-2, and rVR-2 gene, 8) a non-wild type level of an 30 hVR-1, hVR-2, and rVR-2-protein, 9) allelic loss of an hVR-1, hVR-2, and rVR-2 gene,

- 77 -

and 10) inappropriate post-translational modification of an hVR-1, hVR-2, and rVR-2-protein. As described herein, there are a large number of assays known in the art which can be used for detecting alterations in an hVR-1, hVR-2, and rVR-2 gene. A biological sample is a tissue or serum sample isolated by conventional means from a subject.

In certain embodiments, detection of the alteration involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al. (1988) Science 241:1077-1080; and Nakazawa et al. (1994) Proc. Natl. Acad. Sci. USA 91:360-364), the latter of which can be particularly useful for detecting point mutations in the hVR-1, hVR-2, and rVR-2-gene (see Abravaya et al. (1995) Nucleic Acids Res .23:675-682). This method can include the steps of collecting a sample of cells from a subject, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to an hVR-1, hVR-2, and rVR-2 gene under conditions such that hybridization and amplification of the hVR-1, hVR-2, and rVR-2-gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli, J.C. et al., (1990) Proc. Natl. Acad. Sci. USA 87:1874-1878), transcriptional amplification system (Kwoh, D.Y. et al., (1989) Proc. Natl. Acad. Sci. USA 86:1173-1177), Q-Beta Replicase (Lizardi, P.M. et al. (1988) Bio-Technology 6:1197), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in an hVR-1, hVR-2, and rVR-2 gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally),

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digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Patent No. 5,498,531) can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in hVR-1, hVR-2, and rVR-2 can be identified by hybridizing a sample and control nucleic acids, e.g., DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin, M.T. et al. (1996) Human Mutation 7: 244-255; Kozal, M.J. et al. (1996) Nature 10 Medicine 2: 753-759). For example, genetic mutations in hVR-1, hVR-2, and rVR-2 can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin, M.T. et al. supra. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping 15 probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene. 20

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence the hVR-1, hVR-2, and rVR-2 gene and detect mutations by comparing the sequence of the sample hVR-1, hVR-2, and rVR-2 with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on techniques developed by Maxam and Gilbert ((1977) *Proc. Natl. Acad. Sci. USA* 74:5463). It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the diagnostic assays ((1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, *e.g.*, PCT International Publication No. WO 94/16101; Cohen *et al.* (1996) *Adv. Chromatogr.* 36:127-162; and Griffin *et al.* (1993) *Appl. Biochem. Biotechnol.* 38:147-159).

- 79 -

Other methods for detecting mutations in the hVR-1, hVR-2, and rVR-2 gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers et al. (1985) Science 230:1242). In general, the art technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wildtype hVR-1, hVR-2, and rVR-2 sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent which cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton et al. (1988) Proc. Natl Acad Sci USA 85:4397; Saleeba et al. (1992) Methods Enzymol. 217:286-295. In a embodiment, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in hVR-1, hVR-2, and rVR-2 cDNAs obtained from samples of cells. For example, the mutY enzyme of E. coli cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu et al. (1994) Carcinogenesis 15:1657-1662). According to an exemplary embodiment, a probe based on an hVR-1, hVR-2, and rVR-2 sequence, e.g., a wild-type hVR-1, hVR-2, and rVR-2 sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Patent No. 5,459,039.

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- 80 -

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in hVR-1, hVR-2, and rVR-2 genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (orita et al. (1989) Proc Natl. Acad. Sci USA: 86:2766, see also Cotton (1993) Mutat. Res. 285:125-144; and Hayashi (1992) Genet. Anal. Tech. Appl. 9:73-79). Single-stranded DNA fragments of sample and control hVR-1, hVR-2, and rVR-2 nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to sequence, the resulting alteration in electrophoretic mobility enables the detection of 10 even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In a embodiment, the subject method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility 15 (Keen et al. (1991) Trends Genet 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers et al. (1985) Nature 313:495). When DGGE is used as the method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) Biophys Chem 265:12753).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions which permit hybridization only if a perfect match is found (Saiki et al. (1986) Nature 324:163); Saiki et al. (1989) Proc. Natl Acad. Sci USA 86:6230). Such allele specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different

- 81 -

mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology which depends on selective PCR amplification may be used in conjunction with the instant invention.

Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs et al. (1989) Nucleic Acids Res. 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner (1993) Tibtech 11:238). In addition it may be desirable to introduce a novel restriction site in the region of the mutation to create cleavage-based detection (Gasparini et al. (1992) Mol. Cell Probes 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) Proc. Natl. Acad. Sci USA 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing prepackaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, e.g., in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving an hVR-1, hVR-2, and rVR-2 gene.

Furthermore, any cell type or tissue in which hVR-1, hVR-2, and rVR-2 is expressed may be utilized in the prognostic assays described herein.

3. Monitoring of Effects During Clinical Trials

Monitoring the influence of agents (e.g., drugs) on the expression or activity of an hVR-1, hVR-2, and rVR-2 protein can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or upregulate hVR-1, hVR-2, and rVR-2 activity, can be monitored in clinical trials of subjects exhibiting decreased hVR-1, hVR-2, and rVR-2

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- 82 -

gene expression, protein levels, or downregulated hVR-1, hVR-2, and rVR-2 activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or downregulate hVR-1, hVR-2, and rVR-2 activity, can be monitored in clinical trials of subjects exhibiting increased hVR-1, hVR-2, and rVR-2 gene expression, protein levels, or upregulated hVR-1, hVR-2, and rVR-2 activity. In such clinical trials, the expression or activity of an hVR-1, hVR-2, and rVR-2 gene, and preferably, other genes that have been implicated in, for example, an hVR-1, hVR-2, and rVR-2-associated disorder can be used as a "read out" or markers of the phenotype of a particular cell.

For example, and not by way of limitation, genes, including hVR-1, hVR-2, and rVR-2, that are modulated in cells by treatment with an agent (e.g., compound, drug or small molecule) which modulates hVR-1, hVR-2, and rVR-2 activity (e.g., identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on hVR-1, hVR-2, and rVR-2-associated disorders (e.g., pain disorders), for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of hVR-1, hVR-2, and rVR-2 and other genes implicated in the hVR-1, hVR-2, and rVR-2-associated disorder, respectively. The levels of gene expression (e.g., a gene expression pattern) can be quantified by northern blot analysis or RT-PCR, as described herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of hVR-1, hVR-2, and rVR-2 or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be determined before, and at various points during treatment of the individual with the agent.

In a embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (e.g., an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) including the steps of (i) obtaining a pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of an hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the preadministration sample; (iii) obtaining one or more post-

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- 83 -

administration samples from the subject; (iv) detecting the level of expression or activity of the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the pre-administration sample with the hVR-1, hVR-2, and rVR-2 protein, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of hVR-1, hVR-2, and rVR-2 to higher levels than detected, *i.e.*, to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of hVR-1, hVR-2, and rVR-2 to lower levels than detected, *i.e.* to decrease the effectiveness of the agent. According to such an embodiment, hVR-1, hVR-2, and rVR-2 expression or activity may be used as an indicator of the effectiveness of an agent, even in the absence of an observable phenotypic response.

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D. Methods of Treatment:

The present invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant hVR-1, hVR-2, and rVR-2 expression or activity. With regards to both prophylactic and therapeutic methods of treatment, such treatments may be specifically tailored or modified, based on knowledge obtained from the field of pharmacogenomics. "Pharmacogenomics", as used herein, refers to the application of genomics technologies such as gene sequencing, statistical genetics, and gene expression analysis to drugs in clinical development and on the market. More specifically, the term refers the study of how a patient's genes determine his or her response to a drug (e.g., a patient's "drug response phenotype", or "drug response genotype".) Thus, another aspect of the invention provides methods for tailoring an individual's prophylactic or therapeutic treatment with either the hVR-1, hVR-2, and rVR-2 molecules of the present invention or hVR-1, hVR-2, and rVR-2 modulators according to that individual's drug response genotype. Pharmacogenomics allows a clinician or physician to target prophylactic or

- 84 -

therapeutic treatments to patients who will most benefit from the treatment and to avoid treatment of patients who will experience toxic drug-related side effects.

1. Prophylactic Methods

5 In one aspect, the invention provides a method for preventing in a subject, a disease or condition associated with an aberrant hVR-1, hVR-2, and rVR-2 expression or activity, by administering to the subject an hVR-1, hVR-2, and rVR-2 or an agent which modulates hVR-1, hVR-2, and rVR-2 expression or at least one hVR-1, hVR-2, and rVR-2 activity. Subjects at risk for a disease which is caused or contributed to by aberrant hVR-1, hVR-2, and rVR-2 expression or activity can be identified by, for 10 example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the hVR-1, hVR-2, and rVR-2 aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of hVR-15 1. hVR-2, and rVR-2 aberrancy, for example, an hVR-1, hVR-2, and rVR-2, hVR-1, hVR-2, and rVR-2 agonist or hVR-1, hVR-2, and rVR-2 antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein.

20 2. Therapeutic Methods

Another aspect of the invention pertains to methods of modulating hVR-1, hVR-2, and rVR-2 expression or activity for therapeutic purposes. Accordingly, in an exemplary embodiment, the modulatory method of the invention involves contacting a cell with an hVR-1, hVR-2, and rVR-2 or agent that modulates one or more of the activities of hVR-1, hVR-2, and rVR-2 protein activity associated with the cell. An agent that modulates hVR-1, hVR-2, and rVR-2 protein activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring target molecule of an hVR-1, hVR-2, and rVR-2 protein (e.g., an hVR-1, hVR-2, and rVR-2 substrate), an hVR-1, hVR-2, and rVR-2 antibody, an hVR-1, hVR-2, and rVR-2 agonist or antagonist, a peptidomimetic of an hVR-1, hVR-2, and rVR-2 agonist or antagonist, or other small molecule. In one embodiment, the agent stimulates one or more hVR-1,

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- 85 -

hVR-2, and rVR-2 activities. Examples of such stimulatory agents include active hVR-1, hVR-2, and rVR-2 protein and a nucleic acid molecule encoding hVR-1, hVR-2, and rVR-2 that has been introduced into the cell. In another embodiment, the agent inhibits one or more hVR-1, hVR-2, and rVR-2 activities. Examples of such inhibitory agents include antisense hVR-1, hVR-2, and rVR-2 nucleic acid molecules, anti-hVR-1, hVR-2, and rVR-2 antibodies, and hVR-1, hVR-2, and rVR-2 inhibitors. These modulatory methods can be performed in vitro (e.g., by culturing the cell with the agent) or, alternatively, in vivo (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of an hVR-1, hVR-2, and rVR-2 protein or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) hVR-1, hVR-2, and rVR-2 expression or activity. In another embodiment, the method involves administering an hVR-1, hVR-2, and rVR-2 protein or nucleic acid molecule as therapy to compensate for reduced or aberrant hVR-1, hVR-2, and rVR-2 expression or activity.

Stimulation of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which hVR-1, hVR-2, and rVR-2 is abnormally downregulated and/or in which increased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect. For example, stimulation of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which an hVR-1, hVR-2, and rVR-2 is downregulated and/or in which increased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect. Likewise, inhibition of hVR-1, hVR-2, and rVR-2 activity is desirable in situations in which hVR-1, hVR-2, and rVR-2 is abnormally upregulated and/or in which decreased hVR-1, hVR-2, and rVR-2 is abnormally upregulated and/or in which decreased hVR-1, hVR-2, and rVR-2 activity is likely to have a beneficial effect.

3. Pharmacogenomics

The hVR-1, hVR-2, and rVR-2 molecules of the present invention, as well as agents, or modulators which have a stimulatory or inhibitory effect on hVR-1, hVR-2, and rVR-2 activity (e.g., hVR-1, hVR-2, and rVR-2 gene expression) as identified by a screening assay described herein can be administered to individuals to treat

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- 86 -

(prophylactically or therapeutically) hVR-1, hVR-2, and rVR-2-associated disorders (e.g., pain disorders) associated with aberrant hVR-1, hVR-2, and rVR-2 activity. In conjunction with such treatment, pharmacogenomics (i.e., the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, a physician or clinician may consider applying knowledge obtained in relevant pharmacogenomics studies in determining whether to administer an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator as well as tailoring the dosage and/or therapeutic regimen of treatment with an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, for example, Eichelbaum, M. et al. (1996) Clin. Exp. Pharmacol. Physiol. 23(10-11):983-985 and Linder, M.W. et al. (1997) Clin. Chem. 43(2):254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur either as rare genetic defects or as naturally-occurring polymorphisms. For example, glucose-6-phosphate dehydrogenase deficiency (G6PD) is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

One pharmacogenomics approach to identifying genes that predict drug response, known as "a genome-wide association", relies primarily on a high-resolution map of the human genome consisting of already known gene-related markers (e.g., a "bi-allelic" gene marker map which consists of 60,000-100,000 polymorphic or variable sites on the human genome, each of which has two variants.) Such a high-resolution genetic map can be compared to a map of the genome of each of a statistically

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- 87 -

significant number of patients taking part in a Phase II/III drug trial to identify markers associated with a particular observed drug response or side effect. Alternatively, such a high resolution map can be generated from a combination of some ten-million known single nucleotide polymorphisms (SNPs) in the human genome. As used herein, a "SNP" is a common alteration that occurs in a single nucleotide base in a stretch of DNA. For example, a SNP may occur once per every 1000 bases of DNA. A SNP may be involved in a disease process, however, the vast majority may not be disease-associated. Given a genetic map based on the occurrence of such SNPs, individuals can be grouped into genetic categories depending on a particular pattern of SNPs in their individual genome. In such a manner, treatment regimens can be tailored to groups of genetically similar individuals, taking into account traits that may be common among such genetically similar individuals.

Alternatively, a method termed the "candidate gene approach", can be utilized to identify genes that predict drug response. According to this method, if a gene that encodes a drugs target is known (e.g., an hVR-1, hVR-2, and rVR-2 protein of the present invention), all common variants of that gene can be fairly easily identified in the population and it can be determined if having one version of the gene versus another is associated with a particular drug response.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a 20 major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (e.g., N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive 25 metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently 30 experience exaggerated drug response and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as

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demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Alternatively, a method termed the "gene expression profiling", can be utilized to identify genes that predict drug response. For example, the gene expression of an animal dosed with a drug (e.g., an hVR-1, hVR-2, and rVR-2 molecule or hVR-1, hVR-2, and rVR-2 modulator of the present invention) can give an indication whether gene pathways related to toxicity have been turned on.

Information generated from more than one of the above pharmacogenomics approaches can be used to determine appropriate dosage and treatment regimens for prophylactic or therapeutic treatment an individual. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with an hVR-1, hVR-2, and rVR-2 modulator, such as a modulator identified by one of the exemplary screening assays described herein.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the Figures and the Sequence Listing are incorporated herein by reference.

EXAMPLES

25 EXAMPLE 1:

IDENTIFICATION AND CHARACTERIZATION OF hVR-1, hVR-2, and rVR-2 cDNA

In this example, the identification and characterization of the genes encoding hVR-1 (clone Fchrb87a6), hVR-2 (clone flh21e11), hVR-2 alternate form (clone frhob12c4), and rVR-2 (clone flrxb147g11) are described.

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Isolation of the hVR-1, hVR-2, and the rVR-2 cDNA

The invention is based, at least in part, on the discovery of two human genes and one rat gene encoding novel members of the Capsaicin/Vanilloid receptor family, referred to herein as hVR-1, hVR-2, and rVR-2, respectively. These clones were identified from a human heart library and a rat dorsal root ganglion (DRG) library, based on sequence homology to the known rat VR-1 (Accession Number AF029310). The sequence of the two human clones and the rat clone was determined and found to contain open reading frames.

The nucleotide sequence of the full length hVR-1 cDNA and the predicted amino acid sequence of the hVR-1 polypeptide are shown in Figure 1 and in SEQ ID NOs:1 and 2, respectively.

The nucleotide sequence of the full length hVR-2 cDNA and the predicted amino acid sequence of the hVR-2 polypeptide are shown in Figure 2 and in SEQ ID NOs:4 and 5, respectively.

The nucleotide sequence of the partial hVR-2 (alternate form) cDNA and the predicted amino acid sequence of the hVR-2 (alternate form) polypeptide are shown in Figure 3 and in SEQ ID NOs:7 and 8, respectively.

The amino acid sequence of the predicted full length human VR-2 protein (alternate form) is shown in Figure 16 and in SEQ ID NO:20.

The nucleotide sequence of the partial rVR-2 cDNA and the predicted amino acid sequence of the rVR-2 polypeptide are shown in Figure 4 and in SEQ ID NOs:10 and 11, respectively.

Analysis of the hVR-1, hVR-2, and rVR-2 Molecules

The hVR-1 protein (SEQ ID NO:2) was aligned with the human VR-2 protein (SEQ ID NO:5) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 46.348% identity and 55.378% similarity between the two sequences (see Figure 5).

The hVR-1 nucleotide sequence (SEQ ID NO:1) was aligned with the human

VR-2 nucleotide sequence (SEQ ID NO:4) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3. The

- 90 -

results showed a 55.316% identity and 55.316% similarity between the two sequences (see Figure 6).

The hVR-2 protein (SEQ ID NO:5) was aligned with the rat VR-2 protein (SEQ ID NO:11) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 7), as well as using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 79.167% identity and 81.703% similarity between the two sequences (see Figure 8).

The hVR-1 nucleotide sequence (SEQ ID NO:1) was aligned with the rat VR-1 nucleotide sequence (Accession Number:AF029310) using the GAP program in the GCG software package (nwsgapdna matrix) and a gap weight of 50 and a length weight of 3. The results showed a 82.125% identity and 82.125% similarity between the two sequences (see Figure 9).

The hVR-1 protein (SEQ ID NO:2) was aligned with the rat VR-1 protein (Accession Number:AF029310) using the GAP program in the GCG software package (Blosum 62 matrix) and a gap weight of 12 and a length weight of 4. The results showed a 86.022% identity and 89.247% similarity between the two sequences (see Figure 10).

The hVR-2 protein (SEQ ID NO:5) was aligned with the human VR-2 protein (alternate form) (SEQ ID NO:8) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 11).

Finally, the hVR-2 protein (SEQ ID NO:5) was aligned with the predicted full length human VR-2 protein (alternate form) (SEQ ID NO:20) using the CLUSTAL W (1.74) multiple sequence alignment program (Figure 17).

A search was performed against the HMM database resulting in the identification of three ankyrin repeat domains in the amino acid sequence of hVR-1 (SEQ ID NO:2) at about residues 201-233, 248-283, and 333-361, and in the amino acid sequence of hVR-2 (SEQ ID NO:5) at about residues 162-194, 208-243, and 293-328. The results of the searches are set forth in Figures 13 and 15, respectively.

Hydropathy plots have identified 6 transmembrane domains in the hVR-1 and the hVR-2 proteins (see Figures 12 and 14, respectively).

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- 91 -

A series of searches have revealed that the hVR-1 protein matches the ProDom entry 141801 for the vanilloid receptor subtype and the ProDom entry 145518 for the vanilloid receptor subtype.

Moreover, a search was performed against the Prosite database resulting in the identification of four N-glycosylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 171-174, 192-195, 604-607, and 749-752), three cGMP-dependent protein kinase phosphorylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 2-5, 368-371, and 499-502), a series of protein kinase C and Casein kinase II phosphorylation sites in the amino acid sequence of SEQ ID NO:5, two tyrosine kinase phosphorylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 368-375 and 622-628), and two myristoylation sites in the amino acid sequence of SEQ ID NO:5 (at about residues 169-174 and 765-770).

Tissue Distribution of hVR-1 and hVR-2 mRNA

This Example describes the tissue distribution of hVR-1 and hVR-2 mRNA as determined by *in situ* hybridization.

For *in situ* analysis, tissues, such as brain regions and whole brain, obtained from human and monkey were first frozen on dry ice. Ten-micrometer-thick coronal sections of the tissues were postfixed with 4% formaldehyde in DEPC treated 1X phosphate-buffered saline at room temperature for 10 minutes before being rinsed twice in DEPC 1X phosphate-buffered saline and once in 0.1 M triethanolamine-HCl (pH 8.0). Following incubation in 0.25% acetic anhydride-0.1 M triethanolamine-HCl for 10 minutes, sections were rinsed in DEPC 2X SSC (1X SSC is 0.15M NaCl plus 0.015M sodium citrate). Tissue was then dehydrated through a series of ethanol washes, incubated in 100% chloroform for 5 minutes, and then rinsed in 100% ethanol for 1 minute and 95% ethanol for 1 minute and allowed to air dry.

Hybridizations were performed with ³⁵S-radiolabeled (5 X 10⁷ cpm/ml) cRNA probes. Probes were incubated in the presence of a solution containing 600 mM NaCl, 10 mM Tris (pH 7.5), 1 mM EDTA, 0.01% sheared salmon sperm DNA, 0.01% yeast tRNA, 0.05% yeast total RNA type X1, 1 X Denhardt's solution, 50% formamide, 10%

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- 92 -

dextran sulfate, 100 mM dithiothreitol, 0.1% sodium dodecyl sulfate (SDS), and 0.1% sodium thiosulfate for 18 hours at 55°C.

After hybridization, slides were washed with 2 X SSC. Sections were then sequentially incubated at 37°C in TNE (a solution containing 10 mM Tris-HCl (pH 7.6), 500 mM NaCl, and 1 mM EDTA), for 10 minutes, in TNE with 10µg of RNase A per ml for 30 minutes, and finally in TNE for 10 minutes. Slides were then rinsed with 2 X SSC at room temperature, washed with 2 X SSC at 50°C for 1 hour, washed with 0.2 X SSC at 55°C for 1 hour, and 0.2 X SSC at 60°C for 1 hour. Sections were then dehydrated rapidly through serial ethanol-0.3 M sodium acetate concentrations before being air dried and exposed to Kodak Biomax MR scientific imaging film for 24 hours and subsequently dipped in NB-2 photoemulsion and exposed at 4°C for 7 days before being developed and counter stained.

The data indicate that the hVR-1 molecule is not expressed in human nor monkey brain. The hVR-1 molecule is expressed in nodose, trigeminal sensory neurons, but is not expressed in sympathetic neurons. Within the nodose sensory neurons and trigeminal sensory neurons, expression was seen in distinct sub-populations. Moreover, hVR1 is expressed in some, but not all, small dorsal root ganglion (DRG) neurons and in a few medium sized DRG neurons. The hVR-1 molecule is partially co-expressed with the neuropeptide CGRP and with substance P which are present in nociceptive neurons.

The data further indicate that the VR-2 molecule is expressed in both human and monkey brain, primarily in cortical neurons. The VR2 molecule is also expressed in other brain regions, for example, the thalamus, striatum, hippocampus, hypothalamus, midbrain, medula and brain stem. In addition, the VR-2 molecule is expressed in parasympathetic neurons of the monkey heart (atrium), nodose sensory neurons, trigeminal (TRG) sensory neurons, dorsal root ganglion sensory neurons, sympathetic neurons, and motor neurons of the spinal cord. The VR2 molecule is widely expressed in TRG and DRG neurons, being present in most small and medium sized neurons and also in a few of the large neurons. VR2, like VR-1, partially co-localizes with CGRP and substance P.

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Trigeminal sensory neurons are recognized pain centers while sympathetic neurons have been shown to be involved in neuropathic pain.

EXAMPLE 2: EXPRESSION OF RECOMBINANT hVR-1, hVR-2, AND rVR-2 PROTEIN IN BACTERIAL CELLS

In this example, hVR-1, hVR-2, and rVR-2 is expressed as a recombinant glutathione-S-transferase (GST) fusion polypeptide in *E. coli* and the fusion polypeptide is isolated and characterized. Specifically, hVR-1, hVR-2, and rVR-2 is fused to GST and this fusion polypeptide is expressed in *E. coli*, *e.g.*, strain PEB199. Expression of the GST-hVR-1, GST-hVR-2, and GST-rVR-2 fusion protein in PEB199 is induced with IPTG. The recombinant fusion polypeptide is purified from crude bacterial lysates of the induced PEB199 strain by affinity chromatography on glutathione beads. Using polyacrylamide gel electrophoretic analysis of the polypeptide purified from the bacterial lysates, the molecular weight of the resultant fusion polypeptide is determined.

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EXAMPLE 3: EXPRESSION OF RECOMBINANT hVR-1, hVR-2, AND rVR-2 PROTEIN IN COS CELLS

To express the hVR-1, hVR-2, and rVR-2 gene in COS cells, the pcDNA/Amp vector from Invitrogen Corporation (San Diego, CA) is used. This vector contains an SV40 origin of replication, an ampicillin resistance gene, an *E. coli* replication origin, a CMV promoter followed by a polylinker region, and an SV40 intron and polyadenylation site. A DNA fragment encoding the entire hVR-1, hVR-2, and rVR-2 protein and an HA tag (Wilson *et al.* (1984) *Cell* 37:767) or a FLAG tag fused in-frame to its 3' end of the fragment is cloned into the polylinker region of the vector, thereby placing the expression of the recombinant protein under the control of the CMV promoter.

To construct the plasmid, the hVR-1, hVR-2, and rVR-2 DNA sequence is amplified by PCR using two primers. The 5' primer contains the restriction site of interest followed by approximately twenty nucleotides of the hVR-1, hVR-2, and rVR-2 coding sequence starting from the initiation codon; the 3' end sequence contains complementary sequences to the other restriction site of interest, a translation stop

- 94 -

codon, the HA tag or FLAG tag and the last 20 nucleotides of the hVR-1, hVR-2, and rVR-2 coding sequence. The PCR amplified fragment and the pCDNA/Amp vector are digested with the appropriate restriction enzymes and the vector is dephosphorylated using the CIAP enzyme (New England Biolabs, Beverly, MA). Preferably the two restriction sites chosen are different so that the hVR-1, hVR-2, and rVR-2 gene is inserted in the correct orientation. The ligation mixture is transformed into *E. coli* cells (strains HB101, DH5a, SURE, available from Stratagene Cloning Systems, La Jolla, CA, can be used), the transformed culture is plated on ampicillin media plates, and resistant colonies are selected. Plasmid DNA is isolated from transformants and examined by restriction analysis for the presence of the correct fragment.

COS cells are subsequently transfected with the hVR-1, hVR-2, and rVR-2pcDNA/Amp plasmid DNA using the calcium phosphate or calcium chloride coprecipitation methods, DEAE-dextran-mediated transfection, lipofection, or electroporation. Other suitable methods for transfecting host cells can be found in Sambrook, J., Fritsh, E. F., and Maniatis, T. Molecular Cloning: A Laboratory Manual. 15 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989. The expression of the hVR-1, hVR-2, and rVR-2 polypeptide is detected by radiolabelling (35S-methionine or 35S-cysteine available from NEN, Boston, MA, can be used) and immunoprecipitation (Harlow, E. and Lane, D. Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring 20 Harbor, NY, 1988) using an HA specific monoclonal antibody. Briefly, the cells are labelled for 8 hours with ³⁵S-methionine (or ³⁵S-cysteine). The culture media are then collected and the cells are lysed using detergents (RIPA buffer, 150 mM NaCl, 1% NP-40, 0.1% SDS, 0.5% DOC, 50 mM Tris, pH 7.5). Both the cell lysate and the culture media are precipitated with an HA specific monoclonal antibody. Precipitated 25 polypeptides are then analyzed by SDS-PAGE.

Alternatively, DNA containing the hVR-1, hVR-2, and rVR-2 coding sequence is cloned directly into the polylinker of the pCDNA/Amp vector using the appropriate restriction sites. The resulting plasmid is transfected into COS cells in the manner described above, and the expression of the hVR-1, hVR-2, and rVR-2 polypeptide is

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- 95 -

detected by radiolabelling and immunoprecipitation using an hVR-1, hVR-2, and rVR-2 specific monoclonal antibody.

EXAMPLE 4: ELECTROPHYSIOLOGICAL STUDIES OF VR2

Human VR2 was functionally characterized in both HEK293 cells and *Xenopus* oocytes using electrophysiological methods. VR2 (in the pcDNA3.1 vector purchased by Invitrogen) was transiently expressed in HEK293 cells (ATCC) and recordings were performed 48 hours after transfection of cells using the whole-cell patch-clamp method (described in Bertil Hille, Ionin Channels of excitable membranes, 1992; Hammill *et al.* (1981) *Pluger Arch.* 391:85-100). The results indicate that heat stimulation (>50 °C) induces a rapid inactivating inward current (1-2 nA). Heat-evoked currents of VR2 displayed profound desensitization and could be reversibly blocked by the VR1 inhibitor capsazepin (at a 10 μM concentration). In contrast to rat VR1, Capsaicin (at a 1-10 μM concentration), resiniferatoxin (at a 0.1-3 μM concentration), and low pH (5.0-6.0) do not induce any currents from VR2. Binding studies of [³H]-resiniferatoxin (NEN) to both human VR1 and VR2 in membranes isolated from HEK293 cell homogenates also indicate that resiniferatoxin (at a 0.1-10 nM concentration) has no specific binding to VR2 while it binds to human VR1 with high affinities.

For the oocyte studies, human VR2 was subcloned into an oocyte expression vector containing 5'- and 3'-UTR of *Xenopus* β-globin (Chiara *et al.* (1999) *Biochemistry* 38(20)6689-6698). *In vitro* transcription was carried out as described in Chiara *et al.* (*supra*) and cRNA (10-100 ng) was then injected into the oocytes. VR2 function was characterized in the oocytes 48 hours after cRNA injection using a standard two-electrode voltage-clamp. Consistent with the data from the HEK293 studies, VR2 can only be activated by heat stimulation (48-50 °C) but not by vanilloid receptor agonists, capsaicin, or resiniferatoxin. The vanilloid receptor antagonist capsazepine (at a 1-10 μM concentration) blocks the heat response of VR2 reversibly.

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EXAMPLE 5: GENERATION OF ANTI-hVR-2 ANTIBODIES AND hVR-2 PROTEIN LOCALIZATION BY IMMUNOSTAINING

Polyclonal antisera were raised in rabbits against the following three peptides derived from the human VR2 amino acid sequence, using the techniques described in Ed Harlow and David Lane (1988) "Antibodies; A Laboratory Manual" Cold Spring harbor Laboratory Press.

Antibody PEPTIDE 1: AFHCKSPHRHRMVVLE (SEQ ID NO:13)

Antibody PEPTIDE 2: RPEAPTGPNATESVQPMEGQEDEGN (SEQ ID NO:14)

Antibody PEPTIDE 3: SVLEMENGYWWCRKKQRAG (SEO ID NO:15)

10 Antiscra were subsequently affinity purified using the peptide immunogen.

The polyclonal antisera were tested for immunostaining of both monkey and rat dorsal root ganglion sensory neurons. Peptides 1 and 3 gave specific staining of subpopulations of sensory neurons that was competed with the corresponding peptide. This pattern of expression was very similar to the one observed using a VR-2 riboprobe.

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EXAMPLE 6: CHROMOSOMAL LOCALIZATION OF hVR-1 AND hVR-2

To chromosomally map the hVR-1 gene, primers were designed based on the sequence of hVR-1 (clone Fchrb87a6) (amplifying a 177 bp product from a human control cell line DNA and multiple faint larger products from a control Hamster cell line DNA by PCR). These primers were used to amplify 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL).

The hVR-1 primers used in the PCR mapping studies were: forward - TAGGAGACCCCGTTGCCACG (SEQ ID NO:16) and reverse -

- GATTCACTTGGGGACAGTGACG (SEQ ID NO:17) and the PCR reactions were performed as follows: 5 μl Template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR Buffer, 1.2μl Pharmacia dNTP mix 2.5 mM, 1.15μl Forward primer 6.6μM, 1.15μl Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq .05U/μl (Hot Start), using an amplification profile of: 95°C for 10 minutes followed by 35 Cycles of 94°C for 40
- 30 seconds, 55°C for 40 seconds, 72°C for 40 seconds, and 72°C for 5 minutes. The PCR

- 97 -

products were run on 2% agarose gels, post-stainedwith SYBR Gold (1:10,000 dilution in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

10 RH linkage analysis was performed using the Map Manager QTb28 software package.

hVR1 was found to map to the p arm of human chromosome 17, 18.9 cR₃₀₀₀ telomeric to the Whitehead Institute framework marker WI-6584, and 7.7 cR₃₀₀₀ centromeric of the Whitehead framework marker WI-5436. LOD scores for linkage were 14.5 for WI-6584 and 19.3 for WI-5436. This region corresponds to the cytogenetic location 17p12-13. This region is syntenic to mouse chromosome 11.

To chromosomally map the hVR-2 gene, primers were designed from 5' UTR sequence of human VR2 (clone Flh21e11) (amplifying a 166 bp product from a human control cell line DNA and 2 much larger faint bands from a control Hamster cell line DNA by PCR). These primers were used to amplify 93 DNAs in duplicate from the Genebridge 4 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL).

The hVR-2 primers used in the PCR mapping studies were: forward TTAAGCTCCCGTTCTCACCG (SEQ ID NO:18) and reverse GCTGCGGGAGGAAGTGAAGC (SEQ ID NO:19) and the PCR reactions were
performed as follows: 5μl Template DNA (10ng/μl), 1.5μl 10X Perkin Elmer PCR
Buffer, 1.2μl Pharmacia dNTP mix 2.5mM, 1.15μl Forward primer 6.6μM, 1.15μl
Reverse primer 6.6μM, 5μl Gibco/BRL Platinum Taq .05U/μl (Hot Start), using an
amplification profile of 95°C for 10 minutes, followed by 35 Cycles of 94°C for 40
seconds, 55°C for 40 seconds, 72°C for 40 seconds, and 72°C for 5 minutes. The PCR
products were run on 2% agarose gels, post-stainedwith SYBR Gold (1:10,000 dilution
in 1X TBE), and scanned on a Molecular Dynamics 595 Fluorimager.

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The following is the vector data for the 93 Genebridge4 hybrid DNAs. These are in order 1-93. A "1" is a positive result, a "-" is a negative result, a "?" is an ambiguous result.

RH linkage analysis was performed using the Map Manager QTb28 software package.

hVR2 was found to map to the p arm of human chromosome 17, 29.3cR cR₃₀₀₀ telomeric to the Whitehead Institute framework marker D17S721, and 23.3 cR₃₀₀₀ centromeric of the Whitehead framework marker AFMA043ZB5. LOD scores for linkage were 11.9 for D17S721 and 13.6 for AFMA043ZB5. This region corresponds to the cytogenetic location 17p11-12. This region is syntenic to mouse chromosome 11.

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Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

What is claimed:

- 1. An isolated nucleic acid molecule selected from the group consisting of:
- (a) a nucleic acid molecule comprising the nucleotide sequence set
- 5 forth in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof; and
 - (b) a nucleic acid molecule consisting of the nucleotide sequence set forth in SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 or a complement thereof.
- 2. An isolated nucleic acid molecule which encodes a polypeptide selected 10 from the group consisting of:
 - (a) a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11; and
 - (b) a polypeptide consisting of the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11.

- 3. An isolated nucleic acid molecule which encodes a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence set forth in SEQ ID NO:2, 5, 8, or 11.
- 20 4. An isolated nucleic acid molecule selected from the group consisting of:
 - a) a nucleic acid molecule comprising a nucleotide sequence which is at least 83% identical to the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof;
- b) a nucleic acid molecule comprising a fragment of at least 20 nucleotides of a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12, or a complement thereof;
 - c) a nucleic acid molecule which encodes a polypeptide comprising an amino acid sequence at least about 87% identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11; and

WO 00/29577

- 100 -

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d) a nucleic acid molecule which encodes a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15 contiguous amino acid residues of the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.

- 5. An isolated nucleic acid molecule comprising the nucleic acid molecule of any one of claims 1, 2, 3, or 4, and a nucleotide sequence encoding a heterologous polypeptide.
- 10 6. A vector comprising the nucleic acid molecule of any one of claims 1, 2, 3, or 4.
 - 7. The vector of claim 6, which is an expression vector.
- 15 8. A host cell transfected with the expression vector of claim 7.
 - 9. A method of expressing a polypeptide comprising culturing the host cell of claim 8 in an appropriate culture medium to, thereby, express the polypeptide.

- 10. An isolated polypeptide selected from the group consisting of:
- a) a fragment of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the fragment comprises at least 15 contiguous amino acids of SEQ ID NO:2, 5, 8, or 11;
- b) a naturally occurring allelic variant of a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11, wherein the polypeptide is encoded by a nucleic acid molecule which hybridizes to a nucleic acid molecule consisting of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12 under stringent conditions;
- c) a polypeptide which is encoded by a nucleic acid molecule comprising a nucleotide sequence which is at least 83% identical to a nucleic acid comprising the nucleotide sequence of SEQ ID NO:1, 3, 4, 6, 7, 9, 10, or 12; and
 - d) a polypeptide comprising an amino acid sequence which is at least 87% identical to the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.
- 15 11. The isolated polypeptide of claim 10 comprising the amino acid sequence of SEQ ID NO:2, 5, 8, or 11.
 - 12. The polypeptide of claim 10, further comprising heterologous amino acid sequences.

13. An antibody which selectively binds to a polypeptide of claim 10.

14. A method for detecting the presence of a polypeptide of claim 10 in a sample comprising:

- 25 a) contacting the sample with a compound which selectively binds to the polypeptide; and
 - b) determining whether the compound binds to the polypeptide in the sample to thereby detect the presence of a polypeptide of claim 10 in the sample.
- 30 15. The method of claim 14, wherein the compound which binds to the polypeptide is an antibody.

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- 16. A kit comprising a compound which selectively binds to a polypeptide of claim 10 and instructions for use.
- 5 17. A method for detecting the presence of a nucleic acid molecule of any one of claims 1, 2, 3, or 4 in a sample comprising:
 - a) contacting the sample with a nucleic acid probe or primer which selectively hybridizes to the nucleic acid molecule; and
- b) determining whether the nucleic acid probe or primer binds to a nucleic acid molecule in the sample to thereby detect the presence of a nucleic acid molecule of any one of claims 1, 2, 3, or 4 in the sample.
 - 18. The method of claim 17, wherein the sample comprises mRNA molecules and is contacted with a nucleic acid probe.
 - 19. A kit comprising a compound which selectively hybridizes to a nucleic acid molecule of any one of claims 1, 2, 3, or 4 and instructions for use.
- 20. A method for identifying a compound which binds to a polypeptide of claim 10 comprising:
 - a) contacting the polypeptide, or a cell expressing the polypeptide with a test compound; and
 - b) determining whether the polypeptide binds to the test compound.
- 25 21. The method of claim 20, wherein the binding of the test compound to the polypeptide is detected by a method selected from the group consisting of:
 - a) detection of binding by direct detection of test compound/polypeptide binding;
 - b) detection of binding using a competition binding assay; and
- 30 c) detection of binding using an assay for hVR-1, hVR-2, or rVR-2 activity.

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22. A method for modulating the activity of a polypeptide of claim 10 comprising contacting the polypeptide or a cell expressing the polypeptide with a compound which binds to the polypeptide in a sufficient concentration to modulate the activity of the polypeptide.

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- 23. A method for identifying a compound which modulates the activity of a polypeptide of claim 10 comprising:
 - a) contacting a polypeptide of claim 10 with a test compound; and
- b) determining the effect of the test compound on the activity of the
 polypeptide to thereby identify a compound which modulates the activity of the polypeptide.
 - 24. A method for treating a subject having a disorder characterized by aberrant hVR-1 or hVR-2 protein activity or nucleic acid expression comprising administering to the subject a hVR-1 or hVR-2 modulator such that treatment of the subject occurs.
 - 25. The method of claim 24, wherein the hVR-1 or hVR-2 modulator is a small molecule.

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26. The method of claim 24, wherein the disorder is a pain disorder.

1/35

humanVR1 gene with translation of open reading frame

Input file Fchrb87a6.seq; Output File Fchrb87a6.tra Sequence length 3909

GTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAA GCAGTGAGCGCAACGCACTTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCCCAACGCACTGCGGGCAGTGAGC GCAACGCACTGCGGGCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCAACGCACT GCGGGCAGTGAGCGCACCGCGCCAGTGAGCGCAACGCACTGCGGGCAGTGAGCGCACCGCACCGCACTGCGGGCAGTG TTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGTGAGCGGATAACAATTTCACACAGGAAACAGCTATGACCATGATT ACGCCAAGCTCTAATACGACTCACTATAGGGAAAGCTGGTACGCCTGCAGGTACCGGTCCGGAATTCCCGGGTCGACCC ACGCGTCCGAAAACACACCTCTCTGCTGGGAAGACTGTGCAATGGCACAGCCGCAGAGCTTGGTTTGGGAGGTTGAA GTGCTCTGGGGAGAATTCGTAGATCATCCTCAGAAAAGCCTTGCCCTGGTGTTCTACCAGAAAAACGTCTCCCAATCAC CCAGAAAAGCTGTCCACAGTAGTCCCCCCTTATCCACGGGTGTCACTTTCCATGGGTTCAGTTATTTGCGGTCAACCAC **GGTCTGCCAATATTAAATGGAAAATTCTTCAAACAGTTCCCAAGTTTTCCCTTGTGCATTGTTCTGAGCAGTGTGATGA** AGAGTCTCTGCCGTGCCATCTGGGATGCAAACCGTCCCTGTGTCCCCACGTCCAGGCCGTAGATGCTCCCCGCCGGTC **AGTCACTTAGTCGTCAGATCGCCCGTCCTGGTATCACAGTGCTTCTGTTCAGGTTGCACACTGGGCCACAGAGGATCCA** w s s T D L G T AADP 18 GCAAGG ATG AAG AAA TGG AGC AGC ACA GAC TTG GGG ACA GCT GCG GAC CCA CTC CAA AAG 54 DTCP D P L D G D P N S R P P PAKP 38 GAC ACC TGC CCA GAC CCC CTG GAT GGA GAC CCT AAC TCC AGG CCA CCT CCA GCC AAG CCC Q L P T A K S R T R L F G K G D S R B A 58 CAG CTC CCC ACG GCC AAG AGC CGC ACC CGG CTC TTT GGG AAG GGT GAC TCG GAG GAG 174 PPVD C P H B B G B L D S C 78 P TTC CCG GTG GAT TGC CCC CAC GAG GAA GGT GAG TTG GAC TCC TGC CCG ACC ATC ACA GTC 234 v ITIQR P G D G P TGAR 98 LLS AGC CCT GTT ATC ACC ATC CAG AGG CCA GGA GAC GGC CCC ACC GGT GCC AGG CTG CTG TCC 294 Q D S V A A S T B K T L R L Y D R R S I 118 CAG GAC TCT GTC GCC GCC AGC ACC GAG AAG ACC CTC AGG CTC TAT GAT CGC AGG AGT ATC Q N N CQDL E 8 L 138 LL

FIGURE 1

TIT GAA GCC GIT GCT CAG AAT AAC TGC CAG GAT CTG GAG AGC CTG CTG CTC TTC CTG CAG

F L

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K D LTDNEF 158 AAG AGC AAG AAG CAC CTC ACA GAC AAC GAG TTC AAA GAC CCT GAG ACA GGG AAG ACC TGT 474 L N L H D G Q N 178 CTG CTG AAA GCC ATG CTC AAC CTG CAC GAC GGA CAG AAC ACC ACC ATC CCC CTG CTC CTG 534 Q T v R D S LKE L N A 198 GAC ATC GCG CGG CAA ACG GAC AGC CTG AAG GAG CTT GTC AAC GCC AGC TAC ACG GAC AGC 594 R R N GOT ALHIAIE 218 TAC TAC AAG GGC CAG ACA GCA CTG CAC ATC GCC ATC GAG AGA CGC AAC ATG GCC CTG GTG 654 H ν A OA A E N G A D 238 ACC CTC CTG GTG GAG AAC GGA GCA GAC GTC CAG GCT GCG GCC CAT GGG GAC TTC TTT AAG P G FYFGELP LSLAA 258 AAA ACC AAA GGG CGG CCT GGA TTC TAC TTC GGT GAA CTG CCC CTG TCC CTG GCC GCG TGC 774 F L L Q N s L G I K 278 ACC AAC CAG CTG GGC ATC GTG AAG TTC CTG CTG CAG AAC TCC TGG CAG ACG GCC GAC ATC 834 RDSVGNTVLHAL v E V 298 AGC GCC AGG GAC TCG GTG GGC AAC ACG GTG CTG CAC GCC CTG GTG GAG GTG GCC GAC AAC 894 T S M Y N E I L M L G A 318 F V ACG GCC GAC AAC ACG AAG TTT GTG ACG AGC ATG TAC AAT GAG ATT CTG ATG CTG GGG GCC n k k G м 338 E E L T L ĸ L AAA CTG CAC CCG ACG CTG AAG CTG GAG GAG CTC ACC AAC AAG AAG GGA ATG ACG CCG CTG 1014 TGKIGVLAYILQREI 358 A A G GCT CTG GCA GCT GGG ACC GGG AAG ATC GGG GTC TTG GCC TAT ATT CTC CAG CGG GAG ATC 1074 RKFTEWA 378 YGPV С R H L S CAG GAG CCC GAG TGC AGG CAC CTG TCC AGG AAG TTC ACC GAG TGG GCC TAC GGG CCC GTG 398 CIDTCE S S ĸ L Y D CAC TCC TCG CTG TAC GAC CTG TCC TGC ATC GAC ACC TGC GAG AAG AAC TCG GTG CTG GAG 1194 418 YSSSRTPNRH M L L A D GTG ATC GCC TAC AGC AGC AGC GAG ACC CCT AAT CGC CAC GAC ATG CTC TTG GTG GAG CCG 1254 438 R L L Q D K W D R F ĸ R I P CTG AAC CGA CTC CTG CAG GAC AAG TGG GAC AGA TTC GTC AAG CGC ATC TTC TAC TTC AAC 1314 458 A F L V Y C L Y M I I T TTC CTG GTC-TAC TGC CTG TAC ATG ATC ATC TTC ACC ATG GCT GCC TAC TAC AGG CCC GTG 478 KIGD Y F R V T G P P F K M E L GAT GGC TTG CCT CCC TTT AAG ATG GAA AAA ATT GGA GAC TAT TTC CGA GTT ACT GGA GAG 498 G GVYFFFR I V L G ATC CTG TCT GTG TTA GGA GGA GTC TAC TTC TTT TTC CGA GGG ATT CAG TAT TTC CTG CAG 1494

R AGG	R CGG	P	S TCG	M GTA	K AAG	T ACC	L CTG	F TTT	V GTG	D GAC	S	Y TAC	S AGT	E GAG	M ATG	L CTI	F	F TTT	L CTG	518 1554
Q	S	L	F	M	L	A	T	V	V	L	Y	F	S	H	L	K	E	Y	v	538
CAG	TCA	CTG		ATG	CTG	GCC	ACC	GTG	GTG	CTG	TAC	TTC	AGC	CAC	CTC	AAG	GAG	TAT	GTG	1614
A	S	M	V	F	S	L	A	L	G	w	T	N	M	L	Y	Y	T	R	G	558
GCT	TCC	ATG	GTA	TTC	TCC	CTG	GCC	TTG	GGC	Tgg	ACC	AAC	ATG		TAC	TAC	ACC	CGC	GGT	1674
F	Q	Q	M	G	I	Y	A	V	M	I	e	K	M	I	L	R	D	L	C	578
TTC	CAG	CAG	ATG	GGC	ATC	TAT	GCC	GTC	ATG	ATA	gag	AAG	ATG	ATC	CTG	AGA	GAC	CTG	TGC	1734
R	F	m	F	V	Y	I	V	F	L	F	G	F	S	T	A	V	V	T	L	598
CGT	TTC	atg	TTT	GTC	TAC	ATC	GTC	TTC	TTG	TTC	GGG	TTT	TCC	ACA	GCG	GTG	GTG	ACG	CTG	1794
I	E	D	G	K	N	D	s	L	P	S	E	S	T	S	H	R	w	R	G	618
TTA	GAA	GAC	GGG	AAG	TAA	GAC	TCC	CTG	CCG	TCT	GAG	TCC	ACG	TCG	CAC	AGG	TGG	CGG	GGG	1854
P	A	C	R	P	P	D	S	S	Y	N	S	L	Y	S	T	C	L	E	L	638
CCT	GCC	TGC	AGG		CCC	GAT	AGC	TCC	TAC	AAC	AGC	CTG	TAC	TCC	ACC	TGC	CTG	GAG	CTG	1914
F	K	F	T	I	G	M	G	D	L	E	F	T	E	n	Y	D	F	K	A	658
TTC	AAG	TTC	ACC	ATC	GGC	ATG	GGC	GAC	CTG	GAG	TTC	ACT	GAG	aac	TAT	GAC	TTC	Aag	GCT	1974
V	F	I	I	L	L	L	A	Y	V	I	L	T	Y	I	L	L	L	N	m	678
GTC	TTC	ATC	ATC	CTG	CTG	CTG	GCC	TAT	GTA	ATT	CTC	ACC		ATC	CTC	CTG	CTC	AAC	atg	2034
L	I	A	L	M	G	E	T	V	N	K	I	A	Q	E	S	K	N	I	W	698
CTC	ATC	GCC	CTC	ATG	GGT	GAG	ACT	GTC	AAC	AAG	ATC	GCA	CAG	GAG	AGC	AAG	AAC	ATC	TGG	2094
k	L	Q	R	A	I	T	I	L	D	T	E	K	S	F	L	K	C	M	r	718
Aag	CTG	CAG	AGA	GCC	ATC	ACC	ATC	CTG	GAC	ACG	GAG	AAG	AGC	TTC	CTT	AAG	TGC	ATG	Agg	2154
k	A	F	R	S	G	K	L	L	Q	V	G	Y	T	P	D	G	K	D	D	738
aag	GCC	TTC	CGC	TCA	GGC	AAG	CTG	CTG	CAG	GTG	GGG	TAC	ACA		GAT	GGC	AAG	GAC	GAC	2214
Y	R	W	C	F	R	V	D	E	V	N	w	T	T	w	N	T	n	GIG	G	758
TAC	CGG	TGG	TGC	TTC	AGG	GTG	GAC	GAG	GTG	AAC	TGG	ACC	ACC	TGG	AAC	ACC	aac	V	GGC	2274
I ATC	I ATC	N	e gaa	D GAC	P CCG	GGC GGC	N AAC	C TGT		G GGC	V GTC	K Aag	R CGC	T ACC	L CTG	S AGC	F TTC	s TCC	L CTG	778 2334
R CGG	S TCA	s agc	R AGA	V GTT	S TCA	G GGC	R AGA	H CAC	W TGG	K AAG	N AÀC	F TTT	A GCC	CTG	V GTC	CCC	L CTT	L TTA	R AGA	798 2394
e	a	s	A	R	D	r	Q	S	A	Q	CCC	E	E	V	Y	L	R	Q	F	818
Gag	GCA	Agt	GCT	CGA	GAT	agg	CAG	TCT	GCT	CAG		GAG	GAA	GTT	TAT	CTG	CGA	CAG	TTT	2454
s	G	e	L	K	P	e	D	A	E	V	F	r	e	CCL	A	A	s	G	e	838
TCA	GGG	TCT	CTG	AAG	CCA	Gag	GAC	GCT	GAG	GTC	TTC	aag	agt		GCC	GCT	TCC	GGG	gag	2514
K AAG	* TGA																			840 2520

GGACGTCACGCAGACAGCACTGTCAACACTGGGCCTTAGGAGACCCCGTTGCCACGGGGGGCTGCTGAGGGAACACCAG

TGCTCTGTCAGCAGCCTGGCCTGGTCTGTGCCTGCCCAGCATGTTCCCAAATCTGTGTGGACAAGCTGTGGGAAGCGT

TCTTGGAAGCATGGGGAGTGATGTACATCCAACCGTCACTGTCCCCCAAGTGAATCTCCTAACAGACTTTCAGGTTTTTA

CTCACTTTACTAAAAAAAAAAAAAAAAAAAGGGCGGCCGCTTA

4/35

228

684

248

5/35

Full-length human VR2

Input file Flh21e11.seq; Output File Flh21e11.tra Sequence length 2809

OGCTAGCCTGTCCTGACAGGGGAGAGTTAAGCTCCCGTTCTCCACCGTGCCGGCTGGCCAGGTGGGCTGAGGGTGACCG GGCAGCCCCTCCCGGCTTCACTTCCTCCCGCAGCCCCTGCTACTGAGAAGCTCCCGGGATCCCAGCAGCCGCCACGCCCT GGCTCAGCCTGCGGGGCTCCAGTCAGGCCAACACCGACGCGCAGCTGGGAGGAAGACAGGACCCTTGACATCTCCATC S P TECACAGAGGTCCTGGCTGGACCGAGCAGCCTCCTCCTAGG ATG ACC TCA CCC TCC AGC TCT CCA 24 E D G E ETLD G G Q 28 OTT TTC AGG TTG GAG ACA TTA GAT GGA GGC CAA GAA GAT GGC TCT GAG GCG GAC AGA GGA ₽ M E 0 G G P 48 AME CTG GAT TIT GGG AGC GGG CTG CCT CCC ATG GAG TCA CAG TTC CAG GGC GAG GAC CGG 144 I R V N L N Y R K G 68 AAA TTC GCC CCT CAG ATA AGA GTC AAC CTC AAC TAC CGA AAG GGA ACA GGT GCC AGT CAG 204 PNRFDRDRL F N 88 COG GAT CCA AAC CGA TIT GAC CGA GAT CGG CTC TTC AAT GOG GTC TCC CGG GGT GTC CCC K Y E L 108 CAG GAT CTG GCT GGA CTT CCA GAG TAC CTG AGC AAG ACC AGC AAG TAC CTC ACC GAC TCG 324 B Y T B G S T G K T C L M ĸ A 128 GAN THE ACA GAG GGC TEE ACA GGT AAG ACG TGE ETG ATG AAG GET GTG ETG AAC ETT AAG C I Q I D R D G 148 P L L N A L CAC GGA GTC AAT GCC TGC ATT CTG CCA CTG CTG CAG ATC GAC AGG GAC TCT GGC AAT CCT 444 G H Ð D Y Y 16B T CAG CCC CTG GTA AAT GCC CAG TGC ACA GAT GAC TAT TAC CGA GGC CAC AGC GCT CTG CAC CVKL ĸ KRSL Q ATC GCC ATT GAG AAG AGG AGT CTG CAG TGT GTG AAG CTC CTG GTG GAG AAT GGG GCC AAT 564 K G Q G T C 208 0 GTG CAT GCC COG GCC TGC GGC CGC TTC TTC CAG ANG GGC CAA GGG ACT TGC TTT TAT TTC 624

L H A L V N I S D N S A E N I A L V T S 268
CTG CAT GCC CTA GTG ATG ATC TCG GAC AAC TCA GCT GAG AAC ATT GCA CTG GTG ACC AGC 804

C T 'K

GOT GAG CTA COC CTC TCT TTG GCC GCT TGC ACC AAG CAG TGG GAT GTG GTA AGC TAC CTC

CTO GAG AAC COA CAC CAG CCC GCC AGC CTG CAG GCC ACT GAC TCC CAG GGC AAC ACA GTC

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IRRLQDLTPLKLAAKEGKIE 308

6/35

ATC COC AAC CTG CAG GAT CTC ACG CCT CTG AAG CTG GCC GCC AAG GAG GGC AAG ATC GAG E F S G L S H L S R K F 328 L Q R ATT TTC AGG CAC ATC CTG CAG CGG GAG TTT TCA GGA CTG AGC CAC CTT TCC CGA AAG TTC L Y D R .V L A P V 8 TEWCYG ACC GAG TOG TGC TAT GGG CCT GTC CGG GTG TCG CTG TAT GAC CTG GCT TCT GTG GAC AGC 1044 FHCKSP E I I A S L TGT GAG GAG AAC TCA GTG CTG GAG ATC ATT GCC TTT CAT TGC AAG AGC CCG CAC CGA CAC 1104 K L L Q E L N CGA ATG GTC GTT TTG GAG CCC CTG AAC AAA CTG CTG CAG GCG AAA TGG GAT CTG CTC ATC 1164 FLNFLCNLIYMFIF CCC AAG TTC TTC TTA AAC TTC CTG TGT AAT CTG ATC TAC ATG TTC ATC TTC ACC GCT GTT 1224 P н K E 428 TLKKO GCC TAC CAT CAG CCT ACC CTG AAG AAG CAG GCC GCC CCT CAC CTG AAA GCG GAG GTT GGA 1284 G G 1 N S M L L T G H I L I L L AAC TCC ATG CTG CTG ACG GGC CAC ATC CTT ATC CTG CTA GGG GGG ATC TAC CTC CTC GTG 1344 I W I S F I D S 468 R H V F Y F W R GGC CAG CTG TGG TAC TTC TGG CGG CGC CAC GTG TTC ATC TGG ATC TCG TTC ATA GAC AGC 1404 ALLTVVSQV YFEIL F 0 TAC TIT GAA ATC CTC TTC CTG TTC CAG GCC CTG CTC ACA GTG GTG TCC CAG GTG CTG TGT 1464 F L Y L P L L V S A L V L 508 TTC CTG GCC ATC GAG TGG TAC CTG CCC CTG CTT GTG TCT GCG CTG GTG CTG GGC TGG CTG 1524 YTRGFQHTG S M I Y ARC CTG CTT TAC TAT ACA CGT GGC TTC CAG CAC ACA GGC ATC TAC AGT GTC ATG ATC CAG 1584 L V KVILRDLLRFLL 1 ANG GTC ATC CTG CGG GAC CTG CTG CGC TTC CTT CTG ATC TAC TTA GTC TTC CTT TTC GGC 1644 RPEA A L V S L S Q E A TTC GCT GTA GCC CTG GTG AGC CTG AGC CAG GAG GCT TGG CGC CCC GAA GCT CCT ACA GGC 1704 G A 588 E B G Q N A T E S V M CCC AAT GCC ACA GAG TCA GTG CAG CCC ATG GAG GGA CAG GAG GAC GAG GGC AAC GGG GCC 1764 P BASLBLFKFT-IG-HG CAG TAC AGG GOT ATC CTG GAA GOC TOC TTG GAG CTC TTC AAA TTC ACC ATC GGC ATG GGC 1824 QYRG I L L. A 628 BQLEFRGHVL L L L GAG CTG GCC TTC CAG GAG CAG CTG CAC TTC CGC GGC ATG GTG CTG CTG CTG CTG GCC 1884 L L T T I L L R N L ALMSET I THE GTG CTG CTC ACC THE ATC CTG CTG CTC ARE ATG CTC ATC GCC CTC ATG AGC GAG ACC 1944 W K L Q K A I S V I TDSW B GTC AAC AGT GTC GCC ACT GAC AGC TOG AGC ATC TOG AAG CTG CAG AAA GCC ATC TCT GTC 2004 C R K K Q 688 R Ħ w CTG GAG ATG GAG AAT GGC TAT TOG TGG TGC AGG AAG AAG CAG CGG GCA GGT GTG ATG CTG 2064 RNENG ¥ K P D G S P D E R W C F R V E E 708

FIGURE 2 (continued)

2124

ACC GIT GGC ACT AAG CCA GAI GGC AGC CCG GAI GAG CGC TGG TGC TIC AGG GTG GAG GAG

7/35

FIGURE 2 (continued)

728 2184 748 . 765 2295 G V P R T L E N P V L A S P P K E D B D GOT OFF CCT CCC ANG GAT GAG GAT V N W A S W E Q T L P T L C E D P S G A GTG AC TOT ACG CTG TGT GAG GAC CCG TCA GGG GCA G A S E E N Y V P V Q L L Q S N * GST GCC TCT CAG TCC AAC TGA

8/35

Partial human VR2 alternate form

Input file frhob12c4.seq; Output File frhob12c4.tra Sequence length 1489

G		c o	R GC	F	F TT	Q C CA	K G AA	G G GG	Q CAJ	G 4. GGG	T G ACT	C TG(F TT	Y AT T		G C GG) P	C CI	19 C 57
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L	C	•	A	G	A	R	L	C	P	T	v	Q	L	8	D	I	R		L	. 0	99
CT	CA	A G	CT	GGG	GC	c cc	CT	C TG	C C C	CAC	GT	CA	CT	T GA	G GA	C AT	c cc	CAA	c cr	G CA	3 297
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D	L	. •	T	P	L	K	L	A	A	K	. E	G	K	I	E	I	F	R	н	1	119
GA?	г ст	CA	œ	CCI	CIC	S AA	CIN	GCC	GCC	: AAC	GAC	GGG	C AAC	TA E	C GA	3 AT	r TT	C AG	G CA	C AT	
L	Q	1	R	E	F	S	G	L	s	H	L	S	R	ĸ	P	T	E	W	C	Y	139
CIG	CA	Gα	G	GAG	TT	TCI	A GGZ	CTG	AGC	CAC	CIT	TCC	: CG1	AA A	TT	: AC	C GA	3 TG	G TG	C TAT	417
	P					_		_	D								_	E		s	159
GGG	CC	r G1	~	CGG	GTG	TO	CIC	TAT	GAC	CIG	GCI	TCI	GIG	GAG	: AGC	TG	GAC	GA(AA E	TCA	477
-	L	_	-	_	_		-		C		_				_		M	v	•	L	179
GTG	CIX	G GA	G	ATC	ATI	. ecc	777	CAT	TGC	AAG	AGC	CCC	CAC	: CGA	CAC	CCF	ATC	GIY	GT	TIG	537
									. A									F	F	L	199
GAG	ccc	CI	G.	AAC	AAA	CIG	CIG	CAG	GCG	AAA	TGG	GAT	CIG	CTC	ATC	ccc	: AAG	TIC	110	TTA	597
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N	_	_		С	N	_			M				T				Y	H	Q	P	219
AAC	110	: CI	G	TGT	AAT	CTG	ATC	TAC	ATG	TTC	ATC	TTC	ACC	GCI	GIT	GCC	TAC	CAT	CAG	CCI	657
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ACC	CIG	, AA	G.	AAG	CAG	တေ	GCC	CCT	CAC	CTG	AAA	GCG	GAG	GIT	GGA	AAC	TCC	ATC	CIG	CTG	717
	_			_	_	_		_	_	_	_		_	_		_	_	_			
T	G	_		I		I			G			Y		L	V	G	.O	L	W	Y	259
ACG	GGC	CA	C A	ATC	CIT	ATC	CIG	CTA	GGG	GGG	ATC	TAC	CIC	CIC	GIG	GGC	CAG	CIG	TGG	TAC	777
		_	_	_	_		_	_		_	_	_	-	_	_	_	_	_	_	_	
_	H			R					. W					D	8	_	F	B	_		- 279
110	100	C.	G (CGC.	CAC	GIG	HC	MC	TGG	ATC	100	110	ALA	GAC	AGC.	TAC	111	GAA	ATC	CIC	837
	T.			_	_			_	v.	•		_	17	L	_		•.		•		299
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			-																		

FIGURE 3

379

V E GTG QAG

C F R TGC TTC AGG G 399

s g TCA GGG **419** 1257

A G V P R T L E N P V L A S P P K B D E GCA GGT GTC CCT CCC AAG GAG GAT GAG

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L T V G T K P D G S P D E R W CTG ACC GTT GGC ACT CAG CCA GAT GGC AGC CCG GAT GAG CGC TGG

TGGCCCAAATGCAGCAGAAGGCAAAGAGCAGAGCAGAGGATCTTTCCAACCACATCTGCTGGCTCTGGGGTCCCAGTG

D G A S E E N Y V P V Q L L Q S N • GAT GGT GCC TC CAG CTC CAG TCC AAC TGA

GCGGACGCGTGGGTCGAC

437

9/35

FIGURE 3 (continued)

Partial rat VR2

Input file Plrxb147g11.seq; Output File Flrxb147g11.tra Sequence length 1794

500	gue		9 -		-															
G 1	s rcc /	T ACC (H CAC	A GCG	s rcc o	A GCT (L CTT :	s rcr (r CTG (A CT (A GCG 1	C TGC	T ACC	K AAG	Q CAG	W TGG	D GAT	V GTG	V GTG	19 57
~	v	7.	1.	E	N	Ð	u	Q	Ð	A	s	L	E	A	T	D	s	L	G	39
ACC	TAC	. cr	c c T	GĀ	AAC	င်္	CAC	CÃ	ccc	s GCC	AGG	CT	GAG	G GC					GGC	117
N	т	v	L	н	A	L	v	м	I	A	D	N	s		E		s	A	L	59
AAC	ACA	GTC	CIC	CAT	GCT	CTG	GTA	ATC	ATT	GCA	GA1	AA 7	TCC	s cc	r gac	AA E	AG1	r GCC	CIG	177
v	I	H	M	Y	D	G	Ĺ	L	Q	M	G	A				P		v		79
GTG	ATC	CAC	ATG	TAC	GAC	GGG	CIT	CTA	CAA	ATG	GGG	GCG	CGC	CIV	1G0	ccc	: ACI	GTG	CAG	237
L		E	Ŧ	s	N	н	0	G	L	T	₽	L	K	L	A	A	ĸ	E	G	99
CTT	GAG	GAA	ATC	TCC	AAC	CAC	CAA	GGC	CTC	ACA	ccc	CIG	AAA	СТА	GCC	GCC	AAG	GAA	GGC	297
ĸ	I	E	I	F	R	H	I	L	Q	R	E	F	s	G	P	Y	Q	P	L	119
AAA	ATC	GAG	ATT	TTC	AGG	CAC	ATT	CTG	CAG	CGG	GAA	TTC	TCA	GGA	CCG	TAC	CAG	ccc	CTT	357
s	R	ĸ	P	T	E	w	C	Y	G	P	v	R	v	s	L	Y	D	L	s	139
TCC	CGA	AAG	TIT	ACT	GAG	TGG	TGT	TAC	GGT	CCT	GTG	CGG	GTA	TCG	CIG	TAC	GAC	CTG	TCC	417
s	v	D	s	W	E	ĸ	· N	s	v			I		A	F		C	K	s	159
TCT	GTG	GAC	AGC	TGG	GAA	AAG	AAC	TCG	GTG	CTG	GAG	ATC	ATC	GCT	TTT	CAT	TGC	AAG	AGC	477
P	N	R	H	R	M	V	v	L	E	P		N		L		Q	E	K	W	179
ccc	AAC	CGG	CAC	œc	ATG	GTG	GTT	TTA	GAA	CCA	CIG	ŸAC	AAG	CIT	CIG	CAG	GAG	AAA	TGG	537
D	R ·	I.	v	s	R	F	F	F	N	F	A			L	v		M	F	I	199
GAT	œc	CTC	CIC	TCA	AGA	TTC	TTC	TTC	AAC	TTC	GCC	TGC	TAC	TTG	GTC	TAC	ATG	TIC	ATC	597
P	T	v	v	A	Y	Ħ	Q	P	s	L	D	Q	P	A	I	P	s	S	K	219
TTC	ACC	GIC	GTT	GCC	TAC	CAC	CAG	ccr	TCC	CTG	GAT	CAG	CCA	GCC	ATC	ccc	TCA	TCA	AAA	657
A	T	F	G	E	s	м	L	L	L	G	Ħ	I		I	L	L	G	G	I	239
ecci	ACT	TIT	GGG	GAA	TCC	ATG	CTG	CTG	CTG	GGC	CAC	ATT	CIG	ATC	CTG	CTT	GGG	GGT	ATT	717
¥	L	t.	L	G	0	L	W	Y	F	W	R	R	R	L	F	I	W	1	S	259
TAC	crc	TTA	CTG	GGC	CAG	CIG	TGG	TAC	TTT	TGG	CGG	CGG	СЗС	CTG	TTT	ATC	TGG	ATC	TCA	777
P		ъ	s-	. 🕶	· p	- E	I	L	F	L	L	Q	A	L	L	T	v	L	8	279
TTC	ATG	GAC	AGC	TAC	111	GAA	ATC	crc	TIT	ĊIC	CIT	CAG	GCT	CTG	CTC	ACA	GTG	CIG	TCC	837
Q	v	t.	R	F	H	R	T	B	Ħ	Y	L	P	L	L	V	L	8	L	V	299
c <u>w</u>	GIG	CL0	œc	TTC	ATG	GAG.	ACT	CAA	TGG	TAC	CIA	œc	CTG	CTA	GTG	TTA.	TCC	CEA	GIG	897
L	G	w	L	N	L	L	Y	T	T	R	G	F	Q	Ħ	T	G	I	Y	8	319
CTG	GGC	TOG	c <u>i</u> a	AAC	CTG	CIT	TAC	TAC	ACA	CGG	GGC	TIT	CAG	CAC	ACA	GGC	ATC	TAC	agt	957
v	M	I	Q	K	v	I	L	R	D	L	L	R	F	L	L	v	Y	L	v	339
orc	ATG	ATC	œ	ANG	arc	ATC	CIT	AEDO	GAC	CTG	CIC	CCT	TTC	CIG	CTG	GTC	TAC	CIG	GIC	1017
F	L	Ŧ	a	F	A	V	A	L	V	. 6	L	6	R	B	A	R	8	P	K.	359
110	CIT	TTC	GGC	111	GCT	GTA	GCC	CIA	GTA	AGC	TTG	AGC	AGA	GAG	GCC	CGA	agt	ccc	AAA	1077

379 1137	399 1197	419	439 1317	459	479	499	519 1557	539 1617	555 1665	
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FIGURE 4 (contimined)

12/35

GAP of: humanvr2.pep check: 5746 from: 1 to: 764 humanVR2 Flh21e11 to: humanvrl.pep check: 6877 from: 1 to: 839 humanVR1 _Fbh18547pat - fchrb87a6, 3909 bases, 4554 checksum. Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430 12 Average Match: 2.912 Gap Weight: 4 Average Mismatch: -2.003 Length Weight: 850 Length: Quality: 1530 Gaps: 10 Ratio: 2.003 Percent Similarity: 55.378 Percent Identity: 46.348 Match display thresholds for the alignment(s): | = IDENTITY2 humanvr2.pep x humanvr1.pepMTSPSSSPVF 10 1 MKKWSSTDLGTAADPLQKDTCPDPLDGDPNSRPPPAKPQLPTAKSRTRLF 50 11 RLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY 60 : 1.11 : . 51 GKGDSEEAFPVDCPHEEGELDSCPTI.TVSPVITIQRPGDGPTGARLLSQ 99 61 RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEY 110 100 DSVAASTEKTLRLYDRRSIFEAVAQNNCQDLESLLLFLQKSKKHLTDNEF 149 111 TEGSTGKTCLMKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDY 160 150 KDPETGKTCLLKAMLNLHDGQNTTIPLLLEIARQTDSLKELVNASYTDSY 199 161 YRGHSALHIAIEKRSLQCVKLLVENGANVHARACGRFFQKGQG.TCFYFG 209 1:1 -1:1111:1:: 1 1111111.1 1 1 1 11.1 .1 1111 200 YKGQTALHIAIERRNMALVTLLVENGADVQAAAHGDFFKKTKGRPGFYFG 249 210 ELPLSLAACTKQWDVVSYLLENPHQPASLQATDSQGNTVLHALVMISDNS 259 250 ELPLSLAACTNQLGIVKFLLQNSWQTADISARDSVGNTVLHALVEVADNT 299 260 AENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKLAAKEGKIEI 309 300 ADNTKFVTSMYNEILMLGAKLHPTLKLEELTNKKGMTPLALAAGTGKIGV 349

310	FRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEI	357
	:	399
406	TAVAYHOPTLKKQAAPHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFW	455
450	TMAAYYRPVDGLPPFKMEKIGDYFRVTGEILSVLGGVYFFFRGIQYFL	497
456	RRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEWYLPLLVSALVL	505
498	QRRPSMKTLFVDSYSEMLFFLQSLFMLATVVLYFSHLKEYVASMVFSLAL	547
506	GWLNLLYYTRGFOHTGIYSVMIQKVILRDLLRFLLIYLVFLFGFAVALVS	555
548	GWTNMLYYTRGFQQMGIYAVMIEKMILRDLCRFMFVYIVFLFGFSTAVVT	5 9 7
556	LSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLELFKFTI	605
598	LIEDGKNDSLPSESTSHRWRGPACRPPDSSYNSLYSTCLELFKFTI	643
606	GMGELAFOEQLHFRGMVLLLLAYVLLTYILLLNMLIALMSETVNSVATD	655
644	GMGDLEFTENYDFKAVFIILLLAYVILTYILLLNMLIALMGETVNKIAQE	693
656	SWSIWKLOKAISVLEMENGYWWC.RKKQRAGVMLTVGTKPDGSPDERWCF	704
694	SKNIWKLQRAITILDTEKSFLKCMRKAFRSGKLLQVGYTPDGKDDYRWCF	743
705	RVEEVNWASWEOTLPTLCEDPSGA.GVFRIBERTVILLETTREE	753
744	RADEANALLANINAGITUEDEGUCEGAKLIPITETETETETETETETETETETETETETETETETETET	789
754	NYVPVQLLQSN	764
790	: . NFALVPLLREASARDRQSAQPEEVYLRQFSGSLKPEDAEVFKSPAASGEK	839

GAP of: humanvr2.seq check: 8853 from: 1 to: 2809 humanVR2 21e11a, 2809 bases, 8853 checksum. to: humanvrl.seq check: 4554 from: 1 to: 3909 humanVR1 Fbh18547pat - Import - complete Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/nwsgapdna.cmp CompCheck: 8760 50 Average Match: 10.000 Gap Weight: 3 Average Mismatch: 0.000 Length Weight: Length: 3934 Quality: 14359 Ratio: 5.112 Gaps: 15 Percent Similarity: 55.316 Percent Identity: 55.316 Match display thresholds for the alignment(s): | = IDENTITY1 humanvr2.seq x humanvr1.seqGGCTAGCCTGTCCTGACAGGGGAGAG 26 1 1 1 1 1 11 111 801 TGTCCACAGTAGTCCCCCCTTATCCACGGGTGTCACTTTCCATGGGTTCA 850 27 TTAAGCTCCCGTTCTCCACCGTGCCGGCTGGCCAGGTGGGCTGAGGGTGA 76 1 1 851 GTTATTTGCGGTCAACCACGGTCTGCCAATATTAAATGGAAAATTCTTCA 900 77 CCGAGAGACCAGAACCTGCTTGCTGGAGCTTAGTGCTCAGAGCTGGGGAG 126 901 AACAGTTCCCAAGTTTTCCCTTGTGCATTGTTCTGAGCAGTGTGATGAAG 950 127 GGAGGTTCCGCCGCTCCTCTGCTGTCAGCGCCGGCAGCCCCTCCCGGCTT 176 111 1 1 951 AGTCTCTGCCGTGCCATCTGGGATGCAAACCGTCCCTGTGTCCCCCACGT 1000 177 CACTTCCTCCCGCAGCCCCTGCTACTGAGAAGCTCCGGGATCCCAGCAGC 226 111 11 11 11 1 11 1 1 1001 CCAGGCCGTAGATGCTCCCCGCCGGTCAGTCACTTAGTCGTCAGATCGCC 1050CTCAGCCTGCGGG 253 227 CGCCACGCCCTGGC..... 11 111 11 * 11 1 1 1 1 1051 CGTCCTGGTATCACAGTGCTTCTGTTCAGGTTGCACACTGGGCCACAGAG 1100

254	GCTCCAGTCAGGCCAACACCGACGCGCAGCTGGGAGGAAG	293
1101	GATCCAGCAAGGATGAAGAAATGGAGCACAGACTTGGGGACAGCTGC	1150
294	ACAGGACCTTGACATCTCCATCTGCACAGAGGTCCTG	331
1151	GGACCCACTCCAAAAGGACACCTGCCCAGACCCCTGGATGGA	1200
	GCTGGACCGAGCAGCCTCCTCCTCCTAGGATGACCTCACCCTCCAGCT	
1201	ACTCCAGGCCACCTCCAGCCAAGCCCCAGGCCCACGGCCAAGAGCCGC	1250
	CTCCAGTTTCAGGTTGGAGACATTAGATGGAGGCCAAGAAGATGGCTCT	
	ACCCGGCTCTTTGGGAAGGGTGACTCGGAGGAGGCTTTCCCGGTGGATTG	
	GAGGCGGACAGAGGAAAGCTGGATTTTGGGAGCGGGCTGCCTCCCATGGA	
	CCCCCACGAGGAAGGTGAGTTGGACTCCTGCCCGACCATCACAGTCAGCC	
	GTCACAGTTCCAGGGCGAGGACCGGAAATTCGCCCCTCAGATAAGAGTCA	
	CTGTTATCACCATCCAGAGGCCAGGAGACGCCCCACCGGTGCCAGGC	
530	ACCTCAACTACCGAAAGGGAACAGGTGCCAGTCAGCCGGATCCAAACCGA	579
1399	TGCTGTCCCAGGACTCTGTCGCCGCCAGCACCGAGAAGACCCTCAGGCTC	1448
580	TTTGACCGAGATCGGCTCTTCAATGCGGTCTCCCGGGGTGTCCCCGAGGA	629
	TATGATCGCAGGAGTATCTTTGAAGCCGTTGCTCAGAATAACTGCCAGGA	
	TCTGGCTGGACTTCCAGAGTACCTGAGCAAGACCAGCAAGTACCTCACCG	
	TCTGGAGAGCCTGCTCTTCCTGCAGAAGAGCAAGAAGCACCTCACAG	
	ACTCGGAATACACAGAGGGCTCCACAGGTAAGACGTGCCTGATGAAGGCT	
	ACAACGAGTTCAAAGACCCTGAGACAGGGAAGACCTGTCTGCTGAAAGCC	
.730	GTGCTGAACCTTAAGGACGGAGTCAATGCCTGCATTCTGCCACTGCTGCA	779
1599	ATGCTCAACCTGCACGACGGACAGAACACCACCATCCCCTGCTCCTGGA	1648
780	GATCGACAGGGACTCTGGCAATCCTCAGCCCCTGGTAAATGCCCAGTGCA	829
1649	GATCGCGCGCAAACGGACAGCCTGAAGGAGCTTGTCAACGCCAGCTACA	1698
830	CAGATGACTATTACCGAGGCCACAGCGCTCTGCACATCGCCATTGAGAAG	879
1699		.1748
880	AGGAGTCTGCAGTGTGAAGCTCCTGGTGGAGAATGGGGCCAATGTGCA	929
1749	CGCAACATGGCCCTGGTGACCCTCCTGGTGGAGAACGGAGCAGACGTCCA	1798
	TGCCCGGGCCTGCGGCCGCTTCTTCCAGAAGGGCCAAGGGACTTGCT	
1799	GGCTGCGGCCCATGGGGACTTCTTTAAGAAAACCAAAGGGCGGCCTGGAT	1848

16/35

977	TTTATTTCGGTGAGCTACCCCTCTCTTTGGCCGCTTGCACCAAGCAGTGG	1026
1849	TCTACTTCGGTGAACTGCCCCTGTCCCTGGCCGCGTGCACCAACCA	1898
1027	GATGTGGTAAGCTACCTCCTGGAGAACCCACACCAGCCCGCCAGCCTGCA	1076
1899	GGCATCGTGAAGTTCCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAG	1948
1077	GGCCACTGACTCCCAGGGCAACACAGTCCTGCATGCCCTAGTGATGATCT	1126
1949	CGCCAGGGACTCGGTGGGCAACACGGTGCTGCACGCCCTGGTGGAGGTGG	1998
1127	CGGACAACTCAGCTGAGAACATTGCACTGGTGACCAGCATGTATGATGGG	1176
1999	CCGACAACACGGCCGACAACACGAAGTTTGTGACGAGCATGTACAATGAG	2048
1177	CTCCTCCAAGCTGGGGCCCGCCTCTGCCCTACCGTGCAGCTTGAGGACAT	1226
2049	ATTCTGATGCTGGGGGCCAAACTGCACCCGACGCTGAAGCTGGAGGAGCT	2098
1227	CCGCAACCTGCAGGATCTCACGCCTCTGAAGCTGGCCGCCAAGGAGGGCA	1276
2099	CACCAACAAGAAGGGAATGACGCCGCTGGCTCTGGCAGCTGGGACCGGGA	2148
	AGATCGAGATTTTCAGGCACATCCTGCAGCGGGAGTTTTCAGGA	
	AGATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAG	
	CTGAGCCACCTTTCCCGAAAGTTCACCGAGTGGTGCTATGGGCCTGTCCG	
	TGCAGGCACCTGTCCAGGAAGTTCACCGAGTGGGCCTACGGGCCCGTGCA	
	GGTGTCGCTGTATGACCTGGCTTCTGTGGACAGCTGTGAGGAGAACTCAG	
	CTCCTCGCTGTACGACCTGTCCTGCATCGACACCTGCGAGAAGAACTCGG	
	TGCTGGAGATCATTGCCTTTCATTGCAAGAGCCCGCACCGACACCGA	
	TGCTGGAGGTGATCGCCTACAGCAGCAGCGAGACCCCTAATCGCCACGAC	
	ATGGTCGTTTTGGAGCCCCTGAACAACTGCTGCAGGCGAAATGGGA	
	ATGCTCTTGGTGGAGCCGCTGAACCGACTCCTGCAGGACAAGTGGGACAG	
	TCTGCTCATCCCCAAGTTCTTCTTAAACTTCCTGTGTAATCTGATCTACA	
	ATTCGTCAAGCGCATCTTCTACTTCAACTTCCTGGTCTACTGCCTGTACA	
	TGTTCATCTTCACCGCTGTTGCCTACCATCAGCCTACCCTGAAGAAGCAG	
	TGATCATCTTCACCATGGCTGCCTACTACAGGCCCGTGGATGGCTT	
	GCCGCCCTCACCTGAAAGCGGAGGTTGGAAACTCCATGCTGCTGACGGG	
2495	GCCTCCCTTTA AGATGGAAAAATTGGAGACTATTTCCGAGTTACTGG	434 2

1665	CCACATCCTTATCCTGCTAGGGGGGATCTACCTCCTCGTGGGCCAGCTGT	171
2543	AGAGATCCTGTCTGTTAGGAGGAGTCTACTTCTTTTTCCGAGGGATTC	2592
1715	GGTACTTCTGGCGGCGCCACGTGTTCATCTGGATCTCGTTCATAGACAGC	1764
2593	AGTATTTCCTGCAGAGGCGGCCGTCGATGAAGACCCTGTTTGTGGACAGC	2642
1765	TACTTTGAAATCCTCTTCCTGTTCCAGGCCCTGCTCACAGTGGTGTCCCA	1814
2643	TACAGTGAGATGCTTTTCTTCTGCAGTCACTGTTCATGCTGGCCACCGT	2692
	GGTGCTGTGTTTCCTGGCCATCGAGTGGTACCTGCCCCTGCTTGTGTCTG	
2693	GGTGCTGTACTTCAGCCACCTCAAGGAGTATGTGGCTTCCATGGTATTCT	2742
1865	CGCTGGTGCTGGCTGAACCTGCTTTACTATACACGTGGCTTCCAG	1914
2743	CCCTGGCCTTGGGCTGGACCAACATGCTCTACTACACCCGCGGTTTCCAG	2792
1915	CACACAGGCATCTACAGTGTCATGATCCAGAAGGTCATCCTGCGGGACCT	1964
2793	CAGATGGGCATCTATGCCGTCATGATAGAGAAGATGATCCTGAGAGACCT	2842
1965	GCTGCGCTTCCTTCTGATCTACTTAGTCTTCCTTTTCGGCTTCGCTGTAG	2014
2843	GTGCCGTTTCATGTTTGTCTACATCGTCTTCTTGTTCGGGTTTTCCACAG	2892
	CCCTGGTGAGCCTGAGCCAGGAGGCTTGGCGCCCCGAAGCTCCTACAGGC	
	CGGTGGTGACGCTGATTGAAGACGGGAAGAATGACTCCCTGCCGTCTGAG	
2065	CCCAATGCCACAGAGTCAGTGCAGCCCATGGAGGACAGGAGGACGAGGG	2114
2943	TCCACGTCGCACAGGTGGCGGGGCCTGCCTGCAGGCC	2980
2115	CAACGGGGCCCAGTACAGGGGTATCCTGGAAGCCTCCTTGGAGCTCTTCA	2164
2981	CCCCGATAGCTCCTACAACAGCCTGTACTCCACCTGCCTG	3030
2165	AATTCACCATCGGCATGGGCGAGCTGCCTTCCAGGAGCAGCTGCACTTC	2214
3031	AGTTCACCATCGGCATGGGCGACCTGGAGTTCACTGAGAACTATGACTTC	3080
2215	CGCGGCATGGTGCTGCTGCTGCTGCTGCTGCTGCTGCTCACCTACAT	2264
3081	AAGGCTGTCTTCATCATCCTGCTGCTGGCCTATGTAATTCTCACCTACAT	3130
2265	CCTGCTGCTCAACATGCTCATCGCCCTCATGAGCGAGACCGTCAACAGTG	2314
3131	CCTCCTGCTCAACATGCTCATCGCCCTCATGGGTGAGACTGTCAACAAGA	3180
2315	TCGCCACTGACAGCTGGAGCATCTGGAAGCCATCTCTGTC	2364
		2220

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	CTGGAGATGGAGAATGGCTATTGGTGGTGCAGGAAGAAGCAGCGGGC	
3231	CTGGACACGGAGAAGACCTTCCTTAAGTGCATGAGGAAGGCCTTCCGCTC	3280
2412	AGGTGTGATGCTGACCGTTGGCACTAAGCCAGATGGCAGCCCGGATGAGC	2461
3281	AGGCAAGCTGCTGCAGGTGGGGTACACCCTGATGGCAAGGACGACTACC	3330
2462	GCTGGTGCTTCAGGGTGGAGGTGAACTGGGCTTCATGGGAGCAGACG	2511
3331	GGTGGTGCTTCAGGGTGGACGAGGTGAACTGGACCACCTGGAACACCAAC	3380
2512	CTGCCTACGCTGTGTGAGGACCCGTCAGGGGCAGGTGTCCCTCGAAC	2558
3381		3430
2559	TCTCGAGAACCCTGTCCTGGCTTCCCCTCCCAAGGAGGATGAGGAT	2604
3431		3480
2605	GGTGCCTCTGAGGAAAACTATGTGCCCGTCCAGCTCCTCCAGTCCAACTG	2654
3481	ACTTTGCCCTGGTCCCCCTTTTAAGAGAGGCAAGTGCTCGAGATAGGCAG	3530
2655	ATGGCCCAGATGCAGCAGGAGGCCAGAGGACAGAGCAGAGGATCTTTCCA	2704
3531	TCTGCTCAGCCCGAGGAAGTTTATCTGCGACAGTTTTCAGGGTCTCTGAA	3580
	· · · · · · · · · · · · · · · · · · ·	
3626	GAAGTGAGGACGTCACGCAGACAGCACTGTCAACACTGGGCCTTAGGAGA	3675
2805	AAAAA	2809
	CCCCGTTGCCACGGGGGGCTGCTGAGGGAACACCAGTGCTCTGTCAGCAG	
.3D/D	CCCCG1 10CC 1CCCCCCCCCC TOTAL TITLE TOTAL	

CLUSTAL W (1.74) multiple sequence alignment

humanVR2 ratVR2	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY
humanVR2 ratVR2	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2 ratVR2	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK
humanVR2 ratVR2	LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQASTHASALSLAACTKQWDVVTYLLENPHQPASLEA
humanVR2 ratVR2	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSLGNTVLHALVMIADNSPENSALVIHMYDGLLQMGARLCPTVQLEEISNHQGLTPLKL
humanVR2 ratVR2	AAKEGKIEIFRHILQREFSG-LSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIA AAKEGKIEIFRHILQREFSGPYQPLSRKFTEWCYGPVRVSLYDLSSVDSWEKNSVLEIIA
humanVR2 ratVR2	FHCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQA FHCKSPNRHRMVVLEPLNKLLQEKWDRLVSRFPFNFACYLVYMFIFTVVAYHQPSLDQPA
humanVR2 ratVR2	APHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQAL IPSSKATFGESMLLLGHILILLGGIYLLLGQLWYFWRRRLFIWISFMDSYFEILFLLQAL * * .*:*** ****************************
humanVR2 ratVR2	Ltvvsqvicflaiewylpilvsalvigwinilyytrgfqhtgiysvmiqkvilrdilrfl Ltvlsqvirfmetewylpilvisivigwinilyytrgfqhtgiysvmiqkvilrdilrfl
humanVR2 ratVR2	Liylvflfgfavalvslsqeawrpeaptgpnatesvqpmegqedegngaqyrgileasle Lvylvflfgfavalvslsrearspkapednnstvteqptvgqeeepapyrsildasle *:***********************************
humanVR2 ratVR2	LFKFTIGNGELAFQEQIHPRGMVILLLAYVILTYIILINMIIAIMSETVNSVATDSNSI LFKFTIGNGELAFQEQIRPRGVVILLILAYVILTYVILINMIIAIMSETVNHVADRSNSI ***********************************
humanVR2 ratVR2	MKLQKAISVLEMENGYNNCR-KKQRAGVML/TVGTKPDGSPDERNCFRVEEVNWASUBQTL MKLQKAISVLEMENGYNNCRRKKERBGRLLKVGTRGDGTPDERNCFRVEEVNWAAWBKTL
humanVR2 ratVR2	PTLCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN PTLSEDPSGPGITGNKKNPTSK-PGKNSASEEDHLPLQVLQSP ***.*****. : : * * : . * * : . * * : . * * * : . * * * : . * * * : . * * * : . * * * : . * * * *

PCT/US99/26701 WO 00/29577

20/35

GAP of: ratvr2.pep check: 9190 from: 1 to: 554

ratVR2 Flrxb147g11

to: humanvr2.pep check: 5746 from: 1 to: 764

humanVR2 Flh21e11

Symbol comparison table: /usr/local/gcg_9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430

Average Match: 2.912 12 Gap Weight: Average Mismatch: -2.003 Length Weight:

766 Length: 2182 Quality: Gaps: Ratio: 3.939 Percent Identity: 79.167 Percent Similarity: 81.703

Match display thresholds for the alignment(s):

= IDENTITY

1

ratvr2.pep x humanvr2.pep

1STHASALSLAACTKQWDVVTYLLENPHQPASLEATDSLGNTVLH 44 ពាពា ពាធារពេយពេយពេយពេយ 201 GQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQATDSQGNTVLH 250 45 ALVMIADNSPENSALVIHMYDGLLQMGARLCPTVQLEEISNHQGLTPLKL 94 251 ALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL 300 95 AAKEGKIEIFRHILQREFSGPYQPLSRKFTEWCYGPVRVSLYDLSSVDSW 144 301 AAKEGKIEIFRHILOREFSG.LSHLSRKFTEWCYGPVRVSLYDLASVDSC 349 145 EKNSVLEI LAFHCKSPNRHRMVVLEPLNKLLQEKHDRLVSRFFFNFACYL 194 195 VYMFIFTVVAYHQPSLDQPAIPSSKATFGESNLLIGHILILIGGIYLLIG 244 245 QLMYFWRRRLFIWISFMDSYFEILFLLQALLTVLSQVLRFMETEWYLPIL 294 તાલામ ના લામનાલાલામાં તેલામનાલા છે. તેલાલા 450 QLMYFWRRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEWYLPLL 499 295 VLSLVLGWLNLLYYTRGFQHTGIYSVNIQKVIIRDLLRFLLVYLVFLFGF 344 500 VEALVLGHINLLYYTRGFQHTGIYEVMIQKVIIRDLLRFLLIYLVFLFGF 549

345 AVALVELSREARSPKAPEDNINSTVTEQPTVGQERE..PAPYRSILDASLE 392

550 AVALVELSQEAMRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLE 599

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202	LFKFTIGMGELAFOEOLRFRGVVLLLLLAYVLLTYVLLLNMLIALMSETV	442
600	LFKFTIGMGELAFQEQLHFRGWVLLLLLAYVLLTYILLLMMLIALMSETV LFKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLMMLIALMSETV	649
000	·	492
443	NHVADNSWSIWKLQKAISVLEMENGYWWC.RKKURGSIWKLQKAISVLEMENGYWWC.RKKQRAGVMLTVGTKPDGSP NSVATDSWSIWKLQKAISVLEMENGYWWC.RKKQRAGVMLTVGTKPDGSP	698
650	NSVATDSWSIWKLQKAIS VLENBROTHMOTHMOTHMOTHMOTHMOTHMOTHMOTHMOTHMOTHM	
493	DERWCFRVEEVNWAAWEKTLPTLSEDPSGPGITGNKKNPTSKPGKN	538
699	DERWCFRVEEVNWASWEQTLPTLCEDPSGAGVPRTLENPVLASPPKEDED DERWCFRVEEVNWASWEQTLPTLCEDPSGAGVPRTLENPVLASPPKEDED	748
	SASEEDHLPLQVLQSP 554	
749	GASEENYVPVQLLQSN 764	

GAP of: humanvrl.seq check: 4554 from: 1 to: 3909 humanVR1 Fbh18547pat - Import - complete to: ratvrl.seq check: 7921 from: 1 to: 2847 ratVR1.seq AF029310 in GenBank Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/nwsgapdna.cmp CompCheck: 8760 Average Match: 10.000 .50 Gap Weight: 3 Average Mismatch: 0.000 Length Weight: 3914 Length: Quality: 22717 Ratio: 7.979 10 Gaps: Percent Similarity: 82.125 Percent Identity: 82.125 Match display thresholds for the alignment(s): ! = IDENTITY 5 humanvrl.seq x ratvrl.seq .

1001 CCAGGCCGTAGATGCTCCCCGCCGGTCAGTCACTTAGTCGTCAGATCGCC 1050 1 11 1 11 ..CAGCTCCAAGGCACTTGCTCC 21 1051 CGTCCTGGTATCACAGTGCTTCTGTTCAGGTTGCACACTGGGCCACAGAG 1100 22 ATTTGGGGTGTGCCTGCACCT...AGCTGGTTGCAAATTGGGCCACAGAG 68 1101 GATCCAGCAAGGATGAAGAAATGGAGCAGCACAGACTTGGGGACAGCTGC 1150 1111 | 1111111 | 1111 | 111 | 111 | 111 | 1 69 GATCTGGAAAGGATGGAACAACGGGCTAGCTTAGACTCAGAGGAGTCTGA 118 1201 ACTCCAGGCCACCTCCAGCCAAGCCCCAGGCCCACGGCCAAGAGCCGC 1250 iii ir imnimm maana ac ac ac ac ac 169 ACTGCAAGCCACCTCCAGTCAAGCCCCACATCTTCACTACCAGGAGTCGT 218 1251 ACCCGGCTCTTTGGGAAGGGTGACTCGGAGGAGGCTTTCCCGGTGGATTG 1300 219 ACCCGGCTTTTTGGGAAGGGTGACTCGGAGGAGGCCTCTCCCCTGGACTG 268 1301 CCCCCACGAGGAAGGTGAGTTGGACTCCTGCCCGACCATCACAGTCAGCC 1350 269 CCCTTATGAGGAAGGCGGGCTGGCTTCCTGCCCTATCATCACTGTCAGCT 318 1351 CTGTTATCACCATCCAGAGGCCAGGAGACGGCCCCACCGGTGCCAGGCTG 1400 319 CTGTTCTAACTATCCAGAGGCCTGGGGATGGACCTGCCAGTGTCAGGCCG 368

1401	CTGTCCCAGGACTCTGTCGCCGCCAGCACCGAGAAGACCCTCAGGCTCTA	1450
	TGATCGCAGGAGTATCTTTGAAGCCGTTGCTCAGAATAACTGCCAGGATC	
416	TGATCGCAGGAGCATCTTCGATGCTGTGGCTCAGAGTAACTGCCAGGAGC	465
	TGGAGAGCCTGCTGCTCCTGCAGAAGAAGCAAGAAGCACCTCACAGAC	
	TGGAGAGCCTGCCCTTCCTGCAGAGGAGCAAGAAGCGCCTGACTGA	
	AACGAGTTCAAAGACCCTGAGACAGGGAAGACCTGTCTGCTGAAAGCCAT	
	AGCGAGTTCAAAGACCCAGAGACAGGAAAGACCTGTCTGCTAAAAGCCAT	565
	GCTCAACCTGCACGACGACAGACACCACCATCCCCTGCTCCTGGAGA	
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		765
	CTCCCCCACTTCTTTAAGAAACCAAAGGGCGGCCTGGATTC	1850
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	TACTTCGGTGAACTGCCCCTGTCCCTGGCCGCGTGCACCAACCA	
	TACTTTGGTGAGCTGCCCCTGTCCCTGGCTGCGTGCACCAACCA	
	CATCGTGAAGTTCCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAGCG	
866	CATTGTGAAGTTCCTGCTGCAGAACTCCTGGCAGCCTGGAAGTCTTGTGAAGTTCTTGCTGCAGAACTCCTGGCAGCCTGCAGCCTGGCAGCCTGCAGCCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCTGCAGCCCTAGCCCTGCAGCCCTGCAGCCCTGCAGCCCTAGCCCTAGCAAGCCCTAGCCCTAGCCCTAGCAAGCCCTAGCAAGCCCTAGCAAGCCCTAGCAAGCCCTAGCAAGCCCTAGCAAGCCCTAGCAAACCCTAGCAAGCCCTAGCAAACCCTAGCAAACAAA	
	CCAGGGACTCGGTGGCAACACGGTGCTGCACGCCCTGGTGGAGGTGGCC	
	CCCGGGACTCAGTGGGCAACACGGTGCTTCATGCCCTGGTGGAGGTGGCA	
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	GATAACACAGTTGACAACACCCAAGTTCGTGACAAGCATGTACAACGAGAT	
	TCTGATGCTGGGGGCCAAACTGCACCCGACGCTGAAGCTGGAGGAGCTCA	
1016	CTTGATCCTGGGGGCCAAACTCCACCCCACGCTGAAGCTGGAAGAGATCA	1065

2101	CCAACAAGAAGGGAATGACGCCGCTGGCTCTGGCAGCTGGGACCGGGAAG	2150
1066	CCAACAGGAAGGGCTCACGCCACTGGCTCTGGCTGCAGCAGTGGGAAG	1115
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	ATCGGGGTCTTGGCCTACATTCTCCAGAGGGAGATCCATGAACCCGAGTG	
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2598	TTCCTGCAGAGGCGGCCGTCGATGAAGACCCTGTTTGTGGACAGCTACAG	
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	TGAGATGCTTTTCTTCTGCAGTCACTGTTCATGCTGGCCACCGTGGTGC	
	TGAGATACTTTTCTTTGTACAGTCGCTGTTCATGCTGGTGTCTGTGGTAC	
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	TGTACTTCAGCCAACGCAAGGAGTATGTGGCTTCCATGGTGTTCTCCCCTG	
2748	GCCTTGGGCTGGACCAACATGCTCTACTACACCCGCGGTTTCCAGCAGAT	2797
1716	CCCATGGGCTGGACCAACATGCTCTACTATACCCGAGGATTCCAGCAGAT	1765

2798	GGGCATCTATGCCGTCATGATAGAGAAGATGATCCTGAGAGACCTGTGCC	2847
1766	GGGCATCTATGCTGTCATGATTGAGAAGATGATCCTCAGAGACCTGTGCC	1815
2848	GTTTCATGTTTGTCTACATCGTCTTCTTGTTCGGGTTTTCCACAGCGGTG	2897
1816	GGTTTATGTTCGTCTACCTCGTGTTCTTGTTTGGATTTTCCACAGCTGTG	1865
2898	GTGACGCTGATTGAAGACGGGAAGAATGACTCCCTGCCGTCTGAGTCCAC	2947
1866	GTGACACTGATTGAGGATGGGAAGAATAACTCTCTGCCTATGGAGTCCAC	1915
2948	GTCGCACAGGTGGCGGGGCCTGCCTGCAGGCCCCCGATAGCTCCTACA	2997
1916	ACCACACAAGTGCCGGGGGTCTGCCTGCAAGCCAGGTAACTCTTACA	1962
2998	ACAGCCTGTACTCCACCTGCCTGGAGCTGTTCAAGTTCACCATCGGCATG	3047
1963	ACAGCCTGTATTCCACATGTCTGGAGCTGTTCAAGTTCACCATCGGCATG	2012
3048	GGCGACCTGGAGTTCACTGAGAACTATGACTTCAAGGCTGTCTTCATCAT	3097
2013	GGCGACCTGGAGTTCACTGAGAACTACGACTTCAAGGCTGTCTTCATCAT	2062
3098	CCTGCTGCTGGCCTATGTAATTCTCACCTACATCCTCCTGCTCAACATGC	3147
2063	CCTGTTACTGGCCTATGTGATTCTCACCTACATCCTTCTGCTCAACATGC	2112
3148	TCATCGCCCTCATGGGTGAGACTGTCAACAAGATCGCACAGGAGAGCAAG	3197
2113	TCATTGCTCTCATGGGTGAGACCGTCAACAAGATTGCACAAGAGAGCAAG	2162
	AACATCTGGAAGCTGCAGAGAGCCATCACCATCCTGGACACGGAGAAGAG	3247
	AACATCTGGAAGCTGCAGAGAGCCATCACCATCCTGGATACAGAGAAGAG	
	CTTCCTTAAGTGCATGAGGAAGGCCTTCCGCTCAGGCAAGCTGCTGCAGG	
	CTTCCTGAAGTGCATGAGGAAGGCCTTCCGCTCTGGCAAGCTGCTGCAGG	
	TGGGGTACACACCTGATGGCAAGGACGACTACCGGTGGTGCTTCAGGGTG	
	TGGGGTTCACTCCTGACGGCAAGGATGACTACCGGTGGTGTTTCAGGGTG	
	GACGAGGTGAACTGGACCACCTGGAACACCAACGTGGGCATCATCAACGA	
	GACGAGGTAAACTGGACTACCTGGAACACCAATGTGGGTATCATCAACGA	
•-	AGACCCGGGCAACTGTGAGGGCGTCAAGCGCACCCTGAGCTTCTCCCTGC	
	GGACCCAGGCAACTGTGAGGGCGTCAAGCGCACCCTGAGCTTCTCCCTGA	
	GGTCAAGCAGAGTTTCAGGCAGACACTGGAAGAACTTTGCCCTGGTCCCC	
	GGTCAGGCCGAGTTTCAGGGAGAAACTGGAAGAACTTTGCCCTGGTTCCC	
3498	CTTTTAAGAGAGGCAAGTGCTCGAGATAGGCAGTCTGCTCAGCCCGAGGA	3547

2463	CTTCTGAGGGATGCAAGCACTCGAGATAGACATGCCACCCAGCAGGAAGA	2512
	AGTTTATCTGCGACAGTTTTCAGGGTCTCTGAAGCCAGAGGACGCTGAGG	
	AGTTCAACTGAAGCATTATACGGGATCCCTTAAGCCAGAGGATGCTGAGG	
	TCTTCAAGAGTCCTGCCGCTTCCGGGGAGAAGTGA.GGACGTCACGCAGA	
	TTTTCAAGGATTCCATGGTCCCAGGGGAGAATAATGGACACTATGCAGG	
3647	CAGCACTGTCAACACTGGGCCTTAGGAGACCCCGTTGCCACGGGGGGCTG	3696
2613	GATCAATGCGGGGTCTTTGGGTGGTCTG	2640
	CTGAGGGAACACCAGTGCTCTGTCAGCAGCCTGGCCTGG	
	CTTAGGGAAC. CAGCAGGGTTGACGTTATCTGGGTCCACTCTGTGCCTGC	
	CCA.GCATGTTCCCAAATCTGTGCTGGACAAGCTGTGGGAAGCGTTCTTG	
	CTAGGCACATTCCTAGGACTTCGGCGGGCCTGCTGTGGGAA.CTGGGAGG	
	GAAGCATGGGGAGTGATGTACATCCAACCGTCACTGTCCCCAAGTGAATC	
2739	TGTGTGGGAATTGAGATGTGTATCCAACCATGA1C1CCAAACCATTGA	2785
	TCCTAACAGACTTTCAGGTTTTTACTCACTTTACTAAAAAAAA	
2786	GCTTTCAACTCTTTATGGACTTTATTAAACAGAGTGAATGGCAAATCTCT	2835
3896	AGGGCGGCCTTA 3909	
2836	ACTTGGACACAT 2847	

27/35

GAP of: humanvrl.pep check: 6877 from: 1 to: 839 humanVR1 Fbh18547pat - fchrb87a6, 3909 bases, 4554 checksum. to: ratvrl.pep check: 5764 from: 1 to: 838 ratVR1 | AF029310 Rattus norvegicus vanilloid receptor subtype 1 mRNA, complete cds. Symbol comparison table: /ddm_local/gcg/gcg_9.1/gcgcore/data/rundata/blosum62.cmp CompCheck: 6430 12 Gap Weight: Average Match: 2.912 Length Weight: Average Mismatch: -2.003 Quality: Length: 840 Ratio: 4.456 Gaps: Percent Similarity: 89.247 Percent Identity: 86.022 Match display thresholds for the alignment(s): | = IDENTITY2 : = humanvrl.pep x ratvrl.pep 1 MKKWSSTDLGTAADPLOKDTCPDPLDGDPNSRPPPAKPOLPTAKSRTRLF 50 1 MEQRASLDSEESESPPQENSCLDPPDRDPNCKPPPVKPHIFTTRSRTRLF 50 51 GKGDSEEAFPVDCPHEEGELDSCPTITVSPVITIQRPGDGPTGARLLSOD 100 51 GKGDSEEASPLDCPYEEGGLASCPIITVSSVLTIQRPGDGPASVRPSSQD 100 101 SVAASTEKTLRLYDRRSIFEAVAQNNCQDLESLLLFLQKSKKHLTDNEFK 150 101 SVSAG.EKPPRLYDRRSIFDAVAQSNCQELESLLPFLQRSKKRLTDSEFK 149 151 DPETGKTCLLKAMLNLHDGQNTTIPLLLEIARQTDSLKELVNASYTDSYY 200 150 DPETGKTCLLKAMLNLHNGQNDTIALLLDVARKTDSLKQFVNASYTDSYY 199 201 KGOTALHIAIERRNMALVTLLVENGADVQAAAHGDFFKKTKGRPGFYFGE 250 200 KGQTALHIAIERRNMTLVTLLVENGADVQAAANGDFFKKTKGRPGFYFGE 249 251 LPLSLAACTNQLGIVKFLLQNSWQTADISARDSVGNTVLHALVEVADNTA 300 250 LPLSLAACTNQLAIVKFLLQNSWQPADISARDSVGNTVLHALVEVADNTV 299 301 DNTKFVTSMYNEILMLGAKLHPTLKLEELTNKKGMTPLALAAGTGKIGVL 350 300 DNTKFVTSMYNEILILGAKLHPTLKLEEITNRKGLTPLALAASSGKIGVL 349 351 AYILQREIQEPECRHLSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI 400 350 AYILQREIHEPECRHLSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI 399

	AYSSSETPNRHDMLLVEPLNRLLODKWDRFVKRIFYFNFLVYCLYMIIFT	450
400	AYSSSETPNRHDMLLVEPLNRLLQDKWDRFVKRIFYFNFFVYCLYMIIFT	449
451	MAAYYRPVDGLPPFKMEK.IGDYFRVTGEILSVLGGVYFFFRGIQYFLQR	499
		400
450	AAAYYRPVEGLPPYKLKNTVGDYFRVTGEILSVSGGVYFFFRGIQYFLQR	499
500	RPSMKTLFVDSYSEMLFFLOSLFMLATVVLYFSHLKEYVASMVFSLALGW	549
500	RPSLKSLFVDSYSEILFFVQSLFMLVSVVLYFSQRKEYVASMVFSLAMGW	549
c c 0	TNMLYYTRGFQQMGIYAVMIEKMILRDLCRFMFVYIVFLFGFSTAVVTLI	599
550	TNMLYYTRGFQQMGIYAVMIEKMILRDLCRFMFVYLVFLFGFSTAVVTLI	599
600	EDGKNDSLPSESTSHRWRGPACRPPDSSYNSLYSTCLELFKFTIGMGDLE	649
600	EDGKNNSLPMESTPHKCRGSACK.PGNSYNSLYSTCLELFKFTIGMGDLE	648
	FTENYDFKAVFIILLLAYVILTYILLLNMLIALMGETVNKIAQESKNIWK	699
649	FTENYDFKAVFIILLLAYVILTYILLLNMLIALMGETVNKIAQESKNIWK	698
	•	749
699	LQRAITILDTEKSFLKCMRKAFRSGKLLQVGFTPDGKDDYRWCFRVDEVN	748
		799
	WTTWNTNVGIINEDPGNCEGVKRTLSFSLRSSRVSGRHWKNFALVPLLRE	
749	WITWNTNVGIINEDPGNCEGVKRTLSFSLRSGRVSGRNWKNFALVPLLRD	798
	•	
800	ASARDROSAOPEEVYLROFSGSLKPEDAEVFKSPAASGEK 839	
700	ASTRORHATQQEEVQLKHYTGSLKPEDAEVFKDSMVPGEK 838	
199	WO I University Africa - a	

CLUSTAL W (1.74)	multiple sequence alignment
humanVR2.alt humanVR2	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY
humanVR2.alt humanVR2	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2.alt humanVR2	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK
humanVR2.alt humanVR2	LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA
humanVR2.alt humanVR2	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL
humanVR2.alt humanVR2	AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF AAKEGKIBIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF
humanVR2.alt humanVR2	HCKSPHRHRMVVLEPLNKILQAKWDLLIPKPFINFLCNLIYMFIFTAVAYHQPTLKKQAA HCKSPHRHRMVVLEPLNKILQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA
humanVR2.alt humanVR2	PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL
humanVR2.alt humanVR2	TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQTVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLL
humanVR2.alt humanVR2	IYLVFLFGFAVALVSLSQEAWRPBAPTGPNATESVQPMEGQEDEGNGAQYRGILBASLBL
humanVR2.alt humanVR2	PKFTIGMGBLAPQBQLHFRGMVLLLLLAYVLLTYILLLMMLIALMSBTVNSVATDSWSIW
humanVR2.alt humanVR2	KKAISVLEMENGYNNCRKKQRAGVMLTVGTKPDGSPDERNCFRVEEVNWASNEQTLPT KLQKAISVLEMENGYNNCRKKQRAGVMLTVGTKPDGSPDERNCFRVEEVNWASNEQTLPT :************************************
humanVR2.alt humanVR2	LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN

FIGURE 11

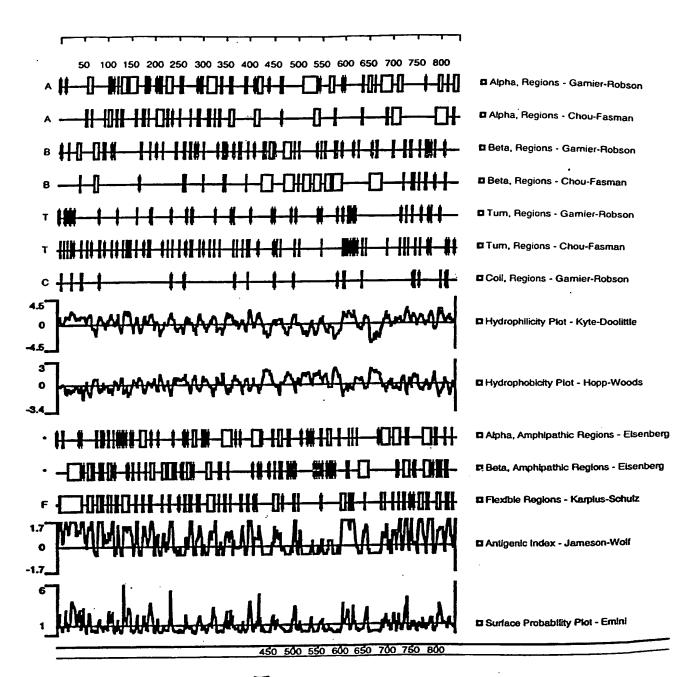


FIGURE 12

Protein Family / Domain Matches, HMMer version 2

```
Searching for complete domains
 hmmpfam - search a single seq against HMM database HMMER 2.1.1 (Dec 1998)
 Copyright (C) 1992-1998 Washington University School of Medicine
 HMMER is freely distributed under the GNU General Public License (GPL).
 HMM file:
                        /prod/ddm/seqanal/PFAM/pfam4.2/Pfam
 Sequence file:
                         /usr/ns-home/docs/seqanal/orfanal/oa-script.18670.seq
 Query: hVR-1
Scores for sequence family classification (score includes all domains):
Model Description
                                                     Score
                                                             E-value N
<u>ank</u>
         Ank repeat
                                                       51.5
                                                             1.9e-11 3
Parsed for domains:
Model
       Domain seq-f seq-t
                              hmm-f hmm-t
                                              score E-value
                 201 233 ..
ank
          1/3
                                1 33 []
                                               34.4
                                                    2.6e-06
                                 ī
          2/3
                        283 ..
ank
                                      33 []
33 []
                  248
                                               13.2
                        361 ..
          3/3
                  333
                                                3.4
Alignments of top-scoring domains:
ank: domain 1 of 3, from 201 to 233: score 34.4, E = 2.6e-06
                  *->nGnTPLHlAarygnvevvklLLehGAdvnartk<-*
                    +G+T+LH+A + n+ +v 1L+e+GAdv a+
              201
                    KGQTALHIAIERRNMALVTLLVENGADVQAAAH
     hVR-1
ank: domain 2 of 3, from 248 to 283: score 13.2, E = 2
                  *->nGnTPLHlAarygnvevvklLLe...hGAdvnartk<-*
                    G PL laa ++++ +vk+LL+++ + Ad+ ar+
FGELPLSLAACTNQLGIVKFLLQnswQTADISARDS
    hVR-1
              248
                                                          283
ank: domain 3 of 3, from 333 to 361: score 3.4, E = 26
                  *->nGnTPLHlAarygnvevvklLLehGAdvnartk<-*
                    +G TPL 1Aa +g++ v ++ L+
                    KGMTPLALAAGTGKIGVLAYILQ----REIQEP
     hVR-1
                                                       361
```

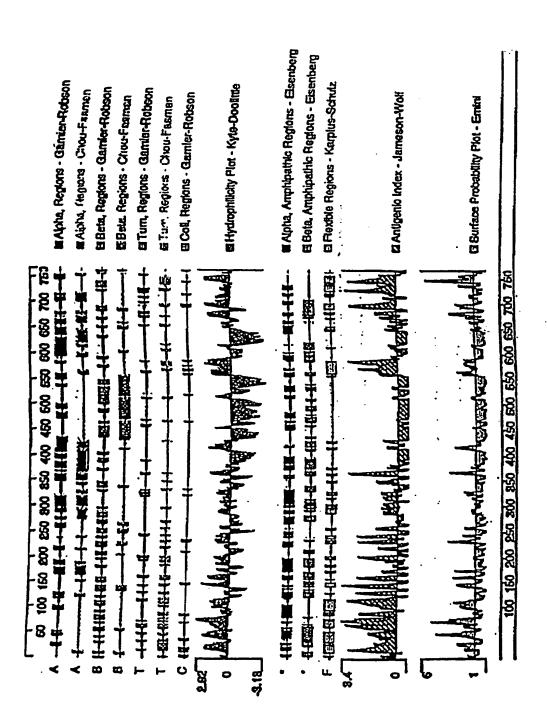


Figure 14

Protein Family / Domain Matches, HMMer version 2

```
Searching for complete domains
humpfam - search a single seq against HM database
HERER 2.1.1 (Dec 1998)
Copyright (C) 1992-1998 Washington University School of Ledicine
RMMER is freely distributed under the CNU General Public License (GPL).
                                                                                                  ---,-,-----
                                                                           /prod/ddm/sequal/PFAM/pfam3.3/Pfam
mer file:
                                                                          /tmp/orfenal.579.aa
Sequence file:
   Query: Plh2iell
Scores for sequence family classification (score includes all domains):
                                                                                                                                                                                        E-value N
                         Description
Model
                                                                                                                                                                                              4e-12
                                                                                                                                                                  53.7
                           PF00023 Ank repeat
 persed for domains:
                                                                                                                                            score E-value
                                                                                           home-f home-t
                          Domain seq-f seq-t
 Kodel
                                                                                                                                                                1.7e-07
                                                                                                                                               38.3
                                                                                                                      33 (i
                                                                        194 ..
                                 1/3
                                                       162
 ank
                                                                                                                                                 6.4
                                                                                                                      33 []
                                                       208
                                                                         243 ..
                                  2/3
 enk
                                                                                                                                                                           2.1
                                                                                                                                                  8.8
                                                                                                                      33 LI
                                                                         328 ..
                                                       293
                                 3/3
 ænk
 Alignments of top-scoring domains:

and domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

ends: domain 1 of 3, from 162 to 194: score 38.3, 8 = 1.7e-

                                                                 4G+4+EE+A 44 44+4Vkline;GA+V401
                                                                RCHBALHTATEKRSLQCVELLVERGAMVHARAC
                                                                                                                                                                         194
                                             162
              P1h21c11
  mike domain 2 of 3. from 208 to 243; soore 6.4. 5 - 4.3
                                                         *->nGotPlatlaryguvovvklide..: hgldvnartke-*
                                                                    G PL IRE + +++TV +ILO++++ A+ E++
                                                                  FGELFLELAACTEOMOVUSYGLESOPHOPASIOATDS
              Flh2le11
   emits domain 3 of 3. from 293 to 328; score 8.8, E=2.1
                                                           ->ngnrpinilacrypuvevvillie...hgadonart <<-*
+ 4TPL last-g+tet + L+t+ G + +r
                                                                  QUIAPLETANKEOKLETERHTLOPACEGLERILSRK?
                                                                                                                                                                                    328
                                              293
               F1h21e11
```

34/35

LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQATDSQGNTVLHALVM ISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKLAAKEGKIEIFRHIL **OREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAFHCKSPHRHRMVVLE GHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALLTVVSQVLCFLAIEW** YLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKKAISVLEMENGYWWCRKKQRAGVML RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL MKAVLNLKDGVNACI LPLLQI DRDSGNPQPLVNAQCTDDYYRGHSALHIA I EKRSLQCVK PLNKLLQAKWDLLI PKFFLNFLCNLI YMFI FTAVAYHQPTLKKQAAPHLKAEVGNSMLLT TVGTKPDGSPDERWCFRVEEVNWASWEQTLPTLCEDPSGAGVPRTLENPVLASPPKEDED MTSPSSSPVFRLETLDGGQEDĞSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLNY >hVR2.altFL (full-length predicted) **SASEENYVPVQLLQSN**

Figure 16

35/35

CLUSTAL W (1.74) multiple sequence alignment

humanVR2 hVR2.altFL	MTSPSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLN) MTSPSSSPVFRLETLDGGQEDGSEADRGKLDFGSGLPPMESQFQGEDRKFAPQIRVNLN)
humanVR2 hVR2.altFL	RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL RKGTGASQPDPNRFDRDRLFNAVSRGVPEDLAGLPEYLSKTSKYLTDSEYTEGSTGKTCL
humanVR2 hVR2.altFL	MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK MKAVLNLKDGVNACILPLLQIDRDSGNPQPLVNAQCTDDYYRGHSALHIAIEKRSLQCVK
humanVR2 hVR2.altFL	LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA LLVENGANVHARACGRFFQKGQGTCFYFGELPLSLAACTKQWDVVSYLLENPHQPASLQA
humanVR2 hVR2.altFL	TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL TDSQGNTVLHALVMISDNSAENIALVTSMYDGLLQAGARLCPTVQLEDIRNLQDLTPLKL
humanVR2 hVR2.altFL	AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF AAKEGKIEIFRHILQREFSGLSHLSRKFTEWCYGPVRVSLYDLASVDSCEENSVLEIIAF
humanVR2 hVR2.altFL	HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA HCKSPHRHRMVVLEPLNKLLQAKWDLLIPKFFLNFLCNLIYMFIFTAVAYHQPTLKKQAA
humanVR2 hVR2.altFL	PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL PHLKAEVGNSMLLTGHILILLGGIYLLVGQLWYFWRRHVFIWISFIDSYFEILFLFQALL
humanVR2 hVR2.altFL	TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQKVILRDLLRFLL TVVSQVLCFLAIEWYLPLLVSALVLGWLNLLYYTRGFQHTGIYSVMIQK
humanVR2 hVR2.altFL	IYLVFLFGFAVALVSLSQEAWRPEAPTGPNATESVQPMEGQEDEGNGAQYRGILEASLEL
humanVR2 hVR2.altFL	FKFTIGMGELAFQEQLHFRGMVLLLLLAYVLLTYILLLNMLIALMSETVNSVATDSWSIW
humanVR2 hVR2.altFL	KLQKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPTKAISVLEMENGYWWCRKKQRAGVMLTVGTKPDGSPDERWCFRVEEVNWASWEQTLPT
humanVR2 hVR2.altFL	LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN LCEDPSGAGVPRTLENPVLASPPKEDEDGASEENYVPVQLLQSN

FIGURE 17

- 1 -

SEQUENCE LISTING

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tatgaccatg attacgccaa gctctaatac gactcactat agggaaagct ggtacgcctg 600

cag	gtac	cgg	tccg	gaat	tc c	cggg	tcga	c cc	acgo	gtcc	gaa	aaca	cac	ctct	ctgctg	660
tgg	gaag	act	gtgc	aatg	gc a	cago	cgca	g ag	cttg	gttt	ggg	aggt	tga	agtg	ctctgg	720
gga	gaat	tcg	taga	tcat	cc t	caga	aaag	c ct	tgcc	ctgg	tgt	tcta	cca	gaaa	aacgtc	780
tcc	caat	cac	ccag	aaaa	gc t	gtcc	acag	t ag	tccc	ccct	tat	ccac	ggg	tgtc	actttc	840
cat	gggt	tca	gtta	tttg	cg g	tcaa	ccac	g gt	ctgc	caat	att	aaat	gga	aaat	tcttca	900
aac	agtt	ccc	aagt	tttc	cc t	tgtg	catt	g tt	ctga	gcag	tgt	gatg	aag	agto	tctgcc	960
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					gag Glu											1512
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WO 00/29577

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	agc Ser															2376
	ggc Gly															2424
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	tgt Cys															2568
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	aac Asn											tgat	ggc	cca		2662
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Thr	Asp	Ser	Gln	Gly 245	Asn	Thr	Val	Leu	His 250	Ala	Leu	Val	Met	Ile 255	Ser
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Tyr	Met	Phe	Ile	Phe 405	Thr	Ala	Val	Ala	Туг 410		Glr	Pro	Thr	Lev 415	_
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Gly	Gln 450	Leu	Trp	Tyr	Phe	Trp 455	Arg	Arg	His	Val	Phe 460		Trp	Ile	Ser
Phe 465	Ile	Asp	Ser	Tyr	Phe 470	Glu	Ile	Leu	Phe	Leu 475		Gln	Ala	Leu	Leu 480
Thr	Val	Val	Ser	Gln 485	Val	Leu	Cys	Phe	Leu 490		Ile	Glu	Trp	Tyr 495	
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Tyr	Thr	Arg 515	Gly	Phe	Gln	His	Thr 520	Gly	Ile	Tyr	Ser	Val 525	Met	Ile	Gln
Lys	Val 530	Ile	Leu	Arg	Asp	Leu 535	Leu	Arg	Phe	Leu	Leu 540	Ile	Tyr	Leu	Val
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Asn	Gly	Tyr 675	Trp	Trp	Cys	Arg	Lys 680	Lys	Gln	Arg	Ala	Gly 685	Val	Met	Leu

Thr Val Gly Thr Lys Pro Asp Gly Ser Pro Asp Glu Arg Trp Cys Phe 690 695 700

Arg Val Glu Glu Val Asn Trp Ala Ser Trp Glu Gln Thr Leu Pro Thr 705 710 715 720

Leu Cys Glu Asp Pro Ser Gly Ala Gly Val Pro Arg Thr Leu Glu Asn 725 730 735

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Lys Phe Ala Pro Gln Ile Arg Val Asn Leu Asn Tyr Arg Lys Gly Thr
50 55 60

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Tyr Leu Ser Lys Thr Ser Lys Tyr Leu Thr Asp Ser Glu Tyr Thr Glu

100 105 110

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ccc Pro	atg Met	gag Glu	gga Gly	cag Gln	gag Glu	gac Asp	gag Glu	ggc Gly	aac Asn	ggg ggg	gcc Ala	cag Gln	tac Tyr	agg Arg	ggt Gly	1776

- 22 **-**

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aag Lys	gag Glu	gat Asp	Glu	gat Asp 420	ggt Gly	gcc Ala	tct Ser	Glu	gaa Glu 425	aac Asn	tat Tyr	gtg Val	Pro	gtc Val 430	cag Gln	1295
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- 25 -

Ser Gln Gly Asn Thr Val Leu His Ala Leu Val Met Ile Ser Asp Asn 50 55 60

Ser Ala Glu Asn Ile Ala Leu Val Thr Ser Met Tyr Asp Gly Leu Leu 65 70 75 80

Gln Ala Gly Ala Arg Leu Cys Pro Thr Val Gln Leu Glu Asp Ile Arg 85 90 95

Asn Leu Gln Asp Leu Thr Pro Leu Lys Leu Ala Ala Lys Glu Gly Lys 100 105 110

Ile Glu Ile Phe Arg His Ile Leu Gln Arg Glu Phe Ser Gly Leu Ser

His Leu Ser Arg Lys Phe Thr Glu Trp Cys Tyr Gly Pro Val Arg Val 130 135 140

Ser Leu Tyr Asp Leu Ala Ser Val Asp Ser Cys Glu Glu Asn Ser Val 145 150 155 160

Leu Glu Ile Ile Ala Phe His Cys Lys Ser Pro His Arg His Arg Met 165 170 175

Val Val Leu Glu Pro Leu Asn Lys Leu Leu Gln Ala Lys Trp Asp Leu 180 185 190

Leu Ile Pro Lys Phe Phe Leu Asn Phe Leu Cys Asn Leu Ile Tyr Met 195 200 205

Phe Ile Phe Thr Ala Val Ala Tyr His Gln Pro Thr Leu Lys Lys Gln 210 215 220

Ala Ala Pro His Leu Lys Ala Glu Val Gly Asn Ser Met Leu Leu Thr 225 230 235 240

Gly His Ile Leu Ile Leu Gly Gly Ile Tyr Leu Leu Val Gly Gln 245 250 255

Leu Trp Tyr Phe Trp Arg Arg His Val Phe Ile Trp Ile Ser Phe Ile

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Leu 305	Val	Ser	Ala	Leu	Val 310	Leu	Gly	Trp	Leu	Asn 315	Leu	Leu	Tyr	Tyr	Thr 320	
Arg	Gly	Phe	Gln	His 325	Thr	Gly	Ile	Tyr	Ser 330	Val	Met	Ile	Gln	Lys 335	Lys	
Ala	Ile	Ser	Val 340	Leu	Glu	Met	Glu	Asn 345	Gly	Tyr	Trp	Trp	Cys 350	Arg	Lys	
Lys	Gln	Arg 355	Ala	Gly	Val	Met	Leu 360	Thr	Val	Gly	Thr	Lys 365	Pro	Asp	Gly	
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Ser 385	Trp	Glu	Gln	Thr	Leu 390	Pro	Thr	Leu	Cys	Glu 395	Asp	Pro	Ser	Gly	Ala 400	
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	Ala					Leu					Туз				c ctc Leu 80	240
					Leu					Glr					cgc Arg	288
				Leu					Leu					Gly	aag Lys	336
								Gln							agc Ser	384
			_	_					-				_		gtg Val	432
	ctg Leu														gtg Val 160	480
	gag Glu															528
_	gtt Val	_						-		-				_	-	576
	atc Ile											_			_	624
	atc Ile 210											Leu				672
	gcc Ala															720
	cac His		Leu					Gly								768
	tgg Trp						His									816
	agc Ser															864

- 28 -

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gtg t Val S										912
ctt g Leu V 305										960
cgt g Arg G										1008
gcc a Ala I										1056
aag c Lys G										1104
agc c Ser P 3										1152
tca t Ser T 385										1200
ggt g Gly V										1248
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			Asp					Thr					Leu		atg Met	145	
												Ile			tac Tyr	193	
								cgc Arg								241	
								ctc Leu								289	
								agg Arg 105								337	
								cga Arg								385	
								gac Asp								433	
								atc Ile								481	
								gaa Glu								529	
								aga Arg 185								577	
	_	-		_				acc Thr	_	-	_			_		625	
tcc	ctg	gat	cag	сса	gcc	atc	ccc	tca	tca	aaa	gcg	act	ttt	ggg	gaa	673	

Ser	Leu 210		Glr	n Pro	Ala	11e 215		Sei	Sei	Lys	s Ala 220		r Phe	e Gl	y Glu	
tcc Ser 225	Met	ctg Leu	ctg Leu	ı ctç ı Lev	ggc Gly 230	, His	att Ile	cto Leu	g ato 1 Il∈	cto Lev 235	ı Let	ggg Gly	g gg1 / Gly	t at	t tac = Tyr 240	721
ctc Leu	tta Leu	ctg Leu	ggc	Gln 245	Leu	tgg Trp	tac Tyr	ttt Phe	tgg Trp 250	Arg	g egg g Arg	g cgo g Aro	cto Lei	y tti 2 Phe 255	atc e Ile	769
				Met					Glu					ı Lei	cag Gln	817
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tgg Trp	tac Tyr 290	cta Leu	ccc Pro	ctg Leu	cta Leu	gtg Val 295	tta Leu	tcc Ser	cta Leu	gtg Val	ctg Leu 300	Gly	tgg Trp	ctg Leu	aac Asn	913
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atg Met	Leu	att Ile 435	gct Ala	ctc Leu	atg Met	Ser	gaa Glu 440	act Thr	gtc Val	aac Asn	cac His	gtt Val 445	gct Ala	gac Asp	aac Asn	1345

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gag Glu 465	aat Asn	ggt Gly	tac Tyr	tgg Trp	tgg Trp 470	tgc Cys	cgg Arg	agg Arg	aag Lys	aaa Lys 475	cat His	cgt Arg	gaa Glu	ggg Gly	agg Arg 480	1441
ctg Leu	ctg Leu	aaa Lys	gtc Val	ggc Gly 485	acc Thr	agg Arg	ggg Gly	gat Asp	ggt Gly 490	acc Thr	cct Pro	gat Asp	gag Glu	cgc Arg 495	tgg Trp	1489
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ccc Pro	acc Thr	tta Leu 515	tct Ser	gag Glu	gat Asp	cca Pro	tca Ser 520	Gl y ggg	cca Pro	ggc Gly	atc Ile	act Thr 525	ggt Gly	aat Asn	aaa Lys	1585
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cat His 545	ctg Leu	ccc Pro	ctt Leu	cag Gln	gtc Val 550	ctc Leu	cag Gln	tcc Ser	ccc Pro	tgat	ggc	cca q	gatgo	cagca	ag	1683
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Glu	Ala	Thr 35	Asp	Ser	Leu	Gly	Asn 40	Thr	Val	Leu	His	Ala 45	Leu	Val	Met	
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Glu	Glu	Ile	Ser	Asn	His	Gln	Gly	Leu	Thr	Pro	Leu	Lys	Leu	Ala	Ala	

90

85

- 32 -

Lys	Glu	Gly	Lys 100		e Glu	Ile	Phe	105		: Ile	e Leu	í Gln	Arg 110		ı Phe
Ser	Gly	Pro 115	_	Gln	Pro	Leu	Ser 120	_	Lys	Phe	Thr	Glu 125	_	Cys	ту
Gly	Pro 130		Arg	Val	Ser	Leu 135		Asp	Leu	Ser	Ser 140		. Asp	Ser	Trp
Glu 145	Lys	Asn	Ser	Val	Leu 150		Ile	Ile	Ala	Phe 155	His	Cys	Lys	Ser	Pro 160
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Tyr	Leu	Val 195	Tyr	Met	Phe	Ile	Phe 200	Thr	Val	Val	Ala	Tyr 205	His	Gln	Pro
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Gly	Glu	Leu	Ala	Phe 405	Gln	Glu	Gln	Leu	Arg 410	Phe	Arg	Gly	Val	Val 415	Leu

- 33 -

Leu Leu Leu Ala Tyr Val Leu Leu Thr Tyr Val Leu Leu Asn Met Leu Ile Ala Leu Met Ser Glu Thr Val Asn His Val Ala Asp Asn 440 Ser Trp Ser Ile Trp Lys Leu Gln Lys Ala Ile Ser Val Leu Glu Met Glu Asn Gly Tyr Trp Trp Cys Arg Arg Lys Lys His Arg Glu Gly Arg 470 475 465 Leu Leu Lys Val Gly Thr Arg Gly Asp Gly Thr Pro Asp Glu Arg Trp 490 Cys Phe Arg Val Glu Glu Val Asn Trp Ala Ala Trp Glu Lys Thr Leu 500 Pro Thr Leu Ser Glu Asp Pro Ser Gly Pro Gly Ile Thr Gly Asn Lys 520 Lys Asn Pro Thr Ser Lys Pro Gly Lys Asn Ser Ala Ser Glu Glu Asp 535 530 His Leu Pro Leu Gln Val Leu Gln Ser Pro 550 545 <210> 12 <211> 1662 <212> DNA <213> Rattus sp. <220> <221> CDS <222> (1)..(1662) <400> 12 tcg acc cac gcg tcc gct ctt tct ctg gct gcg tgc acc aag cag tgg Ser Thr His Ala Ser Ala Leu Ser Leu Ala Ala Cys Thr Lys Gln Trp 5 gat gtg gtg acc tac ctc ctg gag aac cca cac cag ccg gcc agc ctg Asp Val Val Thr Tyr Leu Leu Glu Asn Pro His Gln Pro Ala Ser Leu 20 gag gcc acc gac tcc ctg ggc aac aca gtc ctg cat gct ctg gta atg Glu Ala Thr Asp Ser Leu Gly Asn Thr Val Leu His Ala Leu Val Met 4.5 40 35 att qca gat aac tcg cct gag aac agt gcc ctg gtg atc cac atg tac Ile Ala Asp Asn Ser Pro Glu Asn Ser Ala Leu Val Ile His Met Tyr 50 55 60 gac ggg ctt cta caa atg ggg gcg cgc ctc tgc ccc act gtg cag ctt 240 Asp Gly Leu Leu Gln Met Gly Ala Arg Leu Cys Pro Thr Val Gln Leu 70 75

gaç Glu	ggaa Glu	ato Ile	tco Ser	aac Ası 85	n His	caa Glr	a ggo	c cto / Lev	aca Thi	r Pro	c cto Lev	g aaa 1 Lys	a cta s Lei	a gco Ala 91	c gcc a Ala 5	288
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			Tyr					Arg					Trp		tac Tyr	384
		Val					Tyr					Val			tgg Trp	432
											His				ccg Pro 160	480
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tac Tyr	ttg Leu	gtc Val 195	tac Tyr	atg Met	ttc Phe	atc Ile	ttc Phe 200	acc Thr	gtc Val	gtt Val	gcc Ala	tac Tyr 205	cac His	cag Gln	cct Pro	624
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			ccc Pro													1152
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			gct Ala													1248
			ttg Leu 420													1296
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			gtc Val													1488
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- 37 -

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Ala	Суѕ	Gly 195	Arg	Phe	Phe	Gln	Lys 200	Gly	Gln	Gly	Thr	Cys 205	Phe	Tyr	Phe
Gly	Glu 210	Leu	Pro	Leu	Ser	Leu 215	Ala	Ala	Cys	Thr	Lys 220	Gln	Trp	Asp	Val
Val 225	Ser	Tyr	Leu	Leu	Glu 230	Asn	Pro	His	Gln	Pro 235	Ala	Ser	Leu	Gln	Ala 240
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Ser	Val	Leu 355	Glu	Ile	Ile	Ala	Phe 360		Cys	Lys	Ser	Pro 365		Arg	Hi
Arg	Met 370	Val	Val	Leu	Glu	Pro 375	Leu	Asn	Lys	Leu	Leu 380	Gln	Ala	Lys	Tr
Asp 385	Leu	Leu	Ile	Pro	Lys 390	Phe	Phe	Leu	Asn	Phe 395	Leu	Cys	Asn	Leu	I1 40
Tyr	Met	Phe	Ile	Phe 405	Thr	Ala	Val	Ala	Tyr 410	His	Gln	Pro	Thr	Leu 415	Ly
Lys	Gln	Ala	Ala 420	Pro	His	Leu	Lys	Ala 425		Val	Gly	Asn	Ser 430	Met	Le
Leu	Thr	Gly 435	His	Ile	Leu	Ile	Leu 440	Leu	Gly	Gly	Ile	Tyr 445	Leu	Leu	Va.
Gly	Gln 450	Leu	Trp	Туr	Phe	Trp 455	Arg	Arg	His	Val	Phe 460	Ile	Trp	Ile	Se:
Phe 465	Ile	Asp	Ser	Tyr	Phe 470	Glu	Ile	Leu	Phe	Leu 475	Phe	Gln	Ala	Leu	Le:
Thr	Val	Val	Ser	Gln 485	Val	Leu	Cys	Phe	Leu 490	Ala	Ile	Glu	Trp	Tyr 495	Leu
Pro	Leu	Leu	Val 500	Ser	Ala	Leu	Val	Leu 505	Gly	Trp	Leu	Asn	Leu 510	Leu	Туі
Tyr	Thr	Arg 515	Gly	Phe	Gln	His	Thr 520	Gly	Ile	Tyr	Ser	Val 525	Met	Ile	Glr
Lys	Lys 530	Ala	Ile	Ser	Val	Leu 535	Glu	Met	Glu	Asn	Gly 540	Tyr	Trp	Trp	Cys
Arg 545	Lys	Lys	Gln	Arg	Ala 550	Gly	Val	Met	Leu	Thr 555	Val	Gly	Thr	Lys	Pro 560
Asp	Gly	Ser	Pro	Asp 565	Glu	Arg	Trp	Cys	Phe 570	Arg	Val	Glu	Glu	Val 575	Asr
Trp	Ala	Ser	Trp 580	Glu	Gln	Thr	Leu	Pro 585	Thr	Leu	Cys	Glu	Asp 590	Pro	Ser
Gly	Ala	Gly 595	Val	Pro	Arg	Thr	Leu 600	Glu	Asn	Pro	Val	Leu 605	Ala	Ser	Pro
Pro	Lys 610	Glu	Asp	Glu	Asp	Gly 615	Ala	Ser	Glu	Glu	Asn 620	Tyr	Val	Pro	Val
Gln 625	Leu	Leu	Gln	Ser	Asn 630										

INTERNATIONAL SEARCH REPORT

intern net Application No PCT/US 99/26701

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/12 C12N5/10 C07K14/705 C07K16/28 GO1N33/53 C12Q1/68 A61P25/00 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 7 C12N C07K G01N C12Q A61P Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Category ' Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X CATERINA M J ET AL: "THE CAPSAICIN 1-26 RECEPTOR: A HEAT-ACTIVATED ION CHANNEL IN THE PAIN PATHWAY" NATURE, GB, MACMILLAN JOURNALS LTD. LONDON, vol. 389, 23 October 1997 (1997-10-23), pages 816-824, XP002075020 ISSN: 0028-0836 cited in the application abstract figure 5 page 821, left-hand column, paragraph 2 P,X WO 99 09140 A (BRAKE ANTHONY ; JULIUS DAVID 1-26 J (US); UNIV CALIFORNIA (US); CATERINA) 25 February 1999 (1999-02-25) abstract 100% identity in 1717 BP overlap between SEQ ID NO 3 of W09909140 and SEQ ID NO 10 Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents : T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance Invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention filing date cannot be considered novel or cannot be considered to "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another Involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention citation or other special reason (as specified) cannot be considered to involve an inventive step when the document is combined with one or more other such docu-"O" document referring to an oral disclosure, use, exhibition or other means ents, such combination being obvious to a person skilled *P* document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 28 April 2000 10/05/2000 Name and mailing address of the ISA **Authorized officer** European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Lejeune, R Fax: (+31-70) 340-3016

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INTERNATIONAL SEARCH REPORT

Intern Aal Application No PCT/US 99/26701

		PC1/US 99/26/01
	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	WO 99 37675 A (BRAKE ANTHONY J ;JULIUS DAVID J (US); UNIV CALIFORNIA (US); CATERI) 29 July 1999 (1999-07-29) abstract 99.7% identity in 2533 BP overlap between SEQ ID NO 33 of WO9937675 and SEQ ID NO 1 100% identity in 2380 BP overlap between SEQ ID NO 35 of WO9937675 and SEQ ID NO 4 100% identity in 1717 BP overlap between SEQ ID NO 3 of WO9937675 and SEQ ID NO 10	1-26
P.X	WO 99 37765 A (SMITHKLINE BEECHAM PLC) 29 July 1999 (1999-07-29) abstract 99.9% identity in 2351 BP overlap between SEQ ID NO 1 and 5 of WO9937765 and SEQ ID NO 4	1–26
P,X	EP 0 943 683 A (SMITHKLINE BEECHAM PLC) 22 September 1999 (1999-09-22) abstract 99.6% identity in 2818 BP overlap between SEQ ID NO 1 of EP943683 and SEQ ID NO 1	1-26
P,X	EP 0 953 638 A (SYNTHELABO) 3 November 1999 (1999-11-03) abstract 99.8% identity in 2786 BP overlap between SEQ ID NO 1 of EP953638 and SEQ ID NO 4	1-26
Ρ,Χ	CATERINA ET AL: "A capsaicin-receptor homologue with a high threshold for noxious heat" NATURE, GB, MACMILLAN JOURNALS LTD. LONDON, vol. 398, no. 398, 1 April 1999 (1999-04-01), pages 436-441-441, XP002105951 ISSN: 0028-0836 the whole document 99.8% identity in 2401 BP overlap between the sequence of hVRL and SEQ ID NO 4 100% identity in 1752 BP overlap between the sequence of rVRL and SEQ ID NO 10	1-26
X	WO 98 39448 A (HUMAN GENOME SCI INC) 11 September 1998 (1998-09-11) abstract page 112 page 163 99.3% identity in 1947 BP overlap between SEQ ID NO 191 of WO9839448 and SEQ ID NO 4 98.1% identity in 1709 BP overlap between SEQ ID NO 307 of WO9839448 and SEQ ID NO 4	1-11

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/26701

BOX I OD	servations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Internati	ional Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
bec	ims Nos.: ause they relate to subject matter not required to be searched by this Authority, namely: mark: Although claims 24–26 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
bec	ims Nos.: ause they relate to parts of the International Application that do not comply with the prescribed requirements to such extent that no meaningful International Search can be carried out, specifically:
3. Clai	ms Nos.: ause they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Ob	servations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This Internation	onal Searching Authority found multiple inventions in this international application, as follows:
1. As a	all required additional search fees were timely paid by the applicant, this International Search Report covers all rehable claims.
2. As a of an	all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment ny additional fee.
3. As o	only some of the required additional search fees were timely paid by the applicant, this International Search Report ers only those claims for which fees were paid, specifically claims Nos.:
4. No restr	required additional search fees were timely paid by the applicant. Consequently, this International Search Report is incred to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on P	The additional search fees were accompanied by the applicant's protest. N protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT Information on patent family members

Intern tal Application No PCT/US 99/26701

c	Patent document ited in search report	t	Publication date		Patent family member(s)		Publication dat
h	O 9909140	Α	25-02-1999	AU AU	9115698 2466799		08-03-1999
				WO	9937675	A A	09-08-1999 29-07-1999
- المار	0 9937675	A	29-07-1999	AU		Α	09-08-1999
,				AU Wo		A	08-03-1999
/ _				WU 	9909140	A	25-02-1999
W	10 9937765	A	29-07-1999	NONE	•		
E	P 0943683	Α	22-09-1999	JP	11279196	A	12-10-1999
E	P 0953638	A	03-11-1999	AU	2932799	A	27-09-1999
				WO	9946377	A	16-09-1999
W	0 9839448	A		NONE	, ,		

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